50th Annual Meeting and Golden Jubilee
Celebration

of the



Society for Invertebrate Pathology



13 – 17 August 2017 UC San Diego La Jolla, California

MEETING AT A GLANCE 2017 SIP Annual Meeting and Golden Jubilee Celebration

NOTE: The abstracts included in this book should not publications and should not be cited in print without the author's permission.

POSTERS should be put up after 8:00 am on Monday in the West Ballroom.

They must be removed by noon on Thursday or they will be discarded.

STU indicates papers being judged for graduate student presentation awards.

Presenting authors for oral talks and posters are indicated in bold/italics type.

8:00 - 10:00

Bacteria Division business meeting

SUNDAY – 13 August

	SUNDAY – 13 August	8.00 - 10.00	Dacteria Division business meeting		
8:45 - 5:00	:45 – 5:00 Council Meeting		(Village Tower West, 15th floor, Rm. 15B)		
	(Village Tower West, Room 15C)		Diseases of Beneficial Invertebrates		
2:00 - 6:00	Registration (Village Conference		Division business meeting		
	Center Check-in Desk)		(Village West, Bldg. 2, Rm. 2C)		
6:00 - 8:00	Mixer / Welcome Reception		Microbial Control Division		
(Institute of the Americas, Plaza)		business meeting			
	(mstitute of the Americus, Fluzu)		(Village Tower West, 15th fl., Rm. 15C)		
	MACNIDAY AA Ayyyya		Nematodes Division workshop		
	MONDAY – 14 August		(Village West Bldg. 2, Rms. 2ª/B)		
7:30 – 5:00	Registration (Price Center)				
8:00 – 12:30	Opening Ceremony (Theater)		TUESDAY – 15 August		
	Awards Presentations	6 30 7 30			
	Golden Jubilee	6:30 – 7:30	5K Run/Walk		
	Founders Symposium	8:00 – 10:00	Virus/DBI X-Division		
10:00 - 10:30	Coffee break		Symposium: Honoring		
	Golden Jubilee		Just Vlak (Theater)		
	Founders Symposium (continued)		Bacteria Division #2 (The Forum)		
12:30 - 1:30	Lunch	Fungus Division Symposiu			
1:30 - 3:30	Bacteria/Microbial Control		Insect defense against		
	X-Division Symposium:		fungal pathogens:		
	Challenges for Biocontrol		mechanisms, variations		
	in Latin America (Theater)		and efficiency (Roosevelt Room)		
	Virus Division #1 (The Forum)		Microbial Control Division #1		
	Microsporidia Division #1		(Marshall Room)		
	(Roosevelt Room)	10:00 - 10:30	Coffee break		
	Nematodes Division #1	10:30 - 12:30	Virus Division #2 (Theater)		
	(Marshall Room)		Nematodes Division Symposium:		
3:30 – 4:00	Refreshment break		Nematode Omics (The Forum)		
3.30 4.00	Virus Division Symposium:		Fungi Division #2 (Roosevelt Room)		
	Insect virology: Historical		DBI #2 (Marshall Room)		
	achievements and recent	12:30 - 5:00	EXCURSIONS		
		6:00 - 10:00	Barbeque (Birch Aquarium)		
	, , , , , , , , , , , , , , , , , , , ,		. , , , , , , , , , , , , , , , , , , ,		
	Bacteria Division #1 (The Forum)				
	Fungi Division #1 (Roosevelt Room)				
	DBI #1 (Marshall Room)				



50th Annual Meeting and Golden Jubilee Celebration of the Society for Invertebrate Pathology

University of California San Diego La Jolla, California 13–17 August 2017

Table of Contents

(Monday, Tuesday)	Inside front cover
SIP Officers	2
SIP Committees	2
SIP Division Officers	3
2017 Meeting Organizing Committee	3
Fifty Years of SIP Meetings	4
Program of the Meeting (including abstracts)	7
Short Program (center of book, on colored paper)	SP-1
Index of Authors	89
Addendum	96
Meeting At A Glance (Wednesday, Thursday)	Inside back cover

SIP OFFICERS

President Johannes Jehle Federal Research Center for Cultivated Plants

Julius Kuehn Institute, Institute for Biological Control Heinrichstrasse 243, Darmstadt 64287, Germany

Phone: +49 (6151)-407220 / Email: Johannes.jehle@julius-kuehn.de

Vice President Zhihong (Rose) Hu Wuhan Institute of Virology, Chinese Academy of Sciences

Wuhan 430071, PR China

Phone: +86 (27) 87197180 / Email: huzh@wh.iov.cn

Secretary Juan Luis Jurat-Fuentes Dept. of Entomology and Plant Pathology, The University of Tennessee

370 Plant Biotechnology Bldg, Knoxville, TN 37996, USA Phone: +1 (865) 974-5931 / Email: jurat@utk.edu

Treasurer Stefan Jaronski PO Box 232, Sidney, MT 59270, USA

Phone: +1 (406) 433-9486 / Email: thebugdoc01@gmail.com

Past President Peter Krell University of Guelph, Department of Molecular and Cellular Biology,

50 Stone Rd East, Guelph, ON N162W1, Canada Phone: +1 (519) 824-4120, ext. 53368 / Email:

TRUSTEES: Albrecht Kopperhofer Rutgers University, Entomology Department

Blake Hall, 93 Lipman Drive, New Brunswick, NJ 08901-8524, USA Phone: +1 848-932-9802 / koppenhofer@aesop.rutgers.edu

Monique van Oers Wageningen University, Laboratory of Virology

Droevendaalsesteeg 1, Wageningen, 6708 PB, The Netherlands Phone: 31-317-485082 / Email: monique.vanoers@wur.nl

Helen Hesketh Center for Ecology & Hydrology, Maclean Bldg.

Crowmarsh Fillord, Wallinngfors, OX10 8BB, UK Phone: +44 1491 692574 / Email: hhesketh@cef.ac.uk

Sean Moore Citrus Research International, PO Box 20285, Humewood, Port Elizabeth, 6013, South Africa

Phone: +27 41-5835524 / Email:

SIP COMMITTEES

Archivist Betty Davidson

Awards & Student Contest Monique van Oers (Chair), Patricia Stock (Vice-Chair), Andreas Line

Hyun-Woo Park, Kelly Bateman

Endowment & Financial Support. Roma Gwynn (Chair), Michael Brownbridge, Michael Dimock

James Harper, Dirk Ave, Stefan Jaronski (ex officio)

Founders Lecture James Becnel (Chair), Neil Crickmore, Mark Goettel

Golden Jubilee Peter Krell (Chair), Julie Hopper, Surendra Data, Betty Davidson

Johannes Jehle, Nina Jenkins, Kelli Hoover, Leellen Solter, Grant Stentiford

Monique van Oers

History Elizabeth Davidson (Chair), James Harper, Donald Roberts, Harry Kaya,

Fernando Vega, Juerg Huber, Mark Goettel, Just Vlak

Meetings Mark Goettel (Chair), Nina Jenkins, Elisabeth Herniou, Jörg Wennmann

Membership Surendra Dara (Chair) Peter Krell (Vice Chair) Ambassador Program,

Stefan Jaronski (ex officio)

Nominating Peter Krell (Chair), Jørgen Eilenberg, Leellen Solter, Mark Goettel, Madoka Nakai

Publications David Shapiro-Ilan (Chair), Selcuk Hazir, Albrecht Kopperhöfer, Bryony Bonning,

Johannes Jehle (ex officio), Jean-Louis Schwartz (Newsletter Editor; ex officio),

Cecilia Schmitt (website, ex officio), Lee Solter (JIP, ex officio)

Student Affairs Julie Hopper (Chair), Louise-Marie Roux (Co-Chair), Patricia Stock (Faculty Advisor),

Satomi Adegawa (Bacteria Div.), Katharina Saar and Carina Ehrich (Fungi Div.), Andreas Larem (Microbial Control Div.); Rousel Orozco (Nematodes Div.),

Andreas Earem (wherobial control biv.), Roaser 616266

SIP DIVISION OFFICERS:

Bacteria Kenneth Narva (Chair); Marianne Carey (Chair Elect); Sun Ming (Secretary/Treasurer);

Shuyuan Guo, Juan Ferré (Members as Large); Baltasar Escriche (Past Chair);

Satomi Adegawa (Student Representative)

Diseases of Helen Hesketh (Chair); Mark Freeman (Chair Elect); Kelly Bateman (Secretary/Treasurer);

Beneficial David Bass, Annette Bruun Jensen (Members at Large); Kelly Bateman (Past Chair);

Invertebrates Georgia Ward (Student Representative)

Fungi Nicolai Meyling (Chair); Stefan Jaronski (Chair Elect); Ann Hajek (Secretary/Treasurer); Dietrich Stephan,

Chad Keyser (Members at Large); Katharina Saar, Carina Ehrich (Student Representatives)

Microbial Control Dietrich Stephan (Chair); vacant (Chair Elect); Michael Brownbridge (Secretary/Treasurer);

Mary Barbercheck, vacant (Members at Large); Andreas Larem (Student Representative)

Microsporidia Jyliya Sokolova (Chair); George Kyei-Poku (Chair Elect); Julie Hopper (Secretary/Treasurer);

Naomi Fast, Bryony Williams (Members-at-Large); Susan Björnson (Past Chair);

Sarah Biganski (Student Representative)

Nematodes Glen Stevens (Chair): Raquel Campos-Herrera (Chair Elect); Patricia Stock (Secretary/Treasurer); Ivan

Hiltpold, Barton Slatko (Members-at-Large); David Shapiro-llan (Past Chair); Rousel Orozco (Student

Representative); Sylvia Libro (Facebook Representative)

Viruses Madoka Nakai (Chair); Elisabeth Herniou (Chair Elect); Vera Ros (Secretary/Treasurer);

Holly Popham, Jørg Wennmann (Members-at-Large); Martin Erlandson (Past Chair);

Yue Han, Carina Bannach (Student Representatives)

2017 MEETING ORGANIZING COMMITTEE

Chair Surendra Dara

Scientific Program Richard Humber (Chair), Louela Castrillo (Co-Chair), Ed Lewis,

Lerry Lacey (ex officio)

Program Book Richard Humber

Local Arrangements Kelli Hoover, Dennis Bideshi, Wendy Gelernter, Hyun-Woo Park,

Patricia Stock

Excursions Raffi Aroian

5K Race/Walk Nina Jenkins, Ken Narva

FIFTY YEARS OF SIP MEETINGS

1980 Seattle, Washington, USA 1968 Columbus, Ohio, USA Organizer: Albert Sparks Organizer: John D. Briggs 1981 Bozeman, Montana, USA 1969 **Burlington, Vermont, USA** Organizer: John Henry **Organizer Thomas Cheng** Excursion: Madison River float trip Excursion: Lake Champlain and Green Mountains 1982 Brighton, England, UK 1970 College Park, Maryland, USA 4th International Colloquium with American Institute of Biological Sciences Founder's Lecture Honoree/Speaker: Organizers: James Vaughn, Art Heimpel K.M. Smith / C.F. Rivers 1971 Montpellier, France Organizers: Denis Burges, Chris Payne 1st International Colloquium **Excursion: Brighton Pavilion** Organizer: Max Bergoin 1983 Ithaca, New York, USA Excursion: Pont de Garde, Camargue, San Christol Founder's Lecture Honoree/Speaker: 1972 Minneapolis, Minnesota, USA J. Mackin / A.H. Sparks Organizer: Marian Brooks Organizers: Donald Roberts, Robert Granados Excursion: Water quality laboratory 1984 Davis, California, USA 1973 Oxford, England, UK Founder's Lecture Honoree/Speaker: 2nd International Colloquium E.A. Steinhaus / D.W. Roberts Organizer: Thomas Tinsley Organizers: Harry Kaya, Donald Roberts Excursion: Warwick Castle; Stratford Excursions: City of Sacramento; Bodega Bay Amherst, Massachusetts, USA Marine Laboratoy with American Institute of Biological Sciences 1985 Sault Ste. Marie, Ontario, Canada 1974 Tempe, Arizona, USA Founder's Lecture Honoree/Speaker: with American Institute of Biological Sciences G.H. Bergold / T.A. Angus Organizers: Organizers: Elizabeth Davidson, Patrick Vail Terry Ennis, John Cunningham, Excursion: Wave pool, desert hike Sardar Sohi 1975 Corvallis, Oregon, USA Veldhoven, Netherlands 1986 Organizers: Mauro Martignoni, Michael Mix, 5th International Colloquium Chris Bayne Founder's Lecture Honoree/Speaker: E. Metschnikoff / J. Weiser Organizers: 1976 Kingston, Ontario, Canada Just Vlak, Rob Samson, Dick Peters 3rd International Colloquium Excursion: Philips EM facility; Heinekin Brewery Organizer: Tom Angus, Aaron Rosenfield, Peter Faulkner 1987 Gainesville, Florida, USA **Excursion: Fort Henry** Founder's Lecture Honoree/Speaker: L. Pasteur / C. Vago Organizers: Drion 1977 East Lansing, Michigan, USA Boucias, James Maruniak, with American Institute of Biological Sciences Al Undeen, Clayton McCoy, Organizers: John Bresnak, T. Rizki John Couch, Patrick Greany Excursion: Pont de Garde, Camargue, San Christol 1988 La Jolla, California, USA 1978 Prague, Czechoslovakia Founder's Lecture Honoree/Speaker: Organizer: Jaroslav Weiser

Excursion: Pont de Garde, Camargue, San Christol

Gainesville, Florida, USAOrganizer: John Harshbarger

1979

"State of the Art" / H. Whiteley

Organizer: Brian Federici

1989 College Park, Maryland, USA

Organizers: James Vaughn, Charles Reichelderfter,

Edward Daugherty

Founder's Lecture Honoree/Speaker:

R.R. Kudo / V. Sprague

1990 Adelaide, Australia

6th International Colloquium

Founder's Lecture Honoree/Speaker:

T.D.C. Grace / K. Maramorosch

Organizer: Dudley Pinnock Excursion: Winery tour

1991 Flagstaff, Arizona, USA

5th International Colloquium

Founder's Lecture Honoree/Speaker:

R.W. Glaser / G.O. Poinar Jr.

Organizer: Elizabeth Davidson

Excursion: Philips EM facility; Heinekin Brewery

1992 Heidelberg, Germany

Founder's Lecture Honoree/Speaker:

E. Müller-Kögler / A. Vey

Organizers: Jurg Huber, Gisbert Zimmermann,

Wolfgang Schnetter, Norbert Becker

Excursion: River boat tour

1993 Asheville, North Carolina, USA

Founder's Lecture Honoree/Speaker:

J.N. Couch / H.C. Whisler

Organizers: James Harper, Wayne Brooks

Excursion: Biltmore Estate

1994 Montpellier, France

7th International Colloquium

Founder's Lecture Honoree/Speaker:

C. Vago / L.K. Miller

Organizer: Max Bergoin Excursion: Aigues-Morte tour

1995 Ithaca, New York, USA

Founder's Lecture Honoree/Speaker:

H.T. Dulmage / D.J. Ellar

Organizers: John Vandenberg, Alan Wood Excursion: Taughannock Falls State Park

1996 Cordoba, Spain

Founder's Lecture Honoree/Speaker:

A. Bassi / D.W. Roberts

Organizers: Candido Santiago-Alvarez,

Enrique Vargas-Osuna

Excursion: Alcazar de los Reyes Cristianos

1997 Banff, Alberta, Canada

Founder's Lecture Honorees/Speaker:

T.A. Angus, A.M. Heimpel /

K. van Frankenhuyzen

Organizers: Mark Goettel

Excursions: Columbia Icefields and Banff Springs

Hotel; Nordic Center; Lake Louise

1998 Sapporo, Hokkaido, Japan

Founder's Lecture Honoree/Speaker:

K. Maramorosch / J. Mitsuhashi

Organizer: Toshihiko Iizuka

Excursions: Mt. Tarumae and Shikotsu Lake;

Noboribetsu Hotspring

1999 Irvine, California, USA

Founder's Lecture Honoree/Speaker:

Y. Tanada / J.K. Kaya

Organizers: Brian Federici

Excursions: Wine tasting; Mission San Juan

Capistrano

2000 Guanajuato, Mexico

Founder's Lecture Honoree/Speaker:

H.D. Burges / B.A. Federici

Organizers: Jorge Ibarra, Alejandra Bravo

Excursion: San Miguel de Allende, Teatro Juarez

2001 Noordwijkerhout Netherlands

(relocated from scheduled site in Israel)

Founder's Lecture Honoree/Speaker:

J. Weiser / W. Brooks

Organizers: Just Vlak, Meir Broza

Excursion: Van Gogh Museum, canal boat tour

2002 Foz de Iguassu, Brazil

8th International Colloquium

Founder's Lecture Honoree/Speaker:

H. de Barjac / P. Lüthy

Organizers: Flávio Moscardi, Sérgio Batista Alves

Excursion: Iguassu Falls

2003 Burlington, Vermont, USA

Founder's Lecture Honoree/Speaker:

L. Miller / R.R. Granados Organizers:

John Burand, Ann Hajek

Excursion: Lake Champlain boat cruise

2004 Helsinki, Finland

Founder's Lecture Honoree/Speaker:

H.G. Boman / K. Söderhall

Organizers: Heikki Hokkanen, Jørgen Eilenberg

Excursion: Marimekko factory

2005 Anchorage, Alaska, USA

Founder's Lecture Honoree/Speaker:

E. Canning / J. Becnel

Organizers: Kelli Hoover, Diana Cox-Foster,

Bryony Bonning

Excursion: Train ride to top of Mt. Alyeska

2006 Wuhan, Hubei, China

9th International Colloquium

Founder's Lecture Honoree/Speaker:

Gao S.Y. / J. Vlak

Organizers: Yu Ziniu, Sun Ming,

Hu (Rose) Zhihong

Excursion: Yellow Crane Tower; Three Gorges;

Xi'an

2007 Québec City, Québec, Canada

1st Internat. Forum on Entomopathogenic

Nematodes and Symbiotic Bacteria

Founder's Lecture Honoree/Speaker:

A. Sparks / F. Morado

Organizers: Conrad Cloutier, Jean Louis Schwartz,

Basil Arif, Roland Brousseau

Excursion: St. Lawrence river cruise

2008 Warwick, England, UK

Founder's Lecture Honoree/Speaker:

A. Paillot / J. Jehle

Organizers: David Chandler, Bryonny Bonning,

Doreen Winstanley

Excursion: Warwick Castle; Stratford-upon-Avon

2009 Park City, Utah, USA

Founder's Lecture Honoree/Speaker:

D.W. Roberts / R.J. St. Leger

Organizers: Donald Roberts, Rosalind James

Excursion: Olympic Park, ski jumpers

2010 Trabzon, Turkey

Founder's Lecture Honoree/Speaker:

M. Martignoni / B. Arif

Organizers: Zihni Demirbag, Basil Arif

Excursion: Sumela Monastery

2011 Halifax, Nova Scotia, Canada

Organizers: Susan Bjornson, Christina Noronha,

Michael Agbeti, Thomas Steele

Founder's Lecture Honoree/Speaker:

J.D. Briggs / E. Davidson

Excursion: Peggy's Cove; Citadel, Pier 21

2012 Buenos Aires, Argentina

1st International Congress on Invertebrate

Pathology and Microbial Control

Founder's Lecture Honorees/Speaker:

S.B. Alves, F. Moscardi*

/ I. Delalibera Jr

(* to be the Founder's Lecturer but died

shortly before this meeting)

Organizers: Alicia Sciocco-Cap,

Victor Romanowski, Juan Ferré

Excursion: Tigre-Delta river tour

2013 Pittsburgh, Pennsylvania, USA

Founder's Lecture Honoree/Speaker:

R. Granados / G. Blissard

Organizers: Nina Jenkins, Kelli Hoover

Excursion: Carnegie Museum; Three Rivers

boat tour

2014 Mainz, Germany

Founder's Lecture Honoree/Speaker:

A. Huger / T. Jackson

Organizers: Johannes Jehle

Excursion: Rhine River cruise, Schloss Rheinfels

2015 Vancouver, British Columbia, Canada

Founder's Lecture Honoree/Speaker:

P. Johnson / G. Stentiford

Organizers: Mark Goettel, Todd Kabaluk

Excursion: Cheakamus Center

2016 Tours, France

Founder's Lecture Honoree/Speaker:

D. Ellar / N. Crickmore

Organizers: Elisabeth Herniou, Christine

Nielsen-LeRoux, Jean-Michel Drezen

Excursion: Domaine de Cande

Registration:

Sunday 2– 6 pm, Village Conference Desk

(adjacent to check-in desk for housing)

Monday 7 am -5 pm, Price Center
Tuesday 7: am -1 pm, Price Center
Wednesday 8 am -5 pm, Price Center
Thursday, 8 am -12 noon, Price Center

Posters: Price Center, West Ballroom

Hang on Monday, after 8:30 am

Posters will be displayed throughout meeting

Remove on Thursday, by 12:00 noon

Speaker Ready Room, Slide Loading:

Sunday Adjacent to mixer site

All other days 8:00 am - 5:00 pm, Revelle Room,

Price Center

On-Campus Transportation

A campus shuttle—from 7 am until 6 pm—stops near the Price Center and campus housing at the Village (North campus). It It is a 1/2 mile walk from the Village to the Price Center and for those driving there is parking at Gilman Parking Deck. More information is available at http://transportation.ucsd.edu/shuttles/campus-loop.html #Academic-Break-and-Summer-Sched

Other useful information

A large food court is located inside the Price Center.

The Village Market, at the residence halls, is open 7 am to 8 pm with snacks, beverages, and grab and go food.

There is also an Expresso place in Building 4 on the East side of the Village. Restaurants close by: Ocean View in the Village with a mini food court. One block down and across the street is the restaurant Bella Vista

Uber and Lyft services are available for transportation into La Jolla or elsewhere.

SUNDAY

8:45 – 5:00 Council Meeting,

Village Tower West, 15th Floor, 15C

6:00 – 9:00 **MIXER**

Institute of the Americas - Plaza

MONDAY, 8:00 am - 12:30 pm

Theater

OPENING CEREMONY

8:00 Surendra Dara, Chair, Organizing Committee
Johannes Jehle, SIP President
Glenda Humiston, Vice President of Agriculture
and Natural Resources Division, University of
California

8:20 Awards Presentations
Monique van Oers, Chair, Awards and
Student Committee

SIP Founders' Golden Jubilee Celebration: Historical Perspectives of the Society and Invertebrate Pathology

Organizers: Peter Krell, Jimmy Becnel

Introduction - Jimmy Becnel, Chair, Founder's Lecture Committee

Looking Back: 50 Years of SIP - Elizabeth W. Davidson

•1•

School of Life Sciences, Arizona State University, Tempe, AZ, USA

Corresponding author: e.davidson@asu.edu

The concept of a society for the study of diseases of invertebrates began more than 10 years before the founding of SIP. The International Conference on Insect Pathology and Biological Control, hosted by Jaroslav Weiser in Prague in 1958, brought scientists together from around the world to share findings and ideas. Edward Steinhaus, professor at University of California, Berkeley, mentored many of the early scientists in our field, and was founding editor of the Journal of Insect (now Invertebrate) Pathology. In 1967 Steinhaus brought together 8 scientists to discuss the concept of establishing a Society. The first SIP meeting was held at Ohio State University in 1968. SIP has remained international, diverse, and collaborative ever since. We have met in 16 countries, and welcome members from around the world. We are looking forward to many more wonderful SIP meetings.

Personal reflections of a founding SIP member - Donald W. Roberts



Department of Biology, Utah State University, Logan, Utah, USA.

**Corresponding author: donald.roberts@usu.edu

It will be a pleasure to join you, my fellow Society for Invertebrate Pathology (SIP) members, in celebrating the jubilee (50th) anniversary of our annual meetings. It will be extremely gratifying to note how well our society has survived the vicissitudes of the years, and that it continues to serve its members in very productive and caring ways. My personal SIP history began due to my association with a key member of SIP's organizing committee, Dr. Edward Steinhaus. I joined his laboratory as a student the same year (1959) he initiated publication of the Journal of Invertebrate Pathology, so I was very aware of his desire to increase the visibility of this branch of science. When he and others initiated SIP in 1967, I immediately joined as a Founding Member in the hope that I could, in some way, help this group become established. It grew, became visible, and, in fact, was a key element in distinguishing Invertebrate Pathology as an identifiably separate discipline. Over the years, SIP became very important to my professional development. In fact, I now wonder if I would have fared nearly as well as I have professionally, as limited as that might be, had SIP never come into being. How would you answer that question concerning the existence of SIP in your professional development? We must be very thankful to the small group that 50 years ago expended the time and effort to initiate our unifying organization, SIP. One positive characteristic of our society is our exceptionally high level of friendship between our members. We care about and frequently help each other in both professional and personal advancement, and this provides the climate for developing very useful long-term research connections. SIP has a long history of providing awards for exceptional scientific performance, and these awards were instituted as a way to help our members advance both academically and professionally. Our society welcomes new members and encourages them to volunteer for activities that improve its effectiveness for its members. My advice and invitation to any of you who currently feel you are not fully involved with our SIP movement is that, if you have good ideas to improve SIP, please make them known, and also offer to help implement those ideas, as well as proposals of others that you support. In summary, my experience with SIP has been almost universally positive. Thank you, my fellow members, for making it so.

SIP Divisions, the Beginnings – Peter J. Krell¹, James J. Becnel²



¹Department of Molecular and Cellular Biology, University of Guelph, Canada; ²USDA, ARS, CMAVE, Gainesville, Florida, USA Corresponding author: pkrell@uoguelph.ca

In 1969 Mauro Martignoni was appointed Chair of the Divisions Committee by then President Albert K Sparks with members Wayne Brooks, Horace Denis Burges, Phyllis T Johnson, Carl Sindermann and Marenes R Tripp. The first Division was spear headed by Victor Sprague in August 1969. The now Division of Microsporidia was approved in 1970 electing Chairman Jiří Vávra, Vice Chairman Roy McLaughlin, Secretary Ann Cali with Council Members John Henry and Edwin Hazard. A decade later, the second Division, Division of Microbial Control was conceived in 1979 under Chairman Jaroslav Weiser and later James D Harper as ad hoc Chairman. An executive consisting of Chairperson James D Harper, Vice Chairperson H Denis Burges, Secretary-Treasurer James Fuxa and Members-at-Large Don Roberts and John Cunningham were elected. The third and fourth Divisions were formalized at the 1995 meeting in Ithaca, NY, USA. The Division of Bacteria elected Chair Andre Klier, Vice Chair Barbara Knowles, Secretary Treasurer Bill Moar and Members-at-Large Sue Macintosh and Elizabeth (Betty) Davidson. The Division of Viruses elected Chair Norman Crook, Vice Chair Suzanne Thiem, Secretary Treasurer Just Vlak and Member-at-Large Bryony Bonning. The fifth Division, Division of Fungi, was approved in 1998 and in 2000 elected Chair Stephen Wraight, Chair-Elect Judith Pell, Secretary Treasurer Michael Brownbridge and Members-at-Large Travis Glare and Paresh Shah. The sixth Division, Division of Nematodes, was approved in 1999 and at its 2000 meeting elected Chair Itamar Glazer, Chair-Elect Noel Boemare, Secretary Treasurer Albrecht Koppenhöffer with Members at Large Bryon Adams and Patricia Stock. The seventh Division was the Division of Beneficial Invertebrates and in 2009 elected Chair Grant Stentiford, Chair-Elect Elke Genersch, Secretary Treasurer Kate Aronstein with Trustees Regina Kleespies and Huang Shaokang. The first Division Student Representatives were Christina Campbell (2004) for Microsoporidia, Todd Ugine (2005) for Microbial Control, Claudia Perez (2005) for Bacteria, Ernst Jan Scholte (2005) for Fungi, Heather Smith (2005) for Nematodes, Jondavid de Jong (2006) for Viruses and Kelly Bateman and Eva Forsgren (2009) for Beneficial Insects. The SIP Divisions continue to unify members and provide a unique forum for each discipline of Invertebrate Pathology.

Microsporidia - Ann Cali

•4•

[See Addendum]

Microbial Control of Insects 1967-2017: How far we have come (and how far we still have to go) – *Stefan T. Jaronski*



Agricultural Research Service, U.S. Department of Agriculture Northern Plains Agricultural Research Laboratory Sidney MT USA 59270

Corresponding author: stefan.jaronski@ars.usda.gov

In 1967, the year Ed Steinhaus and colleagues created the Society for Invertebrate Pathology, the biopesticide industry was still, more or less, in its infancy esp. in the U.S. Greater strides had been made in eastern and central Europe, but there were very few commercial products anywhere. The industry struggled through the 1970s, 1980s and even the 1990s, with momentum generally in the hands of small-medium enterprises. In the past two decades, however, the pace of commercial development of microbials has quickened all over the world. Now, even the multinationals have begun to stick their toes in microbial waters. We still have a long way to go. Despite increasing acceptance (e.g., 38% of California growers polled say they use microbial agents to manage pests and pathogens), chemical pesticides still hold sway in most regions. One problem is that many times microbial agents have been cast into a chemical paradigm, in which the microbial is expected to perform like a chemical, as good as chemical, to simply replace a chemical. What is often forgotten is that microbials should be one tool in an integrated system. (Remember the original concept of Integrated Pest Management?) Recently, efforts have been made to integrate microbials with other measures, from cultural practices to other biologicals, to create successful systems to manage insect pests. More research and implementation is needed, however. This, in my opinion, is the path for greater acceptance of

microbials, as well as biologicals in general. This is the direction we need to pursue. Harder? Yes. More complicated? Yes. But necessary.

10:00-10:45 am

Coffee Break

•6•

Insecticidal Bacteria: A remarkable success for invertebrate pathology – Brian A. Federici¹, Jean-Luis Schwartz²

¹Department of Entomology and Institute for Integrative Genome Biology,
University of California, Riverside, California, USA; ²Département de
pharmacologie et physiologie, Faculté de médecine,
Université de Montréal, Montreal, Quebec, H3C 3J7, Canada

Corresponding author: brian.federici@ucr.edu

The modern era of research on insecticidal bacteria, especially Bacillus thuringiensis (Bt), began after WWII when Edward Steinhaus at the University of California and international collaborators such as H. Dulmage, D. Burges, H. de Barjac developed standardized bioassays and serological identification for Bt strains. These advances yielded the first insecticides for lepidopteran pests based on B. t. subsp. kurstaki (HD1). Later, B. t. subsp. israelensis, active against mosquito and black fly larvae, was discovered, commercialized in the 1980's, and used successfully in WHO's Onchocerciasis Control Program in west African countries, and for controlling mosquitoes. Advances in molecular biology led to Schnepf and Whiteley's discovery in 1981 that HD1's bipyramidal crystal contained three different Cry proteins, Cry1Aa, Cry1Ab, and Cry1Ac. This quickly advanced basic and applied Bt research, resulting in new bacterial insecticides and Cry-based genetically engineered crops in the 1990's. Bt cotton and Bt corn were rapidly adopted by farmers in the U.S. and other countries, and current seed sales exceed \$US 10 billion per year, with annual revenues for these crops now more than \$300 billion, which includes engineered soybeans. This success was paralleled by basic research in the 1980's on Cry protein mechanism of action, showing insect gut damage was due to colloid-osmotic lysis, followed in the 1990's by demonstrating Cry proteins formed pores in target cell membranes and triggered cellular signaling. Simultaneously, APN, cadherin and ALP proteins were identified as Bt receptors, and 3-D atomic structures of Cry and Cyt toxins became available, opening the way in the 1990's and 2000's to structure-function analyses of molecular determinants for receptor binding and pore formation. The current millennium has witnessed discovery of new Bt toxins (acting as single or binary toxins against new targets including mammalian tumor cells, displaying completely new 3-D structures); a new class of Cry receptors was identified (ABC transporters); and cellular defense mechanisms induced by these proteins have been discovered. This progress is a remarkable success for insecticidal bacteria, but the future is even brighter made possible by synthetic biology techniques including rapid sequencing of new toxins and retargeting others for improved pest control.

Fifty years of Invertebrate Virology: a stellar course – Just Vlak^{1, 2}, Zhihong Hu²



¹Laboratory of Virology, Wageningen University, Wageningen,
The Netherlands; ² Wuhan Institute of Virology,
Chinese Academy of Sciences, Wuhan, People's Republic of China

Corresponding author: just.vlak@wur.nl

Invertebrate virology over the last fifty years has the Society for Invertebrate Pathology (SIP) a lot to thank. SIP brought together interested (and interesting!) scientists and promoted invertebrate pathology as an integrative discipline. After establishment of the Society (1967) the focus was primarily on virus discovery and use as biocontrol agents. The focus soon shifted to molecular and cell biological approaches. The Society willingly took in and facilitated these emerging disciplines by providing a platform for a new breed of young and committed virologists. Invertebrate virologists also brought their research to the forefront of science and the scientific community at large, with publications in high ranking journals and major fundamental advances like discovery of the anti-apoptotic response and practical applications such as production of pharmaceuticals, diagnostic reagents and even effective vaccines. Although molecular virology was dominant for some time, increased interest in environmental issues in the late 1990s promoted the resurgence of invertebrate virus ecology, also stimulated by major diseases in commercial invertebrates, and re-appreciated field oriented science. The molecular and ecological disciplines, and associated techniques, mutually benefitted and promoted the molecular ecology of invertebrate viruses. The prominence of Invertebrate Virology was institutionalized by the formation of the SIP Virus Division in 1995. While initially analysis of viruses relied on SDS-PAGE for proteins and DNA restriction enzyme profiles, the 2000's that gave way to sequencing of first viral DNA/RNA fragments then genes, genomes and metagenomes. Molecular engineering came to its full potential providing detailed insights in the functioning of virus genes. Molecular detection/identification techniques became indispensable for invertebrate virus research and virus registrations. Understanding the interaction between virus and invertebrate host, at levels ranging from cell to organism to populations to ecosystem, now became feasible. This is paramount to understanding the response of organisms to virus infection and essential to control or mitigate invertebrate virus diseases. It also allows tailored application of viruses for controlling insect populations. The lesson learned here is that fundamental research of societal relevance necessarily precedes a rational exploitation of this knowledge. The SIP can take pride in having fostered this development in Invertebrate Virology over the last fifty years.

Fifty years of Fungi

•8•

in fewer than fifteen minutes?! - Richard A. Humber¹, Jørgen Eilenberg

¹Emerging Pests and Pathogens Research, Robert W. Holley Center for Agriculture & Health, Ithaca, New York 14853-2901, USA; ²Department of Plant and Environmental Sciences, University of Copenhagen, Thorvaldsensvej 40, DK-1871 Frederiksberg C, Denmark Corresponding authors: richard.humber@ars.usda.gov; jei@plen.ku.dk

This presentation attempts to condense a half century of very active progress and significant transitions in our knowledge of all aspects of entomopathogenic fungi (especially EPFs from the most studied taxa fromAscomycota and Entomophthoromycota) and their fungus-like allies (biflagellate, oomycetous pathogens). Subjects to be covered include our expanding knowledge of the biodiversity and ecology of EPFs; new understandings about the life histories and pathobiology of EPFs; their interaction with insects, plants and environment; access to formal training in basic and applied aspects of insect mycology; and the astonishing (and, too often, bewildering) changes in our knowledge of the systematics and classifications of these fungi. The status of the practical uses of EPFs as biological control agents and as sources of biologically active compounds will also be noted.

Symbiosis between entomopathogenic nematodes and bacteria: from taxonomy to ecology and genomics – *Harry K. Kaya*



Department of Entomology and Nematology, University of California, Davis, CA 95616 Corresponding author: hkkaya@ucdavis.edu

Different groups of nematodes including mermithids, tylenchids, diplogasterids, and rhabditids infect insects. Among the rhabditids, two families, Steinernematidae and Heterorhabditidae, referred to as entomopathogenic nematodes (EPNs), have received the most attention over the past four decades. These EPNs are associated with bacteria in the genus Xenorhabdus for Steinernema and Photorhabdus for Heterorhabditis, which form a nematode-bacterium complex that work mutualistically to kill their insect host within 48 hours. Since the first description of Steinernema (Aplectana) kraussei by Gotthold Steiner in 1923, more than 100 species of both nematode genera have been described. In 1936, Rudolf Glaser was the first to produce S. glaseri in vitro and apply it for microbial control of the Japanese beetle, Popillia japonica. Although the application of S. glaseri was partially successful, Glaser considered that the contamination by microorganisms affected nematode production and efficacy. In 1959, S. R. Dutky demonstrated that a symbiotic bacterium, later to be described as Xenorhabdus nematophila, is retained and required for successful production and use of S. carpocapsae. In the 1980s, commercial companies mass produced several EPN species in vitro and made them available to researchers for efficacy trials against a number of soil insect pests. With some successes and many failures, more fundamental research was required to define what environmental and biotic factors affected efficacy.

Understanding the response of EPNs to abiotic and biotic factors is essential for increasing the success of EPNs against an insect pest. Abiotic factors such as soil type, temperature, UV light, and humidity and biotic factors such as nematode foraging behavior and persistence, host susceptibility and resistance, and host density affected nematode efficacy. Recent studies show that EPNs not only respond to host cues but also to nematode pheromones and plant signals. Moreover, genomics and related studies, as well as the indepth knowledge of how the bacteria maintain their mutualistic relationship with EPNs and how bacterial virulence affects efficacy will further advance EPN use in microbial control programs.

Diseases of Beneficial Invertebrates – *Kelly Bateman*¹, *Elke Genersch*², *Grant Stentiford*¹, *Helen Hesketh*³



¹European Union Reference Laboratory (EURL) for Crustacean Disease, Cefas, Barrack Road, The Nothe, Weymouth, Dorset, DT4 8UB, UK; ²Institute for Bee Research, Friedrich-Engels-Str. 32, 16540 Hohen Neuendorf; ³Centre for Ecology & Hydrology, Benson Lane, Crowmarsh Gifford, Wallingford, Oxfordshire, OX10 8BB, UK.

Corresponding authors: kelly.bateman@cefas.co.uk elke.genersch@hu-berlin.de

The Diseases of Beneficial Invertebrate (DBI) division was established in 2009 at the Park City meeting of the Society for Invertebrate Pathology (SIP). The division addresses diseases of invertebrates that are generally considered as non-pest species, covering aquatic and terrestrial invertebrates which are ecologically or economically beneficial. The division focuses on the pathology and host-pathogen interface within various host groups (crustaceans, molluscs, insects and arachnids) and addresses significant bigger picture issues that can be studied via these relationships (e.g. climate change, ocean acidification, epizootiology, pollination, insects for food and feed, disease mitigation). When first established the Society for Invertebrate Pathology possessed a strong interest in aquatic pathology, in particular mollusc and crustacean disease. Over the years this area of the society had diminished, a particular aim of the Division was to attract pathologists and disease experts with an interest in pathogens and pathology of aquatic animals in order to improve the standing of this discipline in the wider society. Alongside this, the significantly increased interest in bee diseases and diseases of other beneficial terrestrial invertebrates co-stimulated the formation of a host directed division. The division has held some key symposia in recent meetings, in 2011 an OECD funded symposia was held to discuss Disease in Aquatic Crustaceans: Problems and Solutions for Global Food Security and in 2015 an additional OECD symposia was funded to examine Microsporidia in the Animal to Human Food Chain: An International Symposium to Address Chronic Epizootic Disease. In 2015, the Founders Lecture honoree was Dr Phyllis Johnson, a pioneer of aquatic invertebrate pathology and a key figure in establishment of the SIP as we know it today. This presentation will provide an overview of the division from conception to now.

The SIP-JIP connection through the years: Supporting invertebrate pathology research – Leellen Solter¹,



Dale Seaton², Brian Federici³

¹Illinois Natural History Survey, Prairie Research Institute, University of Illinois, Champaign, IL, USA; ² Elsevier, New York, USA, ³Department of Entomology & Institute of Genome Biology, University of California, Riverside, CA, USA Corresponding author: Isolter@illinois.edu

The Journal of Invertebrate Pathology and the Society for Invertebrate Pathology have been partners in promoting and supporting invertebrate pathology research since the inception of the Society in 1967. The journal was inaugurated first, in 1959, as Journal of Insect Pathology. The name was changed to Journal of Invertebrate Pathology in 1965, at which time Edward Steinhaus wrote to Albert Sparks that he had proposed a society of the same name. A subsequent global survey of invertebrate pathologists showed overwhelming support for the establishment of the Society and inclusion of the Journal of Invertebrate Pathology as its official organ. In 1968, the Journal of Invertebrate Pathology was first Published under the auspices of the Society for Invertebrate Pathology. Since 1965, over 6,200 papers have been published with Society members well represented as authors. We highlight the history of the JIP-SIP connection as well as researchers and

papers published that garnered significant attention from the scientific community. As well, we celebrate 50 years of enthusiastic encouragement and advancement of invertebrate pathology!

12:15 Presentation of Plaques - Jimmy Becnel

MONDAY, 12:30 - 1:30 pm **LUNCH BREAK**

JIP Editorial Board Meeting and Luncheon Price Center West, Bear Room

MONDAY, 1:30 – 3:30 pm

Theater

BACTERIA AND MICROBIAL CONTROL CROSS-DIVISIONAL SYMPOSIUM

Challenges for Biocontrol in Latin America

Organizers and Moderators: Travis Glare and Ken Narva

The use of Bacillus thuringiensis and Baculovirus based biopesticides in Brazil 1:30 •12•

Fernando H. Valicente

¹Embrapa Maize and Sorghum, Rod. MG. 424 km 65, 35701970, Sete Lagoas, MG, Brazil ${\it Corresponding\ author:} fernando.valicente@embrapa.br$

The search for Baculovirus and Bacillus thuringiensis (Bt) based biopesticides has increased since 2012/13, when Helicoverpa armigera was detected in Brazil. Embrapa Maize and Sorghum has a Bt and a baculovirus collection, with more than 4,600 Bt strains and more than 100 baculovirus isolates, collected from different regions of Brazil. Most of these insect pathogens have been tested to insect pests, such as: fall armyworm Spodoptera frugiperda, black armyworm S. cosmioides, southern armyworm S. eridania, corn earworm Helicoverpa zea, cotton bollworm H. armigera, soybean looper Chrysodeixis includens, the green belly stink bug Dichelops sp and sugar cane borer Diatraea saccharalis. Some baculovirus, all of them NPV, have been developed as biopesticides (wettable powder-WP) to control fall armyworm, soybean looper and cotton bollworm. Baculovirus spodoptera (isolate 6) is very efficient and doesn't cause the disruption of the integument of the caterpillar, and this factor is essential in a large-scale production system. Isolates for cotton bollworm where identified as HearNPV and showed a CL₅₀ of 4.1 x 10⁴, and is formulated as WP. B. thuringiensis isolates have been tested for many cry (1, 2, 3, 4, 5, 6, 8, 9, 12, 13, 14, 22 and 51) and Vip (1, 2, vip3Aa1, vip3Ah1, vip3Ae1, vip3Ba1, vip3Aa2, vip3Af1) genes in order to control from caterpillars to sucking bugs. Some Bt isolates have been developed as biopesticides to control fall armyworm, soybean looper and cotton bollworm. Brazilian companies, with high production capacity, have signed contracts with Embrapa in order to put these products in the market in the near future. Baculovirus (I6) and Bt have been registered. Baculovirus for fall armyworm is already in the market.

Cry1F resistance among lepidopteran pests: A model for improved reistance management? 2:00 •13•

Ana María Vélez

University of Nebraska, Lincoln

The Cry1Fa protein from Bacillus thuringiensis (Bt) is known for its potential to control lepidopteran pests, especially through transgenic expression in maize and cotton. The maize event TC1507 expressing the cry1Fa toxin gene became commercially available in the United States in 2003 for the management of key lepidopteran pests including the European corn borer, Ostrinia nubilalis, and the fall armyworm, Spodoptera frugiperda. A high-

dose/refuge strategy has been widely adopted to delay the evolution of resistance to event TC1507 and other transgenic Bt crops. Efficacy of this strategy depends on the crops expressing a high dose of the Bt toxin to targeted pests and adjacent refuges of non-Bt host plants serving as a source of abundant susceptible insects. While this strategy has proved effective in delaying O. nubilalis resistance, field-evolved resistance to event TC1507 has been reported in S. frugiperda populations in Puerto Rico, Brazil, and the southeastern United States. This talk will examine available information on resistance to Cry1Fa in O. nubilalis and S. frugiperda and discuss how this information identifies opportunities to refine resistance management recommendations for Bt maize with an emphasis in Latin America.

Microbial products for agriculture in Uruguay

Nora Altier¹, Elena Beyhaut¹, Federico Rivas^{1,2} Eduardo Abreo¹, **Trevor A. Jackson**³

¹ INIA, Instituto Nacional de Investigación Agropecuaria, Uruguay; ²BioProtection Research Centre (BPRC), Lincoln University, New Zealand; ³AgResearch, New Zealand. Corresponding author: naltier@inia.org.uv

Biotechnologies based on microbial resources have made possible the development of biological products to improve plant protection and plant nutrition, without compromising the environment. These technologies also contribute to the achievement of high end market requirements for low agrochemical use. The Platform of Microbial Products for Agriculture at INIA-Uruguay (National Institute for Agricultural Research) has emphasized the role of microorganisms in biocontrol of plant pests and diseases, and in nitrogen and phosphorus plant nutrition. The long term strategy lies on strengthening research facilities and team work, recruitment and continuous training of human resources, research and technology ventures with the public and private sectors, fostering a legal framework for registration and intellectual property of products, and promoting education and extension for farmer adoption. Research areas include: (1) biological control, (2) biological nitrogen fixation, (3) microbial mediated phosphorus availability to plants, and (4) bioproduction and formulation. Current biocontrol projects focus on identifying strains of microbe for product development to be incorporated in Integrated Pest Management (IPM) for targeted crops and pests. Biological nitrogen fixation is being efficiently exploited by inoculating legumes with suitable rhizobial inoculants, supported by appropriate legislation, which sets requirements for product registration, mandatory strain recommendation, and quality surveillance of commercial inoculants. Supporting research addresses inoculation and seed coating technologies, fungal endophytes, as well as metagenomics for assessment of microbial diversity and soil health.

Fungal-based biopesticides in Brazil: **Challenges and Opportunities**

3:00



Italo Delalibera Júnior

Department of Entomology and Acarology, University of São Paulo, Brazil Corresponding author: delalibera@usp.br

Brazil is the world's largest consumer of chemical pesticides, but adoption of biological control has grown at a rapid pace in the country. The future of biological control will depend on scientific research guidelines and regulatory mechanisms to ensure the quality of products available on the market. A very limited number of isolates of few species of microorganisms is marketed today in the country in the form of biopesticides. Motivated for facilities and speed in the registration of phytosanitary products for organic agriculture, many companies commercialize the same microorganisms. The same happens with sugarcane mills that use entomopathogenic fungi for their own consumption. The micopesticides in the country are based on aerial conidia produced by solid fermentation almost exclusively using rice, with production system without substantial innovations since the 70's. More efficient production systems must be developed, either by automating stages in production by solid fermentation or by producing other structures such as blastospores and microsclerotia by liquid fermentation to meet the growing demand. Many microbial products in the Brazilian market are of questionable quality, unformulated and with short shelf life. To improve these products it is necessary to develop formulations taking into account the pest and target crop and adapted to the Brazilian climate conditions. Fungi are used for the control of few pests only through aqueous sprays. The

development of biological control strategies in Brazil is a challenge, considering the large continuous areas of the country's agricultural land, growing crops successively over a long period of the year and the various pest cycles per year. It is in this context that scientific research must play a fundamental role in the direction and future of biological control in the country.

The Forum

Virus Division #1

Moderators: John P. Burand, Robert L. Harrison

AcMNPV adaptive evolution in varying environment

1:30 •16•

Aurélien Chateigner, Yannis Moreau, Cindy Pontlevé, Carole Labrousse, Elisabeth A Herniou

Institut de Recherche sur la Biologie de l'Insecte, UMR CNRS 7261. Université de Tours, Faculté des Sciences et Techniques, Tours, France Corresponding author: elisabeth.herniou@univ-tours.fr

Genetic variation underpins the evolutionary process of adaptation. As populations become adapted to different environments, they diverge from one another. Baculoviruses infecting different host species thus usually belong to different species. Yet some viruses, such as Autographa californica multiple Nucleopoly-hedrovirus (AcMNPV), have retained the capacity of infecting many host species. To understand how AcMNPV could draw on standing genetic variation to adapt to different host species we undertook an experimental evolution protocol. A highly polymorphic AcMNPV population was taken through 10 in vivo infection cycles in 4 host species of various susceptibilities, to create 50 evolutionary lines in which the virus was allowed to evolve either as specialist on separate species (4x10 lines) or as generalist on all the hosts (10 lines), whereby the viral population resulted from the infection of all hosts at each generations. We then characterized the genetic make up of the original and evolved baculovirus populations by ultra-deep Illumina sequencing. Using a population genomics approach, we then estimated the global fitness and genetic diversity of each of the evolved populations and their divergence from one another. We found that the specialist lines that evolved on the same host species were more similar to one another than to other lines, especially in the most susceptible hosts, showing experimental evolution could lead to specific adaptation. Furthermore, we found a general diminution of genetic diversity in the specialist lines compared to the ancestral population. The viral lines that could adapt to more resistant host retained higher genomic diversity than those that did not, showing genetic variation is an important component of baculovirus adaptation. Evolution in all the hosts led to high genetic diversity in the generalist lines, which could foster ecological resilience in variable environments such as those encountered by the virus in natural landscapes.

Role of Plutella xylostella nucleopolyhedrovirus-CL3 ie2 in host range adaptation

Rahul P. Hepat¹, Martin A. Erlandson², Robert L. Harrison³, Leslie G. Willis¹. David A. Theilmann¹

¹Summerland Research and Developmental Centre, Agriculture and Agri-Food Canada, Summerland, British Columbia, Canada; ²Saskatoon Research and Developmental Centre, Agriculture and Agri-Food Canada, Saskatoon, Saskatchewan, Canada; ³Invasive Insect Biocontrol and Behavior Laboratory, Beltsville Agricultural Research Center, USDA Agricultural Research Service, Beltsville, MD, USA Corresponding author: david.theilmann@agr.gc.ca

The diamondback moth (Plutella xylostella) is one of the most important destructive insect pests of cruciferous crops worldwide. Previous studies identified both alpha- and betabaculoviruses that can infect and kill this important lepidopteran pest. The genome of the alphabaculovirus isolate $% \left\{ 1,2,...,n\right\}$ Plutella xylostella nucleopolyhedrovirus-CL3 (PlxyNPV-CL3) was recently sequenced and was found to be nearly identical (98.5%) to the genome of Autographa californica multiple nucleopolyhedrovirus (AcMNPV). Interestingly PlxyNPV-CL3 is highly lethal for P. xylostella, whereas AcMNPV is not. Therefore, any differences in the genome of these two viruses may identify genes that are required for host adaptation. The dominant difference between the PlxyNPV-CL3 and AcMNPV-C6 genome is the open reading frame of the ie2 gene. The predicted amino acid sequences of the

two IE2 proteins exhibit only 37.5% similarity. To determine if PlxyNPV-CL3 IE2 is responsible for adaptation of AcMNPV to P. xylostella, recombinant viruses were generated using bacmids. These viruses were an ie2 knockout (vie2KO), and the ie2 knockout repaired with AcMNPV or PlxyNPV-CL3 ie2 tagged with HA epitope (vie2KO-acie2HA and vie2KO-pxie2HA respectively). Each of these viruses was tested for replication in various cell lines which including Sf9, Tn5b1-4, and PxE-Po#583. Previous studies have not analyzed the impact of a complete AcMNPV ie2 knockout. Surprisingly, our results showed that deletion of ie2 led to a greater than 90% reduction in budded virus (BV) production compared to wildtype virus. In addition, occlusion body (OB) production was also greatly reduced. Initial results with vie2KO-pxie2HA indicate that PlxyNPV-CL3 IE2 can rescue vie2KO producing BV titers nearly as high as AcMNPV IE2. Further studies analyzing virus infection in P. xylotella larvae will be reported.

Tracing the origin of nucleopolyhedrovirus from an invasive species, winter moth Operophtera brumata L.







(Lepidoptera: Geometridae)

Hannah J. Broadley¹, Woo Jin Kim², John P. Burand³, Joseph S. Elkinton¹ ¹Organismic and Evolutionary Biology, University of Massachusetts,

> Amherst, MA, USA.; ²Department of Agricultural Biotechnology, Seoul National University, South Korea ³Department of Microbiology, University of Massachusetts, Amherst, MA, USA Corresponding author: hbroadley@cns.umass.edu

The exchange of pathogens between populations or related species can mediate insect invasions in complex ways, either suppressing or stimulating population growth. We evaluated sources of nucleopoly-hedroviruses (NPV) in winter moth Operophtera brumata L. (Lepidoptera: Geometridae). Invasive populations of this destructive insect from Europe may be infected with NPV from its source population in Europe or from native, sympatric geometrids. Winter moth may also introduce novel isolates of NPV to native species when infesting new regions. We sequenced two NPV loci (polyhedrin and p74) from winter moth and its congeners across winter moths' introduced and native range and ran phylogenetic analyses to (1) evaluate differences between NPV isolates of the three populations of invasive winter moth in the United States and (2) compare the NPV from these invasive populations to potential native winter moth source populations and to sympatric geometrids. We found that the virus from winter moth and Bruce spanworm are closely related and constitute a clade within the Alphabaculovirus Group 2 NPVs but are distinct, suggesting that both strains are highly host specific. However, evidence indicates that winter moth NPV may cross-infect related geometrids in Europe. Further, we suggest that the winter moth virus found in the northeastern U.S. may originate from northern Europe.

Genome features of Troides aeacus nucleopolyhedrovirus (TraeNPV) from golden birdwing larvae (Troides aeacus)



Yu-Feng Huang¹, Tzu-Han Chen¹, Zi-Ting Chang², Se Jin Lee³ Jong Cheol Kim³, Jae Su Kim³, Chung-Hsiung Wang⁴, Kuo-Ping Chiua¹, Yu-Shin Nai²

¹Genomic Research center, Academic Sinica, Taiwan; ²Dept. of Biotechnology and Animal Science, National Ilan University, Yilan, Taiwan; ³Dept. of Agricultural Biology, College of Agriculture & Life Sciences, Chonbuk National University, Jeonju, Korea; ⁴Department of Entomology, National Taiwan University, Taipei, Taiwan Corresponding author: vsnai@niu.edu.tw

Golden birdwing butterfly (Troides aeacus) is rarely observed species in Taiwan, but recently we found a population, which showed a classic symptom of nucleopolyhedrovirus infection. Primary PCR was performed to confirm NPV infection in T. gegcus larvae. To better understand this NPV, the nucleotide sequence of the NPV DNA genome was determined by a high throughput genome sequencing (NGS) from infected larvae. The genome of TraeNPV consisted of 125,523 bases, and had a G+C content of 40.35% and contained 142 putative open reading frames (ORFs). The gene content and gene order of TraeMNPV were similar to those of AcMNPV, with 90 ORFs identified as homologous to those reported in the AcMNPV genome. Eight homologous regions (hrs) were identified containing 29 repeated sequences

composed of 30bp imperfect palindromes. Analyses of the gene parity, gene homologues and a phylogenetic analysis suggested that TraeMNPV is a Group I NPV that is closely related to AcMNPV, BmNPV and PlxyMNPV. From the preliminary result of range test, it showed that TraeNPV expanded host range to Silkworm (Bomyx mori). More biological data and the further genome-wide analysis would clarify the identity of TraeNPV.

The Operophtera brumata nucleopolyhedrovirus (OpbuNPV) represents a new and distinct lineage within genus Alphabaculovirus

2:30 •20•

Robert L. Harrison¹, Daniel L. Rowley¹, Joseph Mowery², Gary R. Bauchan², John P. Burand³

¹Invasive Insect Biocontrol and Behavior Laboratory, Beltsville Agricultural Research Center, USDA Agricultural Research Service, Beltsville, Maryland, USA; ²Electron and Confocal Microscopy Unit, Beltsville Agricultural Research Center, USDA Agricultural Research Service, Beltsville, Maryland, USA; ³Department of Microbiology, University of Massachusetts-Amherst, Amherst, Massachussetts, USA

Corresponding author: Robert.L.Harrison@ars.usda.gov

Operophtera brumata nucleopolyhedrovirus infects larvae of the winter moth, Operophtera brumata. Recently, an isolate of this virus, OpbuNPV-MA, was described from a population of winter moth in Massachusetts, USA. This isolate was characterized by electron microscopy (EM) of OpbuNPV occlusion bodies and by sequencing of the viral genome. The occlusion bodies of OpbuNPV-MA were polyhedral in shape and measured approximately 1-1.5μm in diameter. Virions within the occlusion bodies consisted of a single nucleocapsid within each unit envelope. The OpbuNPV-MA genome was found to be 119,054 bp in length, with a 39.83% G+C nucleotide distribution. Five homologous repeat regions consisting of 40 – 44 nt imperfect inverted repeats were identified in the sequence. A total of 130 ORFs were identified and annotated, including the 38 core genes found in all viruses of family Baculoviridae. ORFs encoding homologues of DNA photolyase, DNA ligase III, and LdMNPV host range factor-1 were found, as well as two separate ORFs encoding homologues of nicotinamide riboside kinase 1. In addition, 22 ORFs with no homologues in baculovirus genomes were identified, including an ORF with sequence similarity to an uncharacterized winter moth transcript. OpbuNPV-MA homologues of ORF1629/ac9, ribonucleoside-diphosphate reductase small and large subunits, dUTPase, chaB/ac58/59/60, and ubiquitin/ac35 were identified that exhibited little to no sequence similarity with other baculovirus homologues and did not group with baculovirus taxa in phylogenetic trees. Phylogenetic inference with concatenated alignments of core gene amino acid sequences placed OpbuNPV-MA on a branch lying outside the group I and group II clades of genus Alphabaculovirus. These results collectively indicate that OpbuNPV-MA represents an alphabaculovirus lineage that appeared early during the divergence of Alphabaculovirus.

Crossing kingdoms: Providence virus (Family Carmotetraviridae) infects and replicates in plants, insects and mammalian cell lines

> Rosemary A Dorrington¹, Meesbah Jiwaji¹, Janet Awino Awando¹, Ritah Nakayinga^{1,2}, Mart-Mari de Bruyn¹

> > ¹Dept of Biochemistry and Microbiology, Rhodes University, Grahamstown, South Africa, 6140; ²2Institute of Allied Health Sciences, International Health Sciences University, Kampala, Uganda $Corresponding\ author: r.dorrington@ru.ac.za$

Providence virus (PrV) is a positive-sense single-stranded RNA virus belonging to the Family Carmotetraviridae. It was first isolated from a persistently infected Helicoverpa zea midgut cell line and until recently was the only tetravirus able to replicate in cell culture. While the PrV capsid displays the T=4 icosahedral architecture that is typical of the insect-infecting tetraviruses, its Carmovirus-like replicase is related to those of the plant-infecting Tombusviridae and umbraviruses. This has led us to hypothesize that PrV originated by re-assortment between a plant and an insect virus. Here we present evidence that PrV has indeed arisen via horizontal virus transfer between plants and animals. In addition to infecting lepidopteran insects, PrV readily infects and replicates in human tissue culture cells, establishing

persistent infections in cervical (HeLa) and breast (MCF7) cancer cell lines. We show that functional conservation between the PrV and Tombusvirus replicases extends to the ability of PrV to replicate in plants. We also present data demonstrating that the spread of the virus from the point of infection and the production of infectious PrV particles in Vigna unguiculata (cowpea) plants. These and other data lead us to conclude that PrV is an extant example of horizontal virus transfer between plants and animals which, remarkably, has retained its roots in the plant world, while expanding its host range from insects to mammals.

No longer a simple virus: The alphatetravirus, Helicoverpa armigera stunt virus, expresses three small proteins that co-localize with the viral replicase

> Rosemary A Dorrington¹, Mart-Mari de Bruyn¹, Gareth Edward Hughes^{1,2}, James Roswell Short^{1,3}, Meesbah Jiwaji²

> > ¹Dept of Biochemistry and Microbiology, Rhodes University, Grahamstown, South Africa, 6140; ²Celtic Molecular Diagnostics (Pty) Ltd, Wynberg, Cape Town, South Africa, 780; 3 Illumina Inc. 5200 Illumina Way, San Diego, California, USA, 92122 Corresponding author: r.dorrington@ru.ac.za

Tetraviruses are small insect RNA viruses with capsids that exhibit T=4 icosahedral symmetry. They are classified into three families (Alphatetraviridae, Permutotetraviridae and Carmotetraviridae) based on the divergent characteristics of their viral replicase (REP). The Tetraviruses are considered to be amongst the simplest viruses known, encoding only the viral REP and capsid protein (CP) precursor, but recent studies show that this is no longer true. The genome of the alphatetravirus Helicoverpa armigera stunt virus (HaSV) comprises two RNAs: RNA2 encodes the CP and p17, a non-structural protein that is involved in RNA packaging. In addition to REP, RNA1 encodes three small ORFs, which overlap with the REP ORF at its 3' terminus, potentially producing p11, p15 and p8, all proteins of unknown function. These ORFs are arranged in tandem, separated by UGA (between p11 and p15) and UAA GGG (between p15 and p8). We investigated whether p11, p15 and p8 play a role in the viral life cycle. We report that the three ORFs are expressed in the midgut cells of HaSV-infected Helicoverpa armigera larvae as p11, p8 as well as a 34 kDa polyprotein comprising p11p15-p8. Analysis of the sequences surrounding the stop codons suggests that the polyprotein is likely expressed via a novel translation mechanism involving two read-through stop codons. EGFP-tagged p11 and p15 localize to punctate structures that co-localize with the viral REP in the cytoplasm of Spodoptera frugiperda (Sf9) cells. These punctate structures are associated with the Golgi apparatus and the vesicles of the secretory pathway. These data lead us to propose that p11, p15 and p8 play a role in the infectious viral lifecycle and likely in the initiation of viral replication in HaSV-infected cells.

Characterization of a Group I alphabaculovirus specific gene, ac5, from Autographa californica nucleopolyhedrovirus

STU 3:15

3:00 •22•

Xi Wang, Cheng Chen, Fenghua Zhang, Fei Deng, Hualin Wang, Zhihong Hu, Manli Wang

State Key Laboratory of Virology, Wuhan Institute of Virology, Chinese Academy of Sciences, Wuhan, PR China Corresponding author: huzh@wh.iov.cn: wanaml@wh.iov.cn

Autographa californica nucleopolyhedrovirus (AcMNPV) orf5 (ac5) is a Group I alphabaculovirus-specific gene with unknown function, although AC5 was reported previously to be associated with per os infectivity factor (PIF) complex. The purpose of this study is to characterize AC5 during AcMNPV infection. Transcription and expression analyses suggested that ac5 was a viral late gene. Western blot and immunoelectron microscopy showed that AC5 was an occlusion body (OB) structural protein, but was not detected as a component of budded virus (BV) or occlusion-derived virus (ODV). An ac5deleted recombinant AcMNPV was generated by homologous recombination to investigate the role of ac5 in virus life cycle. One step growth curve assay indicated that ac5 was not required for BV production and infection in Sf9 cells. Scanning electron microscopy and transmission electron microscopy demonstrated that the deletion of ac5 didn't affect OB morphogenesis and embedding of ODV into OB. Bioassay showed that the ac5-deleted virus was

2:45 •21•

still orally infectious to host larvae and the LC_{50} value was not significantly different from that of the ac5-repaired virus. Partial denatured SDS-PAGE and co-immunoprecipitation assay suggested that AC5 was not associated with PIF complex. In conclusion, AC5 is a structural protein of OB instead of a component of PIF complex, and is not essential for either BV- or ODV infection.

Roosevelt Room

Microsporidia Division #1

Moderator: TBD

Genome size in Microsporidia: An evolutionary hypothesis

Charles R. Vossbrinck, Bettina A. Vossbrinck, Jinshan Xu

Department of Environmental Sciences, The Connecticut Agricultural Experiment Station, New Haven, CT, USA Corresponding author: Charles Vossbrinck@ct.gov

It has long been suspected that the Microsporidia may, as obligate intracellular parasites, have small genomes because, for example, they could use Krebs cycle intermediates generated by the host. Genomic analysis has borne out the fact that microsporidia have small genomes. However the idea that an organism might have a small genome because it doesn't need a large genome is not an evolutionary explanation. In fact may organisms have genomes 10 to 100 times larger than the proteome. In addition studies comparing genome size versus phylogeny have shown that microsporidia genomes grow and shrink over evolutionary time. While we are aware that genetic drift could be responsible for the changing of microsporidial genome size we hypothesize that genome size may be a limiting factor in the rate of reproduction in the microsporidial genome. As such there would be a tradeoff between having a small genome and reproducing rapidly versus having a larger genome and giving the parasite more genetic options with which to challenge a host.

Genome, virulence factors, and specific molecular diagnosis STU 1:45 •24• of the microsporidian Enterocytozoon hepatopenaei (EHP)

Pattana Jaroenlak1,2, Piyachat Sanguanrut^{2,3}, Bryony A. P. Williams⁴, Dominic Wiredu-Boakye⁴, Grant D. Stentiford⁵, Timothy W. Flegel^{2,1} Kallaya Sritunyalucksana^{3,6}, Ornchuma Itsathitphaisarn^{1,2}

Department of Biochemistry, Faculty of Science, Mahidol University, Bangkok, Thailand; Center of Excellence for Shrimp Molecular Biology and Biotechnology (Centex Shrimp), Faculty of Science, Mahidol University, Bangkok, Thailand; Shrimp Pathogen Interaction Laboratory (SPI), National Center for Genetic Engineering and Biotechnology (BIOTEC), Bangkok, Thailand; Biosciences, College of Life and Environmental Sciences, University of Exeter, Exeter, United Kingdom; European Community Reference Laboratory for Crustacean Diseases, Center for Environment, Fisheries and Aquaculture Science (Cefas), Weymouth, Dorset, United Kingdom; National Center for Genetic Engineering and Biotechnology (BIOTEC), National Science and Technology Development Agency (NSTDA), Pathum Thani, Thailand Corresponding author: ornchuma.its@mahidol.ac.th

Hepatopancreatic microsporidiosis (HPM), a new emerging disease caused by Enterocytozoon hepatopenaei (EHP), has become a major issue in the Asian shrimp industry. EHP has been reported to cause slow growth in both black tiger shrimp (Penaeus monodon) and white-legged shrimp (P. vannamei) resulting in significant economic loss. Unfortunately, treatment against EHP infection has not been developed due to the lack of the insight into the parasite infection mechanism. Therefore, a genomic study of EHP was carried out to identify potential pathogenic genes of EHP. Among the virulent factors identified are spore wall proteins (SWP) and nucleotide transporters. The former has already been used to as a marker to improve the specificity of EHP diagnosis. To further characterize subcellular localization and function of SWP in spore attachment, antibody against SWP has been produced for immunofluorescent analysis. If the SWP is located in the exospore layer, it may provide a new marker for the development of a pond-side EHP diagnostic kit. In addition to SWP, this study will investigate the role of the nucleotide transporters of EHP in energy parasitism. Taken together, the insight from the functional characterization of the aforementioned proteins may lead to a pathway to control EHP infection.

First finding of a microsporidium of the family Tubulinosematidae infecting Drosophila suzukii

STU 2:00 •25•



Sarah Biganski, Johannes A. Jehle, Regina G. Kleespies

Julius Kühn – Institut, Federal Research Centre for Cultivated Plants, Heinrichstraße 243, 64287 Darmstadt, Germany Corresponding author: sarah.biganski@julius-kuehn.de

The demand of biological control agents against Drosophila suzukii (spotted wing drosophila, SWD) has become urgent around five years ago since the pest has been noticed as a major thread to commercially grown fruits in Europe. Microbial antagonists isolated from these invasive fly might be qualified as biological control agents to SWD. To find potential microbial antagonists for biological control we focused on specimens from countries of origin and early introduction like the USA, where natural control factors may be already established. In wild populations from different origins, microsporidia were detected. With one species infections of lab populations of SWD were successful. Molecular studies revealed a nearest relatedness with the family Tubulinosematidae. Blast results showed about 98% identity to Tubulinosema loxostegi with only 28 bp difference compared to the sequenced SSU-ITS-LSU region and phylogenetic reconstruction implies an assignment to the clade III microsporidia (Aquasporidia). Further gene regions will be sequenced. Morphological analyses by light and electron microscopy are conducted and will be compared to the molecular data. Bioassays in the lab with this microsporidium to investigate possible reduction of life span, fertility and fecundity of SWD are ongoing.

The new species, Nosema maddoxi is a widespread pathogen of the green stink bug Chinavia hilaris and the

brown marmorated stink bug Halyomorpha halys

Ann E. Hajek¹, Leellen F. Solter², James J. Becnel³

¹Dept. Entomology, Cornell University, Ithaca, New York, US, ²Illinois Natural History Survey, Prairie Research Institute, University of Illinois, Champaign, Illinois, US, ³Center for Medical, Agricultural and Veterinary Entomology -CMAVE (USDA, ARS). Gainesville, Florida, US Corresponding author: aeh4@cornell.edu

A new microsporidian species infects the green stink bug. Chinavia hilaris. the brown marmorated stink bug, Halyomorpha halys, the brown stink bug, Euschistus servus, and the dusky stink bug, Euschistus tristigmus. All life stages are unikaryotic, but analysis of the consensus small subunit region of the ribosomal gene places this microsporidium in the genus Nosema, which historically has been characterized by diplokaryotic life stages. It is also characterized by having the reversed arrangement of the ribosomal gene (LSU -ITS- SSU) found in species within the true Nosema clade. This microsporidium is apparently Holarctic in distribution. It is present in H. halys both where it is native in Asia and where it is invasive in North America, as well as in samples of North American native C. hilaris collected prior to the introduction of H. halys from Asia. Prevalence in H. halys from mid-Atlantic, North America in 2015-2016 ranged from 0.0-28.3%, while prevalence in C. hilaris collected in Illinois in 1970-1972 ranged from 14.3-58.8%. Oral infectivity and pathogenicity were confirmed in H. halys and C. hilaris. Morphological, ultrastructural and ecological features of the microsporidium, together with a molecular phylogeny, establish a new species named Nosema maddoxi sp. nov.

Experimental infection of caged honey bees and bumblebees with Nosema ceranae



Sebastian Gisder¹, Franziska Pieper¹, Vivian Schüler¹, Lennard Horchler¹, Peter Šima², Elke Genersch^{1,3}

¹Institute for Bee Research e.V. Friedrich-Engels-Str. 32, 16450 Hohen Neuendorf, Germany; ²Koppert s.r.o., Komárňanská cesta 13, 940 01 Nové Zámky, Slovakia: ³Freie Universität Berlin, Fachbereich Veterinärmedizin, Institut für Mikrobiologie und Tierseuchen, Robert-von-Ostertag-Str. 7-13, 14163 Berlin, Germany. Corresponding author: elke.genersch@hu-berlin.de 2:45

Marshall Room

Nematodes Division #1

Moderator: Gwen Stevens

Formulation of three species of entomopathogenic nematodes in alginate beads and diatomaceous earth

STU 1:30



N Kagimu¹, T Ferreira², AP Malan¹

¹Department of Conservation Ecology and Entomology, Faculty of AgriSciences, Stellenbosch University, Private Bag X1,Matieland 7602; ²Nemlab, c/o R44 and Anyswortelrug Road, Klapmuts 7625, South Africa Corresponding author: nkagimu@sun.ac.za

Costly and scarce formulation and application techniques are hindering the use of entomopathogenic nematodes (EPNs) as biological control agents in integrated pest management programs. For plausible solutions, three local South African species of entomopathogenic nematodes including; Steinernema yirgalemense, S. jeffreyense and Heterorhabditis bacteriophora using different formulation techniques were studied. Encapsulation of the infective juveniles (IJs) in alginate beads, in addition to the use of diatomaceous earth (DE) with reduced water activity (aw -value at 0.97) to induce quiescence; and to reduce the metabolism of the IJs were attempted. Survival of the IJs in the formulations was determined at 6°C, 14°C and 25°C for 4 weeks. It was observed that 10-20% of the IJs could escape from the beads, depending on temperature, and hence, readily survived the encapsulation process. Accordingly, desiccation effect of DE, on the survival of EPNs on the was pronounced at 6°C. Hence, in both formulations, survival significantly differed at 6°C, as compared to at 14°C and 25°C, with a drastic decrease over time for all species. The species exhibited poor survival at low temperature in either formulations. It should be noted that EPNs require low metabolism during storage and transportation. Thus, future research should investigate survival of these species between 8°C and 10°C for better results. The beads successfully retained most of the IJs, and, thus, can be stored for a longer time. Of the two methods studied, formulating EPNs in DE should be further investigated, due to ease of optimisation.

The microsporida *Nosema ceranae* has a unique Heat Shock Response system and is highly sensitive to proteotoxic stress

did not lead to a detectable N. ceranae infection of the midgut.

Managed colonies of A. mellifera and Bombus sp. are both widely used as

human food security. Therefore, diseases of bees (honey bees and

commercial pollinators in agriculture, thus, they considerably contribute to

bumblebees) are of concern not only for scientists but also for beekeepers, farmers, and consumers alike. Among the parasites and pathogens that

fungal-related, obligate intracellular parasites. Three microsporidian species are known to infect bees: *Nosema apis*, *N. bombi* and *N. ceranae*. While *N.*

apis is a specific parasite of A. mellifera, N. bombi is specific for Bombus sp..

for long to be specific for the Eastern honey bee (A. cerana), but extended its

via spore-contaminated fecal spots left on flowers by defecating forager bees.

In contrast, N. ceranae is a rather promiscuous parasite. It was considered

host range to A. mellifera probably in the last century. Recently, it was

The reported host-parasite range expansion of N. ceranae may pose a

serious threat to wild bees in general; however, productive infection of

results obtained from experimental infection of caged honey bees and

bumblebees with N. ceranae spores. Productive N. ceranae infection was

analyzed via PCR analysis of midgut extracts as well as via N. ceranae-specific

fluorescence in situ hybridization analysis of midgut sections. We show that

oral infection of caged naïve honey bees with N. ceranae spores resulted in

100% infected animals, whereas oral infection of caged naïve bumblebees

Bombus sp. by N. ceranae has not been proven so far. We here present our

suggested that *N. ceranae* also switched from *A. mellifera* to *Bombus* sp.. Infected *A. mellifera* colonies are the most likely indirect source of infection

cause disease in and death of bee colonies are microsporidia, which are

Mia McKinstry¹, Brittany Johnston², Jonathan W. Snow¹

¹Biology Department, Barnard College, New York, NY, 10027, USA ²Biology Department, The City College of New York – CUNY, New York, NY, 10031 *Corresponding author: jsnow@barnard.edu*

Nosema ceranae is a microsporidian parasite that is pathogenic to honey bees and can cause disease at the individual and colony level. While N. ceranae infection can currently be controlled by treatment with the drug Fumagillin. high doses of this drug are toxic to host cells and the potential for development of resistance suggests the need for alternative treatment strategies. Fumagillin, as a methionine aminopeptidase 2 inhibitor, works by interfering with protein synthesis and disrupting proteostasis. In addition to a pronounced sensitivity to Fumagillin, Nosema spp. exhibit an increased vulnerability to heat shock, a canonical trigger of proteotoxic stress. Thus, we hypothesized that a more complete understanding of proteostasis in this species, focusing on the Heat Shock Response (HSR), could help identify novel treatment strategies for microsporidia infection in honey bees. We confirmed that N. ceranae demonstrates increased sensitivity to heat stress relative to its host, the honey bee. In characterizing the HSR in N. ceranae, we found that this and other microsporidian species have lost the canonical heat shock transcriptional regulator HSF and possess a reduced core HSR pathway. Ongoing work aims to understand the apparently novel mechanism of HSR regulation in this species. We also found that N. ceranae cells display a striking sensitivity to pharmacological disruption of cytosolic proteostasis, suggesting the possibility of exploring such stress-inducing compounds as novel anti-Nosema chemical agents. Such discoveries, if applicable to microsporidian infections more broadly, may have widespread impact on microsporidia infection in food production and human health.

Influence of *Xenorhabdus* symbionts on ascaroside production and development of first-generation adults of their *Steinernema* hosts

STU 1:45

•29•

Alexandra Roder¹, S.P. Stock^{1,2}

¹School of Animal and Comparative Biomedical Sciences, University of Arizona, Tucson, AZ; ²Department of Entomology, University of Arizona, Tucson AZ Corresponding author: alexandra2310@email.arizona.edu

Entomopathogenic Steinernema nematodes have a mutualistic relationship with Xenorhabdus bacteria. The two partners form an insecticidal alliance that is successful in killing a wide range of insects. A few studies have shown that Steinernema IJs have an enhanced virulence and reproductive fitness when they associate with their cognate symbionts. However, there are unanswered questions regarding the physiological interactions that govern and perpetuate the interactions between different nematode developmental stages and their bacterial partners. Hence, in this study, we evaluated gonad development and maturation time of 1st-generation S. carpocapsae adults when reared under four bacterial scenarios: a) cognate-symbiotic bacteria, b) closely related non-cognate-symbiotic bacteria, c) distantly related noncognate-symbiotic bacteria and d) non-symbiotic bacteria. Furthermore, we also measured production of signaling pheromones (ascarosides) that are known to play a key role in mating and reproduction. For this purpose, we considered in vitro rearing methods (with/without X. nematophila symbiont) to quali- and quantitatively characterize signaling pheromone molecules produced by first generation adults. Our data showed that gonad development and adult maturation time are tightly dependent on the presence of the nematodes' cognate symbiont. Ascaroside analysis also revealed quantitative diversity in the ascarosides produced by S. carpocapsae adults in relation to the different bacteria culture conditions.

Activated entomopathogenic nematode infective juveniles release lethal venom proteins

2:00 •30•

Dihong Lu¹, Marissa Macchietto², Dennis Chang¹, Mirayana M. Barros¹, James Baldwin¹, Ali Mortazavi², Adler R. Dillman¹

> ¹Department of Nematology, University of California, Riverside, California, USA; ²Department of Developmental and Cell Biology, Center for Complex Biological Systems. University of California, Irvine, California, USA Corresponding author: adlerd@ucr.edu

Entomopathogenic nematodes (EPNs) are unique parasites due to their symbiosis with entomopathogenic bacteria and their ability to kill insect hosts quickly after infection. It is widely believed that EPNs rely on their bacterial partners for killing hosts. Here we disproved this theory by demonstrating that the in vitro activated infective juveniles (IJs) of Steinernema carpocapsae (a well-studied EPN species) release venom proteins that are lethal to several insects including Drosophila melanogaster. We confirmed that the *in vitro* activation is a good approximation of the *in* vivo process by comparing the transcriptomes of individual in vitro and in vivo activated IJs. We further analyzed the transcriptomes of non-activated and activated IJs and revealed a dramatic shift in gene expression during IJ activation. We also analyzed the venom proteome using mass spectrometry. Among the 472 venom proteins, proteases and protease inhibitors are especially abundant; and toxin-related proteins such as Shk domaincontaining proteins and fatty acid- and retinol-binding proteins are also detected, which are potential candidates for suppressing the host immune system. Many of the venom proteins have conserved orthologs in vertebrateparasitic nematodes and are differentially expressed during IJ activation, suggesting conserved functions in nematode parasitism. In summary, our findings strongly support a new model that S. carpocapsae and likely other Steinernema EPNs have a more active role in contributing to the pathogenicity of the nematode-bacterium complex than simply relying on their symbiotic bacteria. Furthermore, we propose that EPNs are a good model system for investigating vertebrate- and human-parasitic nematodes, especially regarding the function of excretory/secretory products.

Cross-talk between nitric oxide and eicosanoid: a specific inhibition of Xenorhabdus hominickii metabolite

2:15 •31•



Yonggyun Kim

Department of Plant Medicals, Andong National University, Andong 36729, Korea Corresponding author: hosanna@anu.ac.kr

Upon nonself recognition, biochemical mediators act in signaling insect immune functions. These include biogenic amines, insect cytokines, eicosanoids and nitric oxide (NO). Treating insects or isolated hemocyte populations with different mediators often leads to similar results. Separate treatments with an insect cytokine, two biogenic amines and an eicosanoid lead to a single result, hemocyte spreading, understood in terms of intracellular cross-talk among these signaling systems. This study focuses on cross-talk between NO and eicosanoid signaling in our model insect, Spodoptera exigua. Bacterial injection increased NO concentrations in larval hemocytes and fat body and RNA interference (RNAi) of SeNOS suppressed NO concentrations. RNAi-treatment also led to a significant reduction in hemocyte nodulation following bacterial injection. Similar RNAi treatments led to significantly reduced PLA2 activities in hemocytes and fat body compared to control larvae. Injection of L-NAME also prevented the induction of PLA₂ activity following bacterial challenge. An injected NO donor, S-nitroso-N-acetyl-DL-penicillamine, increased PLA2 activity in a dosedependent manner. An entomopathogenic bacterium, Xenorhabdus hominickii, inhibited PLA2 activity, but did not suppress NO production. We infer that NO and eicosanoid signaling operate via cross-talk mechanisms in which the elevated NO concentrations activate PLA₂ and eicosanoid biosynthesis, which finally mediates various immune responses.

Root-feeding insects adapt their behavior to bypass plant inducible defenses and attraction of entomopathogenic nematodes

Ivan Hiltpold

¹Department of Entomology and Wild Life Ecology, University of Delaware, Newark, DE Corresponding author: Hiltpold@udel.edu

Living belowground is tough; soil-dwelling insect pest constantly face rough dense soil material, wearing their cuticles out, rendering any movement a challenge, and putting them in constant contact with microorganisms which cannot be always considered allies. If herbivores are lucky enough to find roots to sustain them, they then have to face an arsenal of plant defenses in order to turn chewy roots into biomass. Plant defenses can be characterized into two broad categories: constitutive and induced. Constitutive defenses are mechanical barriers or metabolites always present on or in the plant tissues whereas induced defenses are triggered only in presence of the threatening herbivore. Despite plants investing in root defenses, it seems common for root herbivores to thrive and eventually overcome their host plants. Is it possible that root herbivorous insects circumvent inducible root defenses? Here we show for the first time that root herbivores may clip part of the roots, disconnecting them from the main root system before feeding on the detached root material. We hypothesize that this behavior prevents the build-up of inducible defenses (emission of volatiles to attract entomopathogenic nematodes) which relies on primary metabolites and energy from the main root system. In addition, clipped roots could be left vulnerable to detritivores microbiota which could assist in the detoxification of constitutive defenses.

Susceptibility of invasive gastropods and non-targets to different Phasmarhabditis species in the US

Irma Tandingan De Ley¹, Rory J. McDonnell², Timothy D. Paine³,

¹Department of Nematology, University of California, Riverside, CA 92521, USA; ²Department of Crop and Soil Science, Oregon State University, Corvallis, OR 97331; ³Department of Entomology, University of California, Riverside, CA, 92521, USA Corresponding author: irma.deley@ucr.edu

Three Phasmarhabditis spp including P. hermaphrodita were recently discovered in the US, and tests were conducted to evaluate their lethal effects on invasive slugs, snails and non-targets. Invasive gastropods showed different levels of susceptibility to the three Phasmarhabditis species. P. hermaphrodita caused significant to highly significant mortality on Lehmannia valentiana (Valencia slug), Deroceras reticulatum (grey field slug), Cornu aspersum (European brown garden snail) and Lissachatina fulica (giant African land snail) neonates at rates equivalent to or higher than Nemaslug®'s recommendation of 30 IJs/sq cm (1x). It also infects Limacus flavus (Tawny garden slug) but does not cause mortality and is safe to three earthworm species Amynthas gracilis (Alabama jumper), Eisenia fetida (Red wigglers), and Eisenia hortensis (European night crawler). L. valentiana is more susceptible to P. papillosa than P. hermaphrodita and P. californica. However, a month after exposure, all three killed more than 50% of the pest slugs with P. papillosa and P. californica showing more virulence than P. hermaphrodita. Exposure to the three Phasmarhabditis species at 1x or 5x recommended rate is safe to non-target species Rumina decollata. L. valentiana, L. flavus, and L. fulica are new host records for Phasmarhabditis.

Improved RNAi for Brugia malayi parasitic nematodes in Aedes aegypti





Paul M Airs¹, Watcharatip Dedkhad², Mostafa Zamanian¹, Michael J. Kimber³, Lyric C. Bartholomay

¹University of Wisconsin Madison, Department of Pathobiological Sciences, USA: ²University of Chiang Mai, Department of Parasitology, Thailand; ³Iowa State University, Department of Biomedical Sciences, USA Corresponding author: airs@wisc.edu

The parasitic nematode, Brugia malayi, exacts a significant burden on human health but remains difficult to study using RNA interference (RNAi) due to limited viability of larval stages in culture and low RNAi sensitivity. In Caenorhabditis elegans, suppression of anti-RNAi eri-1 and rrf-3 genes dramatically increases RNAi knockdown success; however the function of B.

malayi homologs is not known. In RNAi hypersensitive backgrounds study of *C. elegans* essential genes have been highlighted as potential drug targets for *B. malayi*. To validate 'essential' gene knockdown impact in *B. malayi* larvae we must first improve RNAi efficacy during larval development. To address this we utilize a previously pioneered 'in squito' approach that exposes *B. malayi* infected *Aedes aegypti* to RNAi triggers via direct haemolymph injection. Using fluorescence microscopy and tailored Northern blotting techniques we track the bio-distribution and persistence of RNAi triggers through larval life-stages of *B. malayi*. Additionally, we show life-stage expression profiles of *Bma-eri-1*, *Bma-rrf-3*, and select essential *B. malayi*. Finally, through co-suppression of Bma-eri-1 or Bma-rrf-3 and essential genes we assess the possibility for RNAi to induce desirable phenotypes that can be utilized in controlling filarial worms as well as optimizing RNAi practices

An enterotoxin-like binary protein from *Pseudomonas* protegens with potent nematicidal activity

3:15 •35•

Jun-Zhi Wei¹, Daniel L. Siehl¹, Barbara Rosen¹, Jarred Oral¹, Christopher G. Taylor², Gusui Wu³

¹DuPont Pioneer, Hayward, California, USA; ²Department of Plant Pathology,
Ohio State University, Columbus, Ohio, USA; ³DuPont Pioneer, Johnston, Iowa, USA

Corresponding author: jun-zhi.wei@pioneer.com

Soil microbes are a major food source for free-living soil nematodes. It is known that certain soil bacteria have evolved systems to combat predation. We identified the nematode-antagonistic Pseudomonas protegens strain 15G2 from screening of microbes. Through protein purification we identified a binary protein, designated as Pp-ANP, that is responsible for the nematicidal activity. This binary protein inhibits Caenorhabditis elegans growth and development by arresting larvae at L1 stage and killing older staged worms. The two subunits, Pp-ANP1a and Pp-ANP2a are active when reconstituted from separate expression in Escherichia coli. The binary toxin also shows strong activity on three other free-living nematodes, Pristionchus pacificus, Panagrellus redivivus, and Acrobeloides sp., but no activity on insects and fungi under test conditions, indicating specificity for nematodes. Pp-ANP1a has no significant homology to any known proteins, while Pp-ANP2a shows 30% homology to E. coli heat-labile enterotoxin subunit A (LT-A) and cholera enterotoxin subunit A (CT-A). Despite sequence similarity, Pp-ANP2a shows several characteristics distinct from enterotoxins. Namely, the A-like subunit has no C-terminal ER-retention signal or cleavage site and the two conserved cysteines are in different positions. Our results indicate that Pp-ANP is a potent toxin, specific to nematodes, that appears to be analogous to yet distinct from previously known AB5 enterotoxins. The potency and specificity of Pp-ANP suggests applications in controlling parasitic nematodes and opens an avenue for further research on its mechanism of action and role in bacterium-nematode interaction.

3:30–4:00 pm Refreshments Break

MONDAY, 4:00 – 6:00 pm

Theater

VIRUS DIVISION SYMPOSIUM

Insect virology: Historical achievements and recent advances

Organizer and Moderator: Elisabeth Herniou

The development and impact (or impacts) of the baculovirus expression system

4:00

0 •36

Robert D Possee^{1,2}

¹Department of Biological and Molecular Sciences, Oxford Brookes University, Oxford UK; ²Oxford Expression Technologies, Oxford UK Corresponding author: rdpossee@brookes.ac.uk

Baculovirus expression vectors have developed as an important tool for the production of recombinant proteins over the past 30+ years. From uncertain beginnings where non-specialists struggled with an arcane polyhedrinnegative virus selection method, they have progressed to a point where very little knowledge of virology is required to use them successfully. In parallel with the improvements in recombinant virus selection systems insect cell culture methods have also advanced. Cloned cell lines are available that can be propagated to high density in serum-free media to provide high concentrations of virus-infected cells for recombinant protein extraction. The number of examples of different recombinant proteins made using the baculovirus expression system continues to grow. The vectors are extremely flexible and can be used to make both simple and complicated recombinant products alike. This is demonstrated admirably by their use to produce multicomponent virus-like particles or enzyme complexes. Frequently, baculoviruses will save the day when all other vector systems have failed. Therefore, much is owed to those pioneers of insect virology who performed the early biological and molecular studies on baculoviruses that resulted in the expression system we have today. However, perhaps is now the time to take a pragmatic look at the impact that baculovirus vectors have made to society in general. Have they had a positive benefit to the health and well being of the wider population? As scientists, we may rejoice in the way they have bolstered our publication lists and curriculum vitae, but beyond our narrow discipline do baculovirus vectors really matter? The introduction of clinical products in healthcare suggests that we are on the right track but they are few in number. We must ask what the future for baculovirus expression vectors is in the next 30 years. What would a similar talk at SIP2047 report?

Taking the next step: from bugs to cells

4:24

24 **937**

Taro Ohkawa¹, Loy E. Volkman², Matthew D. Welch

¹Department of Molecular and Cell Biology, University of California, Berkeley, California, U. S. A.; ²Department of Plant and Microbial Biology, University of California, Berkeley, California, U. S. A. *Corresponding author: tohkawa@berkeley.edu*

The study of molecular insect virology has largely focused on the virus particle. Structural studies on virus morphology moving into molecular studies of viral genes and gene expression. Effects on the host have been studied in the pathology of disease in insects, virus spread and host killing. But only fairly recently have insect host-pathogen interactions focused more finely on the level of virus-cell interactions: the crossroads of virology and cell biology. In the late 1980s a link between baculovirus infection and the host cytoskeleton was discovered. And further examination revealed the great extent to which a virus can appropriate and manipulate the cytoskeleton of the cell. These findings were significant in that we can gain important knowledge about both how a virus can hijack host systems for their own ends, as well as how cells regulate their own cytoskeletal operations. This wedding of cell biology and insect virology has led to the discovery of many unexpected phenomena that occur during virus infection. at the cellular level. My studies have focused on the baculovirus AcMNPV, which has been shown to manipulate host actin at numerous steps during the viral life cycle. AcMNPV uses actin-based motility, polymerizing actin

'comet' tails for propulsion through the cell cytoplasm following entry into the cells. Furthermore, AcMNPV has been demonstrated to mediate the translocation of actin into the nucleus, where it accumulates and polymerizes during the late stage of infection. After virus nucleocapsids are assembled, they initiate intranuclear actin-based motility, which is critical for egress from the nucleus, involving the formation of protrusions of the nuclear envelope. Nucleocapsids escape from the nucleus following a disruption of nuclear envelope integrity, and utilize cytoplasmic actin-based motility to transit to the plasma membrane. Studying the mechanisms of this viral manipulation of cellular actin will shed light both on viral appropriation of cellular processes, as well as how these processes may be regulated in non-infected

Bee viruses and honey bee health

4:48 •38•

Laura Brutscher^{1,2,3}, Alex McMenamin^{1,2,3}, Katie Daughenbaugh^{1,3}, **Michelle Flenniken**^{1,3} ¹Department of Plant Sciences and Plant Pathology; ²Department of Microbiology; ³Pollinator Health Center, Montana State University, Bozeman, MT, USA Corresponding author: michelle.flenniken@montana.edu

Bees are important pollinators of numerous crops and plant species that enhance the biodiversity of both agricultural and non-agricultural landscapes. Unfortunately, bee populations are declining in many regions of the world. Many factors negatively impact bee health including reduced habitat, lack of quality forage, climate change, and pesticide-exposure, and pathogens. Bees are infected by a variety of pathogens (i.e., viruses, bacteria, microsporidia, trypanosomatids, and the Varroa destructor) and honey bee colony losses in North American and Europe are associated with elevated pathogen incidence and abundance. The largest group of bee pathogens is made up of positive sense, single-stranded RNA viruses, thus understanding virus pathogenicity, inter-species transmission, and bee antiviral defense mechanisms are important areas of investigation that may result in the development of strategies that limit bee losses associated with virus infections. Michelle Flenniken will summarize the current understanding of the impact of viruses on bee health, interspecies transmission, and bee antiviral defense mechanisms with a particular focus on recent results in the field.

Antiviral immunity in insects: the impact of RNA interference and Wolbachia.

5:12 •39•

Karyn N. Johnson

School of Biological Sciences, The University of Queensland, 4072, Australia Corresponding author: karvni@ua.edu.au

The interaction between a virus and its insect host can have important consequences for disease transmission. Understanding these interactions may facilitate development of approaches to control insect vectored viral diseases of both plants and animals. Drosophila has emerged as an important model to understand the host-virus interaction. In insects, the outcome of virus infection is greatly impacted by the insect innate immune response, which includes both inducible responses and RNAi. Arguably the most robust immune response in insects is the RNAi response. The presence of doublestranded RNA generated during virus replication stimulates the siRNA pathway leading to the degradation of virus RNA. In response, many insect viruses encode proteins that suppress the RNAi machinery, indicating coevolution of the host and virus. In addition to the host antiviral mechanisms, tripartite interactions between viruses, bacteria and hosts can have significant implications for the outcome of virus infection. The endosymbiotic bacterium Wolbachia, which is present in an estimated 40% of all insect species, has the ability to alter viral dynamics in both Drosophila and mosquitoes, a feature that in mosquitoes may be utilised to limit the spread of important arboviruses.

What dawns at the horizon for invertebrate pathology?



Monique M. van Oers, Vera I.D. Ros

Laboratory of Virology, Wageningen University, Droevendaalsesteeg 1, 6708 PB, Wageningen, The Netherlands Corresponding author: monique.vanoers@wur.nl

In recent years, we see a shift in invertebrate virology from functional viral genomics to an integrative analysis of virus-host interactions. Current

applications are extending from biotechnology and biocontrol of pest insects to prevention of viral scourges in pollinators, aquatic invertebrate farming, and insect rearing for food and feed; all areas in which virus outbreaks can be devastating. Additional virological risks arise from invading invertebrates through global trade and travel, and through climate change. Next generation sequencing approaches are revealing a wealth of new viral sequences, underscoring the large diversity of invertebrate viruses and opening up untapped potential of viruses awaiting to be discovered. The challenge is to understand the biological meaning of all the 'omics' data being generated, starting by asking the relevant biological questions. Considering the large number of apparently orphan viruses, for which accompanying diseases are not known, virus latency may be more a rule than an exception, forcing us to adjust our perspective of viruses. Instead of being merely pathogenic entities they will turn out to be important ecological players. Only by combining disciplines, such as genetics, cell biology, synthetic biology, physiology and ecology, we will be able to fully comprehend how viral infections affect invertebrate hosts as well as other biological entities that rely on these hosts, e.g., co-infecting pathogens or mutualist organisms, or other organisms that are part of various multitrophic interactions. Some of the complex processes still to be understood, such as virus latency and persistency, might be exploited to control pathogenic viruses in the future.

The Forum

Bacteria Division #1

Moderators: Baltasar Escriche, Juan Luis Jurat-Fuentes

Understanding the basis of Cry2A toxin specificity towards Aedes aegypti



STU 4:00 •41•

Lazarus Joseph, Elmi Dahiru, Thomas Courty, Neil Crickmore.

School of Life Sciences, University of Sussex. Falmer, Brighton BN1 9QG, United Kingdom. Corresponding author: L.Joseph@sussex.ac.uk

High specificity is one of the important attributes of Bacillus thuringiensis insecticidal Cry proteins. Therefore, understanding the basis of specificity through determining regions of specificity and molecular interactions of the toxins, is essential not only for the development of novel insecticides but also for the assessment of their risk to non-target organisms. This study involved determining the activity of various Cry2A toxins, and in vitro created hybrids, towards the mosquito A. aegypti, combined with bioinformatic analyses to define regions that determine or influence activity towards this pest. Previous studies have identified a D-block within domain II that is believed to influence activity towards A. aegypti. We have identified an alternative region within domain I and found that substituting just four amino acids within this region can convert Cry2Ab, which is inactive against

Importance of Domain III in Cry1Ac for toxicity against the soybean looper (Chrysodeixis includens)



Rubina Mushtaq¹, Abdul Rauf Shakoori¹, **Juan Luis Jurat-Fuentes**²

¹School of Biological Sciences, University of the Punjab, Lahore, Pakistan; ²Department of Entomology and Plant Pathology, University of Tennessee, Knoxville, USA Corresponding author: jurat@utk.edu

The Cry1Ac insecticidal protein from the bacterium Bacillus thuringiensis (Bt) is produced by transgenic soybeans to control larvae of the soybean looper (Chrysodeixis includens). In this three-domain protein, domain III is critical for binding to protein receptors in the insect midgut. In this study, we performed substitution of domain III of Cry1Ac with the same domain in Cry2Ac, a toxin also active against C. includens larvae. Bioassays comparing parental and hybrid proteins showed lack of activity against C. includens. Binding assays with radiolabeled toxins demonstrated lack of hybrid toxin binding to brush border membrane vesicles from C. includens. In contrast, significant toxicity and binding were detected for the hybrid toxin against larvae of another soybean pest, the velvetbean caterpillar (Anticarsia gemmatalis). Taken together, these results support that domain III is critical for Cry1Ac receptor binding in C. includens and suggest different critical regions for toxicity against A. gemmatalis.

Study of Bacillus thuringiensis Cry1Ab and Cry1Ac protoxins interaction with cadherin-like receptor and GPI-anchored proteins extract from Manduca sexta

4:30 •43•

Arlen Peña-Cardeña, Alejandra Bravo, Mario Soberón, Isabel Gómez

Departamento de Microbiología Molecular, Instituto de Biotecnología, Universidad Nacional Autónoma de México Corresponding author: isabela@ibt.unam.mx

Bacillus thuringiensis (Bt) produces insecticidal Cry toxins. To induce the activity, the crystals have to be ingested by susceptible larva, solubilized in the gut and activated by midgut proteases to yield an activated toxin. In the case of Cry1A, toxins undergo a sequential binding mechanism with glycosylphosphatidylinositol (GPI) anchored proteins such as alkaline phosphatase (ALP) or aminopeptidase-N (APN), and cadherin-like protein resulting in the formation of a prepore oligomeric structure and its insertion in membrane. The currently accepted paradigm asserts that only activated toxins can bind to larval midgut receptors and exert toxic effects. However, previously we reported that Cry1Ab protoxin or activated toxin bind cadherin-like receptor with similar affinities and two different prepores are produced. Here we test the interaction of Cry1Ab and Cry1Ac protoxins with a extract of GPI-anchored proteins obtained from midgut of M. sexta. Our results showed that Crv1Ab protoxin binds to the GPI-anchored proteins extract with the same affinity than activated toxin suggesting that, in vivo, both would be able to bind to these receptors. In contrast, Cry1Ac protoxin has no interaction with cadherin-like receptor neither GPI-anchored extract, suggesting that another protein may act as receptor for Cry1Ac protoxin that is important to identify.

Toxicity of Cry1A toxins to CF1 cells does not involve activation of adenylate cyclase/PKA signaling pathway

Leivi Portugal, Mario Soberón, Alejandra Bravo

Instituto de Biotecnología, Universidad Nacional Autonoma de México, Mexico Corresponding author: bravo@ibt.unam.mx

Here we analyzed the mechanism of action of Cry1Ab and Cry1Ac toxins and a collection of mutants from these toxins in the insect cell line CF-1, from the lepidopteran Choristoneura fumiferana, that is naturally sensitive to these toxins. We show that different non-toxic mutant toxins affected in receptor binding, oligomerization or pore formation lost also toxicity against CF1 cells. Our results show that both Cry1Ab and Cry1Ac toxins induced permeability of K⁺ ions into the cells. The initial response involves the activation of a defense response that involves the phosphorylation of MAPK p38. Analysis of activation of PKA and AC activities indicated that the signal transduction involving PKA, AC and cAMP was not activated during Cry1Ab or Cry1Ac intoxication. In contrast we show that Cry1Ab and Cry1Ac activated apoptosis. These data indicate that Cry1A toxins in CF1 cells can induce an apoptotic death response triggered by pore-formation not related with AC/PKA activation. Our results also show that defense responses triggered by Cry1A toxin activating phosphorilation of MAPK p38 is conserved in different insect species and in CF1 cell line, in contrast to PKA death response that has only been observed in another cell line, Trichoplusia ni H5, transfected with cadherin receptor from Manduca sexta.

Utility of Cry1Ja for transgenic insect control: Competitive binding and in planta assays in three lepidopteran species





John Mathis, Catherine Finke, Gilda Rauscher, Mark Nelson, Gusui Wu

DuPont Pioneer, Johnston, IA 50131 USA Corresponding author: john.mathis@pioneer.com

Insect control traits are a key component to improving efficiency and maximizing crop yields for growers. Insect traits based on proteins expressed by the bacteria Bacillus thuringiensis (Bt) have proven to be very effective tools in achieving this goal. The adaptability of insects has led to resistance to certain proteins in current commercial products. Therefore, new insecticidal traits that represent a different mode of action (MoA) than those currently in use are needed. Cry1Ja has good insecticidal activity against various lepidopteran species and we find that it provides robust protection against insect feeding with in planta expression. For Bt, different MoAs are determined by their binding sites in the insect midgut. We performed

competitive binding assays using brush border membrane vesicles from Helicoverpa zea, Spodoptera frugiperda and Chrysodeixis includens to evaluate the MoA of Cry1Ja relative to the classes of Bt proteins that are expressed in current commercial products. These data illustrate the potential for Cry1Ja for new insect trait development.

Structures of Bombyx mori ABCC transporters responsible for mediating Cry1A toxin intoxication





Haruka Endo, Shiho Tanaka, Ryoichi Sato

Corresponding author: osca1990@gmail.com

Graduate School of Bio-Applications and Systems Engineering. Tokyo University of Agriculture and Technology, Koganei, Tokyo, Japan

The ABC transporter C2 (ABCC2) in lepidopteran insects is a powerful receptor for Bacillus thuringiensis Cry1A toxins due to the outstanding activity that directly mediates pore formation. In the present study, we screened regions of the two Bombyx mori ABCC transporters BmABCC2 and its paralog BmABCC3 that affect receptor function for Cry1A toxins. Analysis using BmABCC2 variants having a mutation in either of six extracellular loops (ECLs) revealed that the structure comprised of ECL 1 and 4 is the putative interaction site with Crv1A toxins. Next, we generated BmABCC2 variants lacking transport activity based on the hypothesis that the powerful receptor function is derived from transport activity or following conformational change. Results indicated that the transport activity is not essential for receptor function. Finally, we sought regions generating different Cry1A receptor activities between BmABCC2 and BmABCC3. BmABCC3 mutants, whose ECL 1 or 3 are partially substituted with those of BmABCC2 increased and gained receptor activities for Cry1Aa and Cry1Ab, respectively. These results indicate that ABCC2 is a just binding target of Cry1A toxins as well as other Crv1A receptors and that structure of ECLs influences Crv toxin specificity and ABCC-mediated toxicity levels.

Participation of different regions of Cyt1Aa toxin from Bacillus thuringiensis in hemolysis, synergism and toxicity to Aedes aegypti larvae.





Mary Carmen Torres-Quintero¹, Jazmin A. López-Diaz¹, Sarjeet S. Gill², Mario Soberón¹, Alejandra Bravo¹.

¹ Instituto de Biotecnología, Universidad Nacional Autónoma de México. Apdo. Postal 510-3, Cuernavaca 62250, Morelos (Mexico); ² Cell Biology and Neuroscience Department, University of California, Riverside, Riverside, CA, 92521, USA. Corresponding author: mara@ibt.unam.mx

Cyt toxins from Bacillus thuringiensis display insecticidal activity principally against dipteran larvae. However, these toxins also show hemolytic activity. Cyt1Aa is pore-forming toxin, which plays an important role in mosquito control since it synergizes the activity of Cry toxins resulting in a potent activity against mosquitoes. In addition, it was shown that Cyt toxin is able to overcome the resistance developed by Culex guinguefaciatus to Cry toxins. Cyt toxins do not interact with protein-receptors but bind to specific lipids found in apical membrane of the midgut cells leading to oligomerization, membrane insertion and pore formation. Helices $\alpha\text{-A}$ and $\alpha\text{-C}$ from Cyt1Aa had been proposed to be involved in initial membrane binding and oligomerization. To determine the role of Cyt1Aa α-helices different residues were mutated and characterized regarging to their effects on oligomerization, hemolysis and synergism with Cry11A and toxicity. Some helix α -C mutations severely affacted oligomerization, hemolytic activity and toxicity to Aedes aegypti larvae. However these mutants were still able to synergize Cry11Aa toxicity indicating that toxicity of Cyt1Aa and synergism of Cry11Aa are independent activities. Some mutants in helix α -A are affected hemolysis but not in toxicity to A. aegypti larvae indicating that insecticidal and hemolytic activities of Cyt1Aa are also independent events.

A Cry1B protein showing high activity against Helicoverpa zea

5:45 •48•

Michi Izumi Willcoxon, Kishore Kakani, Ruth Cong, Sabina Lau, Yi Zheng, Jingtong Hou, Zhenglin Hou, Takashi Yamamoto

DuPont Pioneer, Ag Biotechnology, Plant Protection, Hayward, CA 94545, USA Corresponding author: naga.kakani@pioneer.com

Cry1B is a family of Bacillus thuringiensis (Bt) three-domain type insecticidal proteins having diversified Domain III. For example, Cry1Bd has a Cry1Ac/htype Domain III while Cry1Be has a Cry1Cb-type Domain III. Previously, we reported the discovery of a cry gene fragment consisting of sequences encoding a Cry1Ac-type Domain III and the protoxin region in a Bt isolate containing cry1B-type genes (Yamamoto, et al. Pac. Rim Conference on the Biotechnol. of B. thuringiensis, 2005). This indicates that Bt is naturally capable of swapping Domain III by homologous recombination. Since Cry1Ac has high specificity against the Heliothis/Helicoverpa complex, it seems Bt is trying to acquire a new specificity to this insect complex by combining the Cry1B Domain II with a Cry1Ac-type Domain III. However, H. zea activity of a Cry1B protein, Cry1Bd, which has a Cry1Ac-type Domain III, was not as high as Cry1Ac. Recently, Abad et al. (KT952325) reported a new Cry1B protein, Cry1Bj, from a naturally occurring Bt. Cry1Bj has a Cry1Ac-type Domain III similar to Cry1Bd, but H. zea activity of Cry1Bj was exceptionally high. Certain chimeric proteins made between Cry1Bd and Cry1Bj showed that the unique Domain I of Cry1Bj is important for high H. zea activity of that protein. This finding was confirmed by single amino acid swapping between Cry1Bd and Cry1Bj. The activity of Cry1Bj was further improved by saturation mutagenesis followed by high throughput insect screening.

Engineering of an insecticidal protein specific to corn rootworm



Ruth Cong, Jingtong Hou, Ericka Bermudez, Hana Ali, Erica Corson, Katie Harding, Zhenglin Hou, Phil Patten, Gusui Wu, Mike Lassner, Takashi Yamamoto

> DuPont Pioneer, Ag Biotechnology, Plant Protection, Hayward, CA 94545, USA Corresponding author: takashi.yamamoto@pioneer.com

A perforin-like protein containing a MACPF domain and a DUF946 domain from Chromobacterium piscinae was found to be active against western corn rootworm (Diabrotica virgifera virgifera) (Sampson et al., 2016, J. Invertebr. Path., 142, 34-43). We improved the activity of a native perforin-like protein by saturation mutagenesis. We focused primarily on making certain mutations to hydrophilic amino acid residues. The site-specific, limited mutagenesis identified a number of sites which appeared to be important for the insecticidal activity of the protein. We subjected these sites to full saturation mutagenesis to identify additional mutations for further activity improvements. Single amino acid mutations with substantial activity increase were subsequently combined to further improve activity. Over 300 different combinations were found to have at least 5-fold activity improvement over the original wild type protein. Several of these combinations showed levels of activity increase of more than 30 folds.

Roosevelt Room

Fungi Division #1

Moderators: Louela Castrillo, Jørgen Eilenberg

A new insect pathogenic fungus from Entomophthorales with potential for psyllid control



Annette Hjorthøj Jensen^{1,2}, Louisa Görg¹, Jürgen Gross¹, Anant Patel³, Annette Bruun Jensen², **Jørgen Eilenberg**²

¹Julius Kühn-Institut, Institut für Pflanzenschutz in Obst- und Weinbau, Schwabenheimer Str. 101, 69221 Dossenheim, Germany; ²University of Copenhagen, Department of Plant and Environmental Sciences, Thorvaldsensvej 40, 1871 Frederiksberg C, Denmark; ³Universität Bielefeld, Technische Fakultät, Universitätsstr. 25. D-33615 Bielefeld Corresponding authors: zgv470@alumni.ku.dk and jei@plen.ku.dk

Psyllid pests in fruit orchards can have major economic consequences as they often cause direct and indirect damage on the host trees. Obligate entomopathogenic fungal species from the order Entomophthorales normally have a narrow host range and can cause natural epidemics in insect

populations. We investigated the potential of entomophthoralean fungi for biological control of psyllids in fruit orchards. During late summer in 2016, live psyllids were collected from both apple and pear trees in North Zealand, Denmark, in order to discover natural infections by entomopathogenic fungi. The collected psyllids (approximate 4000 individuals) were incubated alive in the laboratory, and observed daily for Entomophthorales infection. Infection was found and documented for the first time in Cacopsylla spp. collected from pear trees. The fungus was successfully isolated and cultivated in vitro from four sporulating cadavers on Sabouraud dextrose agar-medium (SDA) supplemented with egg yolk and milk using the 'ascending conidia' showering method. Based on morphological characters of the conidia and DNA sequences of the ITS-region, the fungus proved to be a species from genus Pandora (Entomophthorales: Entomophthoraceae). The pathogenicity of the fungus was examined in the laboratory of Julius Kühn-Institut (Dossenheim, Germany) on three different psyllid species (Cacopsylla pyri, C. pyricola and C. mali). The results showed that the isolated Pandora sp. was able to infect all species of psyllids. Our preliminary results indicate the potential of Pandora sp. as a biological control agent of psyllid pests in fruit orchards.

Interaction of entomopathogenic fungus Beauveria STU 4:15 0510 bassiana and predatory mite Amblyseius swirskii on their shared prey, western flower thrips Frankliniella occidentalis

> Marjan Heidarian Dehkordi¹, Hossein Allahyari¹, Reza Talaei Hassanlouei¹, Bruce L. Parker²

¹ Department of Plant Protection, College of Agriculture and Natural Resources, University of Tehran, Karaj, Iran; ² Entomology Research Laboratory, University of Vermont, Burlington, VT 05405-0105, USA Corresponding author: marjan.heidarian@gmail.com

For successful biological control programs, knowing the interaction between biocontrol agents, such as competition for the shared prey in an intraguild predation system, is necessary. Towards this end, bioagents including a commercial Beauveria bassiana (Bals.) Vuill. (Ascomycota, Cordycipitaceae) (strain GHA) and an experimental isolate, JEF-007 and the predatory mite Amblyseius swirskii Athias-Henriot (Acari: Phytoseiidae) were evaluated in single and combined applications for controlling western flower thrips (WFT), Frankliniella occidentalis (Thysanoptera: Thripidae), on marigold. As WFT generally pupate in the soil, we also assessed efficacy of B. bassiana in a granular formulation, in single and combined use with the predatory mite against pupating thrips. Based on our studies, control efficacy of thrips was not significantly improved when the predator and fungus were combined. The number of thrips per plant was higher in those treatments that used A. swirskii and fungus simultaneously than in treatments that used fungus and predatory mite alone. We observed a self-grooming behavior for mites exposed to the fungus conidia in which the mites spent a considerable amount of time that reduced effective searching and predation. Our results suggest that the simultaneous application of these two control agents (A. swirskii and B. bassiana) will not result in an additive or synergistic effect on the thrips population, but it seems separating of each other with regard to timing in an IPM program is important: One for preventative control and another as a curative treatment.

Entomopathogenic activity of Aspergillus flavus against tomato leaf miner, Tuta absoluta (Meyrick; 1917) in Tanzania



Never Zekeya, Musa Chacha, Patrick A. Ndakidemi, Ernest R Mbega

Department of Sustainable Agriculture and Biodiversity Conservation, School of Life Science and Bioengineering, Nelson Mandela African Institution of Science and Technology, P. O. Box 447, Tengeru Arusha, Tanzania. Corresponding author: neverz@nm-aist.ac.tz

Microbial controls of insect pest are of great focus toward developing an integrated management strategy and reduce development resistance among pest populations. Tomato leaf miner, Tuta absoluta (Meyrick) has become a major threat for tomato production and market in Tanzania. The pest is reported to cause up to 100% loss of tomato yield in the field. Management of T. absoluta by chemical pesticides is economically and environmentally unaffordable. However, management of T. absoluta by biocontrols from fungal sources is of economic interest due to the fact that insect cannot

develop resistance against biocontrols. Screening for entomopathogic activity was conducted by using two local isolates of Aspergillus flavus. The efficacy of A. flavus was evaluated against larvae and adult T. absoluta. In larval bioassay, leaves were used as experimental units whereas in adult bioassay plastic vials were treated as experimental unit. In both larval and adult bioassays units were treated with 1x10⁶, 1x10⁷, 1x10⁸, spores/ml of fungal solution. Mortality was recorded at interval of 24 hours. Results showed that A. flavus caused up to 70% larval mortality within 3 days whereas in untreated on 23.3% larval mortality was recorded in 16.6 days. A.flavus significantly reduced survival period of adult T. absoluta in which treated insects survived for 8 days only while untreated (control) survived for 25 days. This study reported the novel native biocontrol against *T. absoluta* in Tanzania. However further studies to isolate and characterize active compound on progress and will tested to evaluate its effectiveness under field conditions and might be potential biocontrol to other areas where T. absoluta is intensifying in Sub Saharan African.

Beauveria bassiana, Isaria fumosorosea, and Metarhizium brunneum antagonizing Fusarium oxysporum f.sp. vasinfectum in cotton

STU 4:45 •53•



Suchitra S. Dara¹, Sumanth S. R. Dara¹, Surendra K. Dara², Tim Anderson³

¹Global Agricultural Solutions, Bakersfield, USA, ²University of California Cooperative Extension, San Luis Obispo, USA, 3Dow, Shafter, USA Corresponding author: skdara@ucdavis.edu

In an effort to explore the potential of entomopathogenic fungi (EPF) in antagonizing plant pathogens, a greenhouse study was conducted using commercial formulations of three EPF against Fusarium oxysporum f.sp. vasinfectum Race 4 (FOV Race 4) . Pima cotton seeds susceptible to the pathogen were planted in potting medium infested with FOV Race 4 and Beauveria bassiana (BotaniGard ES), Isaria fumosorosea (Pfr-97), and Metarhizium brunneum (Met 52) formulations were applied in three treatment regimens. Three biofungicide formulations based on Streptomyces lydicus (Actinovate AG), extract of Reynoutria sachalinensis (Regalia), and Bacillus amyloliquefaciens (MBI 110) were also used to compare the efficacy of the EPF. The three treatment regimens were: A) soil application rate immediately after planting, B) soil application rate immediately and 1 and 2 weeks after planting, and C) foliar application rate immediately after planting. Plant health and growth were monitored 3, 4, and 5 weeks after planting. EPF offered varying degrees of protection against FOV Race 4 depending on the regimen and time after treatment. Isaria fumosorosea-treated plants looked relatively healthier 3 weeks after planting in regimens A and B while B. bassiana-treated plants looked healthier 3-5 weeks after planting in regimen C. This study helps expand the understanding of the alternative uses of EPF in agriculture.

Epizootiological studies of Beauveria bassiana infection in coffee berry borers in Hawaii

5:00 •54•



Louela A. Castrillo¹, Stephen P. Wraight¹, Sandy Galaini-Wraight², Tracie K. Matsumoto³, Lisa Keith³

¹ USDA ARS, Robert W. Holley Center, Ithaca, NY 14853, USA; ²Dept of Entomology, University of Hawaii, Honolulu, HI, 96822, USA; 3USDA ARS, Daniel Inouye Pacific Basin Research Center, Hilo, HI

Corresponding author: Louela.Castrillo@ars.usda.gov

As part of a project aimed at area wide control of the coffee berry borer (CBB) in Hawaii, we conducted a survey of indigenous Beauveria associated with the beetle and assessed their prevalence in fields where a commercial strain of Beauveria bassiana, GHA, has been applied. Surveys of infected CBB collected from study farms in South Kona and at survey sites in various districts in Hawaii and in Oahu from 2014 to 2017 resulted in > 1800 isolates of the fungus. Isolates were characterized initially using colony morphology to differentiate GHA from indigenous conspecifics. This facilitated rapid assessment of the prevalence of strain GHA in sprayed commercial farms. Molecular characterization of indigenous isolates using eight microsatellite markers revealed at least 10 haplotypes. Sequencing of the intergenic Bloc identified all strains as B. bassiana s. s., with six Bloc haplotypes present, one of which has been isolated from other insects collected in national parks and prior to CBB detection in the island. Prevalence data showed that observed

mycoses in coffee farms at the beginning of the new crop cycle are predominantly due to the indigenous strains, with GHA being reestablished following resumption of spray applications. In most farms the prevalence of fungal infection due to native B. bassiana strains rarely exceeded 5%. But one highelevation farm, with no history of GHA applications and surrounded by oldgrowth ohia forest, had 44% infection in CBB, suggesting that under highly favorable environmental conditions and with compatible cultural practices indigenous strains could play a significant role in controlling CBB populations.

Pathogenicity of Beauveria bassiana native strain in tobacco budworm Heliothis virescens

5:15 •55•



Cipriano García-Gutiérrez, Cosme Bojórquez-Ramos, Cesar Marcial Bonilla.

Laboratory of bioinsecticides . Instituto Politécnico Nacional. COFAA-CIIDIR IPN Unidad Guasave, Sinaloa, México . Corresponding autor: aarciaciprian@hotmail.com

Pathogenicity of a native strain of Beauveria bassiana Vuill. (Ascomycota: Hypocreales) was evaluated on third-and fifth-instar larvae of tobacco budworm, (TBW) Heliothis virescens (Fabricius). Larvae were immersed in 1x10⁸ conidia ml⁻¹. After 72 hours, the mortality rate for treating larval instar were 71 and 55%, while to nontreated check had 8 and 11%. The fungus was found across the cuticle and grew into the hemocoel 48 to 60 hours after inoculation in third-instar, and 60 to 72 hours in the fifth instar. The microscope and staining study showed that mycelia grew as filamentous hyphae on the surface and penetrated the cuticle and spiracles killed larvae. Inside the larvae, fungus grew as filamentous hyphae, but blastospores were not noticed, showed a specific infection events in TBW larvae, suggested that a native strain has potential as a bioinsecticide against H.virescens.

Use of generalist pathogens to control the invasive predator of bees Vespa veluting in Europe.





STU 5:30 •56•

Juliette Poidatz¹, Rodrigo J. López Plantey², Denis Thiéry¹

¹UMR-SAVE 1065, INRA, Villenave-d'Ornon, France ²Instituto de Biología Agrícola de Mendoza (CCT Mendoza CONICET-UNCuvo), Facultad de Agronomía, Mendoza, Argentina Corresponding author: Juliette.poidatz@inra.fr

While they arrive in a new area, invasive species often miss their native parasites. This was the case in the invasive hornet predator of bees, Vespa velutina, as he invaded Europe. Finding and destroying V. velutina colonies is problematic for several technical reasons (low detection rate, difficult access, stings risks), and furthermore for products toxicity reasons: destruction is dangerous for the applicator and the environment. In our work, we evaluated the potential efficiency of different French isolates of generalist entomopathogenic fungi, Metarhizium sp. and Beauveria sp. as biocontrol agent against V. velutina. We inoculated the hornets using different methods, mimicking natural ways of contamination: by immersion, by contact with a contaminated surface, in the food, and finally by inter-individual transfers, and kept them in controlled conditions. The isolates and inoculation methods efficiency were assessed by measuring two parameters: the Mortality Index (fungi lethality) and the aggressiveness (fungi killing speed). Our results reveal that the most lethal method of inoculation was immersion, and the less lethal was the contaminated food. No lethality difference was observed between the isolates when considering each inoculation method separately, except for the infected food, where the Metarhizium isolate EF3.5(2) was found deadlier. There was no difference in the aggressiveness between isolates for all and for each inoculation methods. The tested isolates showed a very high efficiency level in the controlled conditions that we tested here. Their potential as biocontrol agents on V. velutina must now be assessed on whole colonies. The possibility of using formulation adjuvants and strains mixes to enhance their efficiency must also be evaluated. This study is the first to test the potentiality of using biocontrol agents to limit the spread of an invasive hornet and to limit its impact.

Field tests on the biological control of leaf-cutter ants, the case of Acromyrmex lundii 5:45 •57 •

Analysis of C3larvin - a putative virulence factor of Paenibacillus larvae, the causative agent of American Foulbrood

STU 4:15 •59•

Patricia J. Folgarait, Daniela Goffré, Ariel Marfetán

Laboratorio de Hormigas, Departamento de Ciencia y Tecnología. Universidad Nacional de Quilmes, Bernal, Argentina. Corresponding author: patricia.folagrait@amail.com

Leaf-cutter ants are considered one of the most important pests of several crops in the Neotropics. Common chemical pesticides utilized to control them present serious problems to the ecosystems and human health, reasons why these compounds are being regulated by several certification organizations. Biological control using fungal pathogens is one of the alternatives which offer promising results in laboratory assays. However, there are few reported cases of utilization of biological control agents in field assays. The aim of this work was to demon-strate that Acromyrmex lundii colonies treated with fungal pathogens baits would reduce their survival, in contrast with colonies treated with control baits without fungi. The granulated baits used in this assay contained either Purpureocillium lilacinum (treatment PI), one entomopathogen that directly affects survival of ants or a combination (treatment BT) of another entomopathogen, Beauveria bassiana, and a mycopathogen, Trichoderma lentiforme, this latter affects the symbiotic fungus cultivated by ants as food (Leucoagaricus sp.). Each colony was assigned to one treatment and the activity (as the number of ants in each trail per minute) was measured weekly for 27 weeks and regularly until 51st week from the beginning, and the last measure was taken in the 58th week, this being the longest activity record known so far. At the end of the assay, 63% and 50% of colonies in treatment BT and PI, respectively, while only 14% of the control, were dead. Furthermore, our results allowed us to determine that 7 consecutive weeks of inactivation was a good criterion of colonies' death. Our results demonstrate the negative effect of these fungal baits on A. lundii colonies' survival and allow us to propose them as biological control agents against this ant species.

Marshall Room

Diseases of Beneficial Invertebrates Division #1

Moderators: Annette Bruun Jensen, Helen Hesketh

Missing sting reflex in Nosema ceranae infected honey bees

STU 4:00 •58•



Wei-Fone Huang, Zhi Ma

College of Bee Science, Fujian Agriculture and Forestry University, Fuzhou, Fujian, China, 350002 Corresponding author: wfhuang@fafu.edu.cn

Nosema ceranae is the most common gut pathogen in adult honey bees. N. ceranae is not identical with the original Nosema pathogen, N. apis, in Apis mellifera. Replacement of N. apis by N. ceranae has led to redefinitions of Nosema disease symptoms in honey bees. N. ceranae is thought causing some-how different symptoms. Many symptoms for N. apis infection were often neglected for N. ceranae without investigation. Missing sting reflex of moribund bees is one symptom that is usually ignored for N. ceranae although it is the first identified behavior change of Nosema disease. Sting behavior is an altruistic suicide for bees and applied for negative feedback learning experiments. We investigated if missing sting reflex is still a symptom for N. ceranae infection using electric stimuli settings in sting extension response (SER) learning experiments. N. ceranae inoculated bees showed significantly higher threshold of electric stimuli for SER. In addition, the infection intensity was correlated with the threshold. Missing sting reflex, however, did not indicate the moribund status that described in the classical symptoms. Bees with higher threshold for SER did not have signi-ficantly higher mortality after the test. In this study, we identified that N. ceranae infection can significantly retard the sting reflex after the infection reaches high intensity, and such infection seems not killing the host soon. We hypothesized that this behavior change can benefit N. ceranae since the hosts fully loaded with N. ceranae spores tended not to sting and lead to altruistic death.

Julia Ebeling¹, Anne Fünfhaus¹, Daniel Krska², Ravikiran Ravulapalli², A. Rod Merrill², Elke Genersch¹

¹Institute for Bee Research, Hohen Neuendorf, Brandenburg, Germany: ²Department of Molecular and Cellular Biology, University of Guelph, ON, Canada Corresponding author: elke.genersch@rz.hu-berlin.de

Paenibacillus larvae is a Gram-positive, spore-forming bacterium causing the devastating honey bee brood disease American Foulbrood (AFB) which is leading to colony losses worldwide. There are four genotypes of P. larvae identified by rep-PCR with enterobacterial repetitive intergenic consensus (ERIC) primers which are thereafter referred to as ERIC I- IV. The genotypes differ phenotypically, amongst others in virulence, and seem to possess different strategies to breach the larval gut epithelium and enter the host's haemocoel. For ERIC I and ERIC II, which are most frequently isolated from current AFB cases, several virulence factors have been identified so far and confirmed in exposure bioassays. Recently, a further putative virulence factor, the C3 exoenzyme C3larvin, has been biochemically and structurally characterized. Yet the role of C3larvin as virulence factor still needs to be examined. The aim of this study was to further analyze the C3larvin locus in the genome of the different P. larvae genotypes in silico and to investigate the biological role of C3larvin in exposure bioassays with knockout mutants.

Solitary bee nesting behavior in the presence of pathogen killed larval cadavers

4:30 •60•



Ellen Klinaer. Diana Cox-Foster

USDA-ARS Pollinating Insect Research Unit; Logan, UT, USA Corresponding author: ellen.klinger@ars.usda.gov

The alfalfa leafcutting bee, Megachile rotundata, is a solitary nesting bee currently managed for commercial production of alfalfa seed in the northwest United States and Canada. Larvae of this bee suffer from a disease caused by fungi in the genus Ascosphaera. This disease, known as chalkbrood, results in hardened larvae that block the nesting cavity and must be bypassed by healthy nestmates when those nestmates emerge as adults. It has been thought that these emerging healthy adults chew through, and may partially consume part of these larval cadavers. We conducted a three year field study to characterize how larval cadavers affected adult bee nesting behavior. This study showed that the presence of cadavers improved the number successful offspring. Using lab based experiments, we quantified this behavior to a greater degree to determine if adult bees are consuming fungal cadavers and propose what this may mean for chalkbrood dynamics in these bee populations.

Deformed Wing Virus of Honeybees - transmission, diversity and impact on honeybee development





Jessica Fannon¹, Dave Chandler¹, David Evans²

¹Warwick Crop Centre, University of Warwick UK; ²School of Biology, University of St Andrews, Scotland UK Corresponding author: j.m.fannon@warwick.ac.uk

The spread of the parasitic mite, Varroa destructor (and the RNA viruses that it transmits between honeybees) is closely linked to increases in overwintering losses and declining honeybee health. Deformed wing virus (DWV; Iflaviridae) a picorna-like single-stranded, positive-sense, RNA virus, is widespread in honeybees and usually present as a low level, asymptomatic infection. However, the transmission of DWV by Varroa to developing pupae causes highly elevated virus levels and characteristic developmental abnormalities. Our work aims to further the understanding of DWV virulence and pathogenicity and reveal how this is linked to Varroa parasitism. We are using molecular techniques and an in vitro larval rearing system to study how transmission route influences DWV accumulation and localisation in honeybee tissues. We are also investigating the impact of Varroa on DWV levels and diversity on both a colony and a landscape level, testing the hypothesis that a single virulent strain has a selective advantage when transmitted by Varroa. We have found that the route of transmission determines DWV infection and tissue location in developing honeybees, as

shown by RNA-FISH. The relative expression levels of key developmental genes from the Homeobox and Ecdysone families have been established using qPCR in an attempt to reveal more about the developmental abnormalities in symptomatic bees. In addition to this, a long-term study of viral diversity in the Ardnamurchan Peninsular, Scotland, illustrates how the viral landscape is affected by the introduction of the Varroa mite.

Metagenomic analysis reveals novel RNA viruses in honey bee colonies in Brazil

5:00 •62



Fernando L. Melo¹, Diouneia L Berlitz², Daniel Ardisson-Araujo³, Lidia Fiuza⁴, Bergmann Morais Ribeiro ¹

¹Laboratory of Baculovirus, Cell Biology Department, University of Brasília, Brasília, DF, Brazil; ²Control Agro Bio Pesquisa e Defesa Agropecuária, Porto Alegre, RS, Brazil: 3 Laboratory of Insect Viruses, Biochemistry and Molecular Biology Department, Federal University of Santa Maria, Santa Maria, RS, Brazil; ⁴Instituto Rio Grandense do Arroz, Cachoeirinha, RS, Brazil. Corresponding author: flucasmelo@amail.com

Bees are considered to be the most important group of pollinators. In recent years populations of these insects have declined worldwide and considerable losses have been also reported in the southern and southeastern states of Brazil. Several viruses have been described infecting honey bees, however the studies are still rare and a number of honey bees viruses are still unknown and remain to be discovered. In this study, adult insects and larvae were collected in several production regions in Rio Grande do Sul State, Brazil. The insects were macerated in SM buffer and centrifuged in a low rotation to remove the solid matter followed by an ultracentrifugation with 20% sucrose cushion to viral particles concentration. RNA extraction was performed using ZR Soil/Fecal RNA MicroPrep (Zymo Research). The sample was processed for rRNA removal using Ribo-Zero rRNA removal kit (Bacteria) (Illumina, San Diego, CA, USA) and cDNA library construction using TruSeq RNA library preparation kit (Illumina). The samples were sequenced at Macrogen (Seoul, Republic of Korea) using Illumina HiSeq 2000 paired-end method. The raw reads were quality trimmed and de novo assembled using CLC Genomics Workbench version 6.3. The resulting contigs were compared to ViralRefSeq complete genomes available in genbank using BLASTx implemented in Geneious version 9.1.5. This revealed several viruses already reported infecting honey bees (such as Bee Macula-like virus, Varroa destructor virus-1, Aphid lethal paralysis virus and Lake Sinai virus) and also two novel viruses, including a Rhabdoviruses and an Orthomyxovirus. All viral genomes were complete or almost complete and their ORF distribution resembles that of their most close relatives. Their host range and geographical distribution are currently under evaluation.

Creating a baseline and examining spillover with an all-taxa parasite inventory of bumble bees in the United States



Amber D. Tripodi, James P. Strange

USDA-ARS, Pollinating Insects Research Unit, Logan, UT, USA Corresponding author: amber.tripodi@ars.usda.gov

Bumble bees are important crop pollinators, and their commercial production has led to wide-scale movement of mass-produced colonies. There are concerns about the potential for pathogen spillover into wild populations, yet little is known about the current distribution of pathogens and parasites in wild bumble bee communities. To understand the distribution of parasites geographically and across bumble bee species, a total of 3670 bumble bee specimens, representing 31 species, were collected from 18 states across the USA in 2015. To address pathogen spillover, we included 13 site pairs in which a site with a known history of commercial bumble-bee use was paired with a wild site nearby. Both microscopy and molecular diagnostics were implemented to inventory invertebrate parasites, pathogenic microorganisms, and the presence of common honey-bee viruses in each specimen. Of the macroparasites observed, conopid flies were most common, although phorid flies, tracheal mites, and nematodes were also observed. Trypanosomatids were common, with Crithidia bombi more common than C. expoeki. Of the microsporidia, only Nosema bombi was observed. Black Queen Cell virus and Sacbrood virus were the most commonly seen viruses, although little is known about the pathology of these viruses in humble bees. Parasite communities differed between

commercial and wild sites, and among species and geographic regions. Our results provide a baseline of parasite pressures faced by bumble bees as well as insight into pathogen spillover across the US.

Viability of Dicrocoelium dendriticum metacercariae in Formica polyctena ants after freezing, boiling or ethanol treatments





Annette. B. Jensen, Joanne Malagocka, Jørgen Eilenberg, Brian L. Fredensborg

University of Copenhagen, Department of Plant and Environmental Sciences, Frederiksberg C. Denmark: Corresponding author: abj@plen.ku.dk

Edible insects have gained increased recognition for their taste, nutritional value, and potential for commercial-scale production of foods for human consumption and animal feed. Insect pathogens and parasites are naturally occurring in insect species used for human consumption. The presence of such pathogens and parasites in insects collected in nature is, however, a concern due to the potential hazards they may cause consumers. Red wood ant Formica polyctena, is an intermediate host of the trematode Dicrocoelium dendriticum and infected ants are often observed in nature. Since D. dendriticum via invertebrate hosts can infect mammals, care is needed when using ants collected in nature for human consumption. In this study we developed a viability assay including trypsin treatment of D. dendriticum metacercariae from the abdomen of the red wood ant Formica polyctena. Then we tested the viability of D. dendriticum metacer-cariae in relation to: 1) freezing (- 20 and -80 °C), 2) boiling (100 °C), and 3) ethanol (50 %). The metacercariae in all control ants were alive and displayed movements either in the cyst, or after excystation, whereas all metacercariae from all treatment groups were recorded as dead. Freezing at -20 °C or -80°C for 30 min, boiling for 1 minute or storage in 50 % ethanol for 24 h proved effective in killing the lancet liver fluke D. dendriticum metacercariae. We therefore recommend treating ants collected in wild by one of the treatments before consumption.

MONDAY, 8:00 - 10:00 pm

Bacteria Division - Business Meeting (Village Tower West, 15th fl., Rm. 15C)

Diseases of Beneficial Invertebrates – Business Meeting, (Village West, Building 2, Room 2C)

Microbial Control - Business Meeting (Village Tower West, 15th fl., Rom. 15B)

Nematodes Division - Workshop

(Village West, Building 2, Rooms 2A/2B)

TUESDAY, 6:30 - 7:30 am

SIP 5K Run/Walk

Assemble on North Point Lane in front of the Spanos Athletic Training Facility.

TUESDAY, 8:00 - 10:00 am

Theater

VIRUS AND DBI CROSS-DIVISIONAL SYMPOSIUM

Honoring Just Vlak

Organizers and Moderators: Martin Erlandson, Kelly Bateman

Introduction - Just Vlak - impact and international collaboration on invertebrate virus research

8:00 •65•

Martin Erlandson

DNA Viruses of Aquatic Invertebrates

8:10 •66•



Kelly Bateman, Grant Stentiford

European Union Reference Laboratory (EURL) for Crustacean Disease, Cefas, Barrack Road, The Nothe, Weymouth, Dorset, DT4 8UB, UK, Corresponding author: Kelly.bateman@cefas.co.uk

The first invertebrate virus discovered from the marine environment was described infecting a crab. Since this discovery there have been numerous virus infections described infecting a range of aquatic invertebrates. Perhaps the most high profile of these is the White Spot Syndrome Virus (WSSV), which despite almost two decades since its discovery, is still considered the most significant known pathogen impacting the sustainability and growth of the global penaeid shrimp farming industry (with annual losses exceeding US \$1bn). The virus was originally discovered in 1991 in Penaeus japonicus in China and Taipei, spreading rapidly throughout Asia and then the Americas during the mid- to late- 1990s. Due to the rapid spread of the virus and the isolation and identification by numerous laboratories the virus has been referred to by a variety of different names, until it was later agreed that these infections were caused by the same agent and the name White Spot Syndrome Virus (WSSV) was adopted. WSSV was classified at the laboratories in Wageningen University, full genome was determined and data showed that this large virus was unlike any other known viruses. A new virus family and genus were created, with WSSV virus as the sole member of the family Nimaviridae. Numerous publications have been written describing the protein structure, infection mechanisms, susceptible hosts and genome composition of this virus. In addition, recent work conducted on wild populations of shore crabs have identified a virus which is similar but not identical to WSSV. Whilst several crab viruses have previously tentatively been classified within the Nimaviridae family based upon structural characteristics, these classifications were later removed due to lack of genomic information. The discovery and description (pathology, morphology, genome) of a novel virus infecting crabs and belonging to the Nimaviridae not only allows for expansion of this single-taxon family but further, shines a light on the potential origin of WSSV in wild animal reservoirs.

Baculovirus Budded Virions: Entry mechanisms and progress toward understanding virus-cell interactions during entry

8:30 •67•

Gary W. Blissard¹, Jeffrey Hodgson¹, Ya Guo^{1,2}, Zhaofei Li², Nicolas Buchon³

¹Boyce Thompson Institute, Cornell University, Ithaca, NY, 14853, USA; ²Northwest A&F University, Yangling, Shaanxi 712100, China; 3Cornell University, Ithaca, NY 14853, USA Corresponding author: gwb1@cornell.edu

In typical baculovirus infections of lepidopteran larvae, infection of the midgut by virions of the Occlusion Derived Virus (ODV) phenotype leads to the production of progeny Budded Virions (BV). BV bud from basal membranes of the polarized midgut epithelial cells and subsequently infect tracheolar cells, hemocytes, and many other cell types. BV entry into the host cell is mediated primarily by a single major envelope glycoprotein that facilitates binding to the (unknown) host cell receptor, a process that appears to stimulate endocytosis. The endosome containing the BV is transported within the cell and progressively acidified, which triggers a conformational change in the major envelope protein and activates its membrane fusion activity. The attachment-and-fusion protein found on

baculovirus BV is named F in some baculoviruses, and GP64 in other baculoviruses. F and GP64 proteins represent two distinct lineages of envelope fusion proteins, each likely acquired by baculoviruses, from either the host genome or from another virus. How BV binding initiates endocytosis and how endosomes are trafficked within the cell during entry are largely unknown. New approaches and advances in our understanding how baculovirus BV envelope proteins mediate attachment and fusion, and how BV navigate entry utilizing cellular pathways, will be reviewed.

Occlusion derived virus the baculovirus virion that is specialized for infection of insect midguts: Identifying proteins essential for oral infectivity

8:45



David A. Theilmann¹, Martin A. Erlandson², Leslie G. Willis¹, Ajay Maghodia¹, Monique van Oers³

¹Summerland Research and Development Centre, Agriculture and Agri-Food Canada, Summerland, BC, VOH 1Z0 Canada; ²Saskatoon Research and Development Centre, Agriculture and Agri-Food Canada, Saskatoon, SK, S7N 0X2, Canada; ³Laboratory of Virology, Wageningen University, Droevendaalsesteeg 1, 6708 PB Wageningen, The Netherlands Corresponding author: david.theilmann@aqr.qc.ca

Baculoviruses are unique in that two virion phenotypes are produced during infection, budded virus (BV) and occlusion derived virus (ODV). In vivo, BV are initially produced in the host midgut cells and cause systemic spread to all susceptible tissues. ODV in contrast are highly specialized for oral infectivity and binding and infection of the host midgut cells. ODV can contain one or many nucleocapsids per virion envelope. The virion envelope is derived from the inner nuclear membrane and becomes associated with the ODV nucleocapsids in the nucleus. The ODV envelope contains a large subset of proteins not found in BV, many of which appear to be required for binding and attachment to midgut cells. These proteins have become known as per os infectivity factors (PIFs). All PIF proteins have common properties which include, being a baculovirus core gene, associated with the ODV envelope and deletion results in the virus being unable to orally infect a host. In addition, deletion of a pif gene does not impact virion replication, BV production or BV infectivity. To date 10 PIFs have been identified PIF0 through PIF9. Extensive work on the individual PIFs has identified specific functionality but they all appear to form a complex in the ODV envelope that is essential for oral infectivity. The core of the complex, PIF1-4, appears to serve as a scaffold to which the other PIFs associate. It is known that the PIFs are required for recognizing and binding to a midgut receptor which is required for membrane fusion. In contrast, PIF proteins do not appear to have any role in cell culture infectivity highlighting the highly specialized role of these proteins. Of significant interest is the discovery of PIF homolgues in many large DNA insect viruses, including the Nudiviridae and Polydnaviridae, suggesting that their mechanism of action involves a pathway that is conserved across a broad range of evolutionarily distant host insects.

Recent advances in the biotechnological applications of baculoviruses: vaccines and gene therapy

9:00

Linda A. King

Oxford Brookes University, Oxford, UK Corresponding author: laking@brookes.ac.uk

It has been more than 30 years since baculovirus expression vectors were first introduced. They now take their place as one of the most commonly used gene expression systems, in both insect and mammalian cells, across the world. In recent years, scientists working both in academia and in industry have taken the system forward far beyond the simple expression of proteins, to produce components for vaccines and therapies, for cancer as an example. Whilst today the number of vaccines on the market are small in number, many more are in various stages of R&D and clinical trials both for human and veterinary markets. There is also a growing body of research aimed at using BacMam vectors for gene therapy, for example, in the field of transplantation. BacMam vectors are being evaluated to deliver genes to alleviate the cause of ischaemic reperfusion injury that is the major cause of transplant failure. The baculovirus expression system is also being used to produce human viral vectors for gene therapy, such as adeno-associated virus. These developments and advances rely on basic science to understand

the replication and biology of baculoviruses, such as that undertaken over many years in Wageningen University under the leadership of the scientist that we are here to honor, Just Vlak.

Basic research supporting the development of baculoviruses as biopesticides

9:20 •70•

Zhihong Hu

¹State Key Laboratory of Virology, Wuhan Institute of Virology, Chinese Academy of Sciences, Wuhan, 430071 People's Republic of China Corresponding author: huzh@wh.iov.cn

The first publication of using baculovirus for pest control was published in 1925. In 1930-1940s, baculovirus played an important role on control European spruce sawfly in North America. These events promoted the early studies of biology and pathology of baculoviruses, as well as safety considerations and the potential of using baculoviruses as pest control agents. The first viral pesticide ElcarTm was registered in USA in 1975, followed by worldwide interest. Successful examples include using Anticarsia gemmatalis multiple nucleopolyhedrovirus (AgMNPV) on soybean in Brazil, Helicoverpa armigra NPV on cotton in China, Cydia pomonella granulovirus (CpGV) on apple in Europe and North America, etc. In 1970s-1980s, the development of recombinant DNA technology helped to identify best strains for pest control. Later on, functional analyses discovered baculoviral genes involved in the insecticidal properties, such as genes encode for ecdysteroid UDP-glucosyltransferase (EGT), polyhedron envelope protein, chitinase, cathepsin, enhancin, ODV-E66, and per os infectivity factors. Despite the widespread interest, the use of baculoviruses for pest control has been limited. This is largely due to the shortcomings of baculoviral pesticides, namely narrow host range, slow speed of kill, sensitive to UV lights and high does needed for control high instar larvae. During late 1980s to middle 1990s, baculovirues were engineered to enhance their properties as pesticides. These include expressing insect hormones or enzymes, deletion of EGT gene, and expression of insect specific toxins. Many of such modifications successfully reduced the LT50 of baculoviruses, however, due to the consequences on the loss of yields and the strict regulation on GMO, recombinant baculoviral pesticides remained at the level of laboratory research or field trials. Recently, the studies of resistance of codling moth to CpGV pesticide revealed the complexity of virus-host interaction during application of baculoviral pesticide. Forty years have been passed since the first registration of a baculoviral pesticide, yet the key factors determine the insecticidal properties still need to be identified. In future, the combination of molecular virology, entomology and ecology will disclose the mechanisms of virus infection and insect behavior. The results of such basic research will promote development for better viral pesticides.

dsDNA Viruses of Invertebrates - Taxonomy and Phylogeny

9:40 •71•



Johannes A. Jehle

Institute for Biological Control, Federal Research Center for Cultivated Plants, Julius Kühn Institute, Darmstadt, Germany Corresponding author: johannes.jehle@Julius-Kuehn.de

Since the very first discovery of viruses infective for invertebrates, numerous groups of DNA viruses have been discovered. Some of these viruses appear to be highly diverse but to share also specific genetic, morphological and pathological characteristics, such as circular double-stranded DNA genomes, a set of common gene othologs, rod shaped nucleocapsids, viral replication in the cell nucleus, and nuclear hypertrophy. Taxonomy and classification of these viruses changed during recent decades of research, depending on classification criteria and methodology used. With the development of molecular techniques, concepts of molecular evolution and mathematical methods to deduce phylogenies, and firmly expedited by the Vlak group in Wageningen, it became obvious that a supergroup of supergroup of nuclear arthropod-specific large DNA viruses, comprising baculoviruses, nudiviruses, hytrosaviruses and nimaviruses most likely shared common ancestor.

The Forum

8:00 •72•

Bacteria Division #2

Moderator: Marianne Carey, Shuyuan Guo

Increasing virulence and producing resistance-busting mutants of Bacillus thuringiensis using experimental evolution

Ben Raymond¹, Neil Crickmore², C. James Manktelow¹, Tatiana Dimitriu¹

¹Department of Bioscience University of Exeter Penryn campus Cornwall LIK: ² School of Life Science, University of Sussex, Brighton, UK Corresponding author: b.raymond@exeter.ac.uk

Insects, and other organisms, can evolve resistance to pathogens under strong selection pressure. In turn, co-evolutionary theory predicts that pathogens should be able to adapt to host resistance and acquire 'resistance busting' mutations that enable them to successfully infect hosts once again. We designed an efficient regime to impose artificial selection for the increased virulence of Bacillus thuringiensis kurstaki against a Cry1Ac resistant population of diamondback moth, Plutella xylostella (with ≈ 10,000 fold resistance). Selection regimes were based on previous work describing the cooperative nature of virulence in B. thuringiensis. Population structure during selection, and bacterial mutation rate, had a strong impact on how bacteria responded to selection. Selection regimes produced mutants that had increased virulence by up to 300-fold; mutants with single order of magnitude increases in virulence were common. Preliminary sequencing results suggest that the best performing mutants have not evolved Cry toxins with new sequences. However, this type of selection regime may be valuable as a means of genetic mining for novel virulence factors or increasing the potency of microbial products.

Resistance to dual-toxin Bt cotton in the pink bollworm, Pectinophora gossypiella

8:15 •73•



Jeffrey A. Fabrick¹, Lolita G. Mathew¹, Jeyakumar Ponnuraj², Xianchun Li³, Yves Carrière³, Bruce E. Tabashnik³

¹USDA ARS, U.S. Arid Land Agricultural Research Center, Maricopa, AZ, USA; ²National Institute of Plant Health Management, Rajendranagar, Hyderabad, Andhra Pradesh, India; 3 University of Arizona, Department of Entomology, Tucson, AZ, USA Corresponding author: jeff.fabrick@ars.usda.gov

Proteins from the bacterium Bacillus thuringiensis (Bt) are produced by transgenic crops targeting several major insect pests. Bt crops can suppress insect pest populations, reduce reliance on conventional insecticides, and increase yields and farmers' profits. Unfortunately, such benefits are being reduced by the rapid evolution of pest resistance to Bt toxins. In the U.S., Bt cotton has been a key tool for managing the pink bollworm and this pest has remained susceptible to both single-toxin (Cry1Ac) or dual-toxin (Cry1Ac + Cry2Ab) cotton. In India, the pink bollworm has evolved practical resistance to both single and dual-toxin Bt cotton. Here we report progress in characterizing the molecular genetics and biochemical mechanisms of pink bollworm resistance to Cry1Ac and Cry2Ab from both India and the U.S.

Functional evaluation by gene editing of a Bacillus thuringiensis Cry1Ac toxin receptor in Helicoverpa zea

8:30 •74•



Omaththage P. Perera¹, Nathan S. Little¹, Calvin A. Pierce¹, Heba Abdelgaffar², Lin Niu², Randall G. Luttrell¹, Juan Luis Jurat-Fuentes²

¹USDA-ARS Southern Insect Management Research Unit, Stoneville, MS; ²Department of Entomology and Plant Pathology, University of Tennessee, Knoxville, TN Corresponding author: op.perera@ars.usda.gov

Naturally occurring mutations in Bacillus thuringiensis (Bt) toxin receptors in insects are known to confer tolerance to Bt toxins. ATP Binding cassette transporter C2 (ABCC2) is a multi-domain Bt toxin receptor that confers tolerance to the Cry1Ac Bt toxin. We report the use of gene editing to truncate ABCC2 in the cotton bollworm (Helicoverpa zea) and test its receptor functionality. Effects on tolerance to Cry1Ac, brush border vesicle membrane binding with Cry1Ac, and fitness costs of mutations were studied using bollworm lines with edited mutations in different domains of ABCC2.

Cry11B enhances efficacy significantly to Aedes aegypti when added to a Bacillus thuringiensis strain producing the Cyt1A-BinA chimera

8:45 •75•

Hyun-Woo Park^{1,2}, Dennis K. Bideshi^{1,2}, Brian A. Federici^{1,3}

¹ Department of Entomology, University of California, Riverside, Riverside, CA, USA; ²Department of Biological Sciences, California Baptist University, Riverside, CA, USA; ³Graduate Division, University of California, Riverside, Riverside, CA, USA Corresponding author: hpark@calbaptist.edu

Two naturally occurring species of bacilli, Bacillus thuringiensis subsp. israelensis (Bti) and Lycinibacillus sphaericus (Ls), are used worldwide for controlling larvae of nuisance and vector mosquitoes. The insecticidal activity of Bti is due to synergistic interactions among its four major proteins (Cry4Aa, Cry4Ba, Cry11Aa, Cyt1Aa), while the activity of Ls is due to a binary toxin (Bin) consisting of a toxin domain, BinA, and a midgut receptor-binding domain, BinB. Although used commercially for almost three decades, reports of mosquito resistance to Bti have been rare. However, levels of resistance greater than 10,000-fold to Bin have been reported where Ls has been used intensively for mosquito control. Cyt1Aa is a lipophilic protein, and in previous studies we showed it delays the evolution of resistance to the Cry proteins of Bti, and can overcome high levels of resistance Bin. In a previous study, we fused Cyt1Aa to BinA, using the lipophile as a broad-spectrum binding domain and showed that the Cyt1Aa-BinA chimera was remarkably toxic to five major vector species of mosquitoes, Anopheles gambiae, An. stephensi, An. quadrimaculatus, Bin-sensitive and Bin-resistant strains of Culex quinquifasciatus, and Aedes aegypti, the latter not normally sensitive to Ls. However, toxicity against Aedes aegypti was not as high as against other mosquito species. Here we show that introducing another highly mosquitocidal protein, Cry11B from B. thuringiensis subsp. jegathesan, enhances the chimera's toxicity against Ae. aegypti significantly.

Interaction of Bacillus thuringiensis toxins and abamectin in Cry1Ac-resistant Heliothis virescens

9:00

David G. Heckel. Andrea Barthel

Max Planck Institute for Chemical Ecology Hans-Knoell-Str. 8, 07745 Jena, Germany Corresponding author: heckel@ice.mpa.de

A previous study reported that a strain of Helicoverpa armigera with a mutation in the ABCC2 gene conferring resistance to the Cry1Ac toxin of Bacillus thuringiensis (Bt) was nine-fold less tolerant to abamectin than a control Bt-susceptible strain (Y. Xiao et al. (2016) PLoS Pathogens 12(2):e1005450). This suggests that abamectin might be useful in combatting Bt resistance when mutations in this ABC transporter are involved. To investigate the generality of this phenomenon, we examined the interaction in the YEE strain of Heliothis virescens which has the same type of Bt resistance. When expressed in Sf9 cells, the ABCC2 protein from H. virescens transported abamectin in inside-out membrane vesicles, confirming the previous results with H. armigera. The two components of abamectin were differentially transported. Bioassays utilizing abamectin incorporated into artificial diet revealed less than a four-fold susceptibility difference between YEE and the JEN2 susceptible strain. These bioassays were complicated by the unique mode of action of orally-administered abamectin. Independent of the concentration, larvae would eat just enough diet until they were paralyzed. Mandibles and body musculature stopped moving; however the heart continued beating for several days. Functionally speaking, the relevant parameter is the median effective concentration for paralysis, not the median lethal concentration. There was no significant difference in tolerance of the YEE and JEN2 strains to topical application of abamectin. This suggests that the utility of abamectin as an strategy to selectively kill Bt-resistant insects in the field will depend on how much mortality, or paralysis, occurs by the oral versus the cuticular route.

Modeling peritrophic matrix defense against pathogens infecting per os in Leptinotarsa decemlineata (Coleoptera: Chrysomelidae)







Emre Caner¹, Umut Toprak¹, Serife Bayram¹, Dwayne Hegedus², Doug Baldwin², Cathy Coutu², Martin Erlandson², David Heckel³

¹Ankara University, Faculty of Agriculture, Dept. of Plant Protection, Ankara, Turkey; ² Agriculture and Agri-Food Canada, Saskatoon Research Centre, Saskatoon, SK Canada; ³ Max Planck Institute for Chemical Ecology, Dept. of Entomology, Jena, Germany Corresponding author: utoprak@agri.ankara.edu.tr

Many insects possess a porous structure lining the midgut called the peritrophic matrix (PM), which is composed of chitin and proteins. One essential role of PM is its capacity as a barrier against pathogens infecting per os such as Bacillus thuringiensis. It is critical to detect PM-associated defense proteins for understanding midgut defense and developing more effective B. thuringiensis formulations. Colorado potato beetle (CPB) is a major pest of potato and has developed resistance against the endotoxin of several B. thuringiensis strains. In this study, midgut specific-cDNA libraries were generated from 4th instar larvae and adults of CPB, and PM defense proteins were identified by liquid chromatography-tandem mass spectrometry and functional annotation. The PMs were found to possess pattern recognition proteins such as thaumatin, peptidoglycan recognition protein, C- type lectin, and beta-galactosidase, beta-mannosidase and REPAT proteins. Transcriptional levels of the corresponding genes were predicted using reads per kilobase per million mapped reads method (RPKM). Presence of defense proteins in the PM and their expression patterns suggest that PM may serve as a biochemical barrier in the resistance mechanisms developed against B. thuringiensis toxins. Finally, a model describing the role of beetle PM in defense is presented.

A chitin binding protein involved in adhesion of Bacillus thuringiensis to the insect peritrophic matrix

Zongxing Tong¹, Yiling Zhan¹, Fuping Song², Christina Nielsen-LeRoux³, Shuyuan Guo¹

¹School of Life Science, Beijing Institute of Technology, Beijing 100081, China; ²State Key Laboratory for Biology of Plant Diseases and Insect Pests, Institute of Plant Protection, Chinese Academy of Agricultural Sciences. Beijing 100193, China; ³ Génétique Microbienne et Environnement, INRA, UMR 1319 Micalis, Domaine de Vilvert , 78253 Jouy en Josas, France Corresponding author:quosy@bit.edu.cn

As an insect pathogen, an important step of infection for Bacillus thuringiensis (Bt) is adhering to the host. The peritrophic matrix (PM) is a natural protective barrier of insect gut, mainly composed of chintin and mucus protein. Most Lepidopteran larval midguts are alkaline. Bt can produce several non-insecticidal extracellular macromolecules such as proteases, chitin binding proteins (CBP), lipases, etc., to facilitate the infection process. Bt HD73 strain can produce two kinds of chitin binding proteins encoded by cbp3152 and cbp3189. A RNA-Seq approach showed that transcription of cbp3189 is induced under alkaline condition. The Cbp3189 gene was cloned and the protein expressed in E.coli BL21 strain. CBP3189 was purified by chitin affinity chromatography, confirming its chitin binding ability. No chitinase activity was detected from this protein, but it could bind to the isolated cell wall of the HD73 strain and the PM of Ostrinia furnacalis (Guenée) and Galleria mellonella. Gfp translational fusion showed the subcellular location of CBP-3189 protein is on both bacterial surface and the spore. A transcriptional fusion with lacZ confirmed that the promoter of cbp3189 is induced by alkaline pH. Based on the above studies, it is speculated that the expression of CBP-3189 on the bacterial surface might trough the N-terminal chitin-binding domain stick to the PM and help Bt to be maintained in contact with the midgut cells in a susceptible insect. Interestingly, in Galleria mellonella, which is not susceptible to the Bt HD73 Cry toxin, the cbp3189 deletion mutant persisted better in the gut than the wild type which might be explained by a decrease in PM binding and thereby a lower expulsion rate with the feces.

The impact of plant pyramids and planting mosaic on the evolution of resistance to Bt toxins

9:45 •79•

Kanglai He, Yueqin Wang, Yudong Quan, Zhenying Wang

The State Key Laboratory for Biology of Plant Diseases & Insect Pests.

Corresponding author: hekanalai@caas.cn

Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing, China

Bt crops have been planted by farmers since 1996. Control failures due to the

evolution of insect resistance have been reported in several field populations.

Preventing or delaying the evolution of resistance to Bt is critical for the sustainable utilization of this state-of-the-art technology. Although Bt crop

pyramids have the advantage for resistance management, deployment of

especially it is usually along with the difficulty of using structured refuges.

We tested how applications of Bt crop pyramids (selection with bi-mixed

different single-gene plants will be a vital in the small-hold farmers in China,

toxins, Cry1Ab+Cry1F, Cry1Ab+Cry1Ie) and planting mosaic (mixed selection with alternating between Cry1Ab and Cry1F and/or Cry1le) affected the

evolution of resistance in Ostrinia furnacalis using multi-generation selection

experiments. The LC50 value of each toxin estimated from a 7-day bioassay

were deployed as selection pressure. There were significant differences in

the rate of evolution of resistance among selection regimes. Selection with

bi-mixed toxins resulted in 10-fold and 50-fold of resistance to Cry1Ab toxin

after 9- and 12-generation of selection with Cry1Ab+Cry1F toxins and

Cry1Ab+Cry1le toxins, respectively. In contrast, mixed selection with alternating between Cry1Ab and Cry1F and/or Cry1le resulted in 47-fold and

83-fold resistance to Cry1Ab toxin. These data indicated that resistance

evolved more quickly with treatments of mixed selection with alternating

between two toxins than that selection with bi-toxins. The results help to

understand better the impact of plant pyramids and planting mosaic on the

Roosevelt Room

FUNGI DIVISION SYMPOSIUM

Insect defense against fungal pathogens: mechanisms, variations and efficacy

Organizer/Moderator: Louela A. Castrillo

8:00 •81•

Nemat O. Keyhani

University of Florida, Dept. of Microbiology and Cell Science, Bldg 981, Museum Rd., Gainesville, FL 32611, USA, and Genetic Engineering Research Center, School of Life Sciences, Chongqing University, Chongqing, China

Defining new cuticle barrier steps in the infection process of insect pathogenic fungi

Corresponding author: kevhani@ufl.edu

Successful mycosis and the process of penetration into and out of the host insect have long been recognized to involve more than just the activities of cuticle-degrading enzymes. Recent evidence suggests that ingress and egress are two distinct steps in the infection process, requiring different genetic factors. The cuticle waxy-layer and its lipid constituents can act as growth substrates that affect conidial virulence and lipid mobilization. For example, Beauveria bassiana conidia grown in the presence of oleic acid displayed a ~4-5 fold decrease in mean lethal dose (LD₅₀) as compared to standard media alone. Manipulation of the ability of entomopathogenic fungi to adapt and/or detoxify host cuticular secretions has also been shown to result in an arms race between Beauveria bassiana and tenebrionid secretions. In this instance, cuticular quinones were detoxified by a fungal benzoquinone reductase, although the beetle, Tribolium castaneum secretes high enough levels of cuticular quinones to inhibit fungal growth for most wild type strains of entomopathogenic fungi. Fungal perilipin and caleosin, proteins involved in lipid assimilation, storage, and turnover, are involved in appressorial turgor pressure and hence cuticle penetration. Caleosin activity is also involved in spore dispersal, and intriguingly, melanization and/or production of secondary metabolites, including oosporein, on the insect cadaver. The production of oosporein has been shown to be a late-stage, post host death event, indicating that the fungus continues to respond to the insect even after death of the host. In this case, oosporein appears to act as an antimicrobial compound on the dead host cuticle in order to minimize growth of competing microbes. These advances have led to an expansion of our model of the pathogenic process employed by entomopathogenic fungi during infection of insect hosts.

Impact of parasitization of Macrocentrus cingulum and Cry1Ac protein on cellular immunity and humoral immunity within hemolymph of susceptible and resistant Asian corn borer

evolution of insect resistance to Bt crops.

Zengxia Wang^{1,2}, Kanglai He¹, Shuxiong Bai¹ Tiantao Zhang¹, Wanzhi Cai², **Zhenying Wang**¹*

STU 10:00 •80•

¹ State Key Laboratory for Biology of Plant Diseases and Insect Pests, MOA – CABI Joint Laboratory for Bio-safety, Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing 100193, China: ² Department of Entomology, China Agriculture University, Beijing 100193, China. Corresponding author: wangzy61@163.com

The potential effects of insect-resistant, genetically engineered crops producing Cry proteins derived from Bacillus thuringiensis (Bt) on nontarget organisms, especially on predators and parasitoids that suppress pest populations, must be evaluated before such crops are commercially planted. The most important pest of maize in China and Southeast Asia is the Asian corn borer (ACB), Ostrinia furnacalis (Guenée) (Lepidoptera: Crambidae). An insect-resistant GE maize line transformed with the cry1Ac gene provides good resistance against stem borers was independently developed by China Agricultural University and has entered the pilot production stage. The current research concerns the potential effects of this GE maize line on Macrocentrus cingulum (Hymenoptera: Braconidae), a dominant and abundant parasitoid that can suppress corn borers in maize. We studied the impact of M. cingulum and Cry1Ac protein on cellular immunity and humoral immunity within hemolymph of susceptible and resistant ACB larvae. According to the results, the hemocytes in hemolymph of susceptible and resistant larvae exhibited a series of pathological changes after 2 days treated by M. cingulum and Cry1Ac protein. Total number of hemocytes changed variously when susceptible larvae were fed on Cry1Ac before or after parasitism. Hemocytes of susceptible larvae feeding on CrylAc had significantly declined cysts capacity to Sephadex A-25 microsphere an apoptosis rate was obviously higher than other treatments. Parasitism of M. cingulum inhibited the PPO activity and melanism reaction in Hemocytes of both susceptible and resistant larvae, however, lysozyme activity increased after parasitism. The differences in these immune responses may be another intrinsic mechanism for the decline of host quality.

Molecular and genomic approaches to microsporidian disease in the European honey bee, Apis mellifera

Yanping (Judy) Chen, Jay D. Evans

USDA-ARS, Bee Research Laboratory, Beltsville, MD 20705 Corresponding author: Judy.Chen@ARS.USDA.GOV

The microsporidia Nosema (a type of spore forming fungus) has been implicated in the steep global population decline of honey bees that are critical pollinators of our food crops and plants. There are two species of Nosema that have been found to infect honey bees. Nosema apis and N. ceranae. N. ceranae was the more common infection of two species. Using genomic, comparative genomics and transcriptomic approaches, we studied the genome-wide genetic differences between two Nosema species and the dynamics of host-parasite interactions at the molecular level. The comparative genomic analysis led to the identification of genes that are unique characteristics of the individual Nosema species. Transcriptional profiling of host responses to N. ceranae infection resulted in the identification of genes and pathways implicated in host-Nosema interactions. Further, we explored the potential of RNAi as a therapy for controlling Nosema disease in honey bees. Specifically, we employed an RNAi strategy to reduce the expression of a honey bee gene, naked cuticle (nkd) which is a negative regulator of host immune function. Our studies found that nkd mRNA levels in adult bees were upregulated by N. ceranae infection (and thus the parasite may use this mechanism to suppress host immune function), and ingestion of dsRNA specific to nkd efficiently silenced its expression. Furthermore, we found that RNAi-mediated knockdown of nkd transcripts in Nosema-infected bees

resulted in upregulation of expression of several immune genes encoding candidate effectors such Abaecin, Apidaecin, and Defensin-1 for the immunerelated Toll pathway, reduction of Nosema spore loads, and extension of honey bee lifespan. The results of our studies clearly indicate that silencing the host nkd gene can activate honey bee immune responses, suppress the reproduction of N. ceranae and improve the overall health of honey bees. Our study represents a novel therapeutic for honey bee Nosema disease treatment and will have positive implications for honey bee disease management practices.

Immune priming in Tenebrio molitor against Metarhizium anisopliae

9:00 0830



Jorge Contreras-Garduño

Escuela Nacional de Estudios Superiores, UNAM, Mexico Corresponding author: jcg@enesmorelia.unam.mx

Immune priming in invertebrates is their ability to mount a more vigorous immune response to a second encounter with a specific pathogen or parasite (a similar process as immune memory in vertebrates). Although at the beginning of its proposal, immune priming was not well accepted by immunologist, now there is evidence of its occurrence in different invertebrate groups such as ctenophores, porifers, nematodes, crustaceans and insects, being this later group the most studied. Now there is evidence that the immune priming protection can be species/strain-specific and can persists for a lifetime or can pass from parents to offspring. Recent research has been focused on its molecular and physiological mechanisms. However, two topics have been less studied in the literature of immune priming: the role of pathogens and its evolutionary costs. Hence, by mainly using Tenebrio molitor against the fungus Metarhizium anisopliae (MA10) as a model system, I will show that the immune primed group compared to the control group, survived for a long time period, and had more haemocytes but less ARN methylated, so I will discuss the potential link between immune response and RNA methylation. In addition, catalase seems to be implicated in host resistance because challenged insects with a more virulent derivative transformant of this fungus that overexpressed catalase (CAT) impaired immune priming. However, administering a ROS-promoter agent (paraguat) restored the protection in terms of survival of T. molitor. I propose that oxidative stress plays a key role in immune priming and discuss the possibility that parasites such as fungi may induce antioxidant production to overcome immune priming defenses. Finally, I will show that although immune priming provides protection to tenebrio larvae, at the adult stage the immune primed group compared to the control group were less preferred to mate, and this may reveal a trade-off between immune priming and reproduction. In general. I will use this host-parasite model system to discuss the potential outcomes and limitations of immune priming.

Honey bee behavioral defences against fungi

9:30 •84•

Renata Borba

University of British Columbia, Vancouver, Canada V6T 1Z4 Corresponding author: renata.borba@canada.ca

The nests of densely populated social insect colonies provide a favorable habitat for a wide range of parasites and pathogens that have evolved to overwhelm or suppress their hosts' immune defenses. In turn, insect societies have evolved remarkable abilities to counter these challenges via dynamic defense mechanisms at both the individual level (individual immunity; e.g., cell phagocytosis, immune gene up-regulation) and the colony level (social immunity). Social immunity describes colony level antiparasitic and anti-pathogenic protection characterized by collective defensive behaviors (e.g. hygienic behavior, grooming), spatial organization, and regulation of contact among nest-mates. One of these behavioral defenses in honey bees is the collection and deposition of antimicrobial plant resins in the nest as a form of cement, called propolis. When honey bee colonies nest in tree cavities, they construct a contiguous propolis layer within the rough inner walls surrounding the nest area called a propolis envelope. Previous work has shown that the presence of a propolis envelope within the nest provides benefits to adult bees' constitutive baseline immune gene expression. Additionally, recent research has shown that colonies experimentally provided with a propolis envelope have significantly lower

levels of a fungal pathogen, Ascosphaera apis, as compared to colonies without the propolis envelope. The presence of a propolis envelope on the inner walls of the nest acts as an antimicrobial layer that enshrouds the colony and benefits individual and social immune defenses of honeybees. The individual and social immunity benefits derived from propolis stem from the diverse composition of resins, each with their own complex mixtures of antimicrobial compounds. This talk will overview the range of honeybee behavioral defenses, but focus on the behavioral defense of resin collection to protect themselves against pathogenic fungi. Investigating the general and specific benefits of resin for promoting honey bees' natural defenses to pathogenic microorganisms may lead to novel and sustainable ways to improve colony health and mitigate honey bee colony loss due to disease

Marshall Room

Microbial Control Division #1

Moderators: Mary Barbercheck, Michael Brownbridge

Using NGS methods to analyze the molecular composition of CpGV isolates and making predictions for field performance



Johannes A. Jehle, Gianpiero Gueli Alletti, Annette J. Sauer, Birgit Weihrauch, Eva Fritsch, Karin Undorf-Spahn, Jörg Wennmann

Institute for Biological Control, Federal Research Center for Cultivated Plants, Julius Kühn Institute, Darmstadt, Germany Corresponding author: johannes.jehle@Julius-Kuehn.de

Cydia pomonella granulovirus (CpGV) is a highly important tool for an efficient control of codling moth (Cydia pomonella, L., CM) in organic and integrated pome fruit production. In recent years more than 40 orchards with CM populations resistant to CpGV products have been identified in Europe. By genetic analyses and resistance testing we identified three types of resistance (type I-III) which differ strongly in their susceptibility to different CpGV isolates of different genome groups A-E and their inheritance pattern. We used mapping of single nucleotide polymorphisms (SNPs) and next generation sequencing (NGS) of different CpGV isolates to correlate their molecular composition and virulence. Our identification method is based on 651 SNP positions either specific for one genome group, or specific for a combination of at least two genome groups. The vast majority (534 positions or 82%) were specific for a single genome group. The SNP distribution was used to quantify the composition of CpGV isolates containing viruses form different genome groups. These results allow predictions for field efficacy and recommendations for the design of resistance management strategies.

New Chinese isolates of Cydia pomonella granulovirus (CpGV) provide novel genetic diversity in the arms-race of resistance of codling moth



Jiangbin Fan^{1,2}, Annette J. Sauer¹, Jörg Wennmann¹, Dun Wang², Johannes A. Jehle¹

¹Julius Kühn-Institut, Federal Research Centre for Cultivated Plants, Heinrichstraße 243, 64287 Darmstadt, Germany; ²Key Laboratory of Plant Protection Resources and Pest Management of Ministry of Education, Northwest A&F University, Yangling 712100, China

Corresponding author: Johannes.Jehle@julius-kuehn.de; dunwang@foxmail.com

Application of Cydia pomonella granulovirus (CpGV) products is an efficient and safe way to control codling moth (CM) in pome fruit orchard. CM field populations, however, have evolved strategies to defend against CpGV infection, and more than 40 resistant CM populations classified into two types of resistance, sex-linked (type I) and autosomal inheritance (type II), have been identified in Europe, again causing serious economic threat to apple and pear growers. Previous research has elucidated that absence of 24 nucleotides in pe38 is the key factor involved breaking type I resistance by certain CpGV isolates. Seven new CpGV isolates were collected in different regions in China. Their virulence was determined in three different susceptible and resistant CM populations with a discriminating concentration of 5.8×10⁴ OBs ml⁻¹. Some of these isolates were breaking both type I and type II resistance. All seven isolates were Illumina sequenced and assembled using Geneious R9. Phylogenetic analyses of these isolates using the whole

genome sequences revealed considerable genetic diversity. It was observed that the isolates CpGV-ZY, -ALE, -KS1, CpGV-KS2 were grouped into CpGV genome group A containing 24 bp insertion in pe38. CpGV-JQ, -ZY2 and -WW were grouped into resistance-breaking genome type E without 24 bp insertion. CpGV-WW, however, could not kill CM with type I resistance efficiently as it would have been expected from the currently existing resistance models. These new CpGV isolates significantly extend the known genetic diversity of CpGV and provide new tools to analyze resistancebreaking mechanisms as well as for field control of CM.

Promoting microbial control as an important part of integrated pest management strategies in California

STU 8:30 •87•



Sumanth S.R. Dara¹, Suchitra S. Dara¹, Surendra K. Dara²

¹Global Agricultural Solutions, Bakersfield, USA, ²University of California Cooperative Extension, San Luis Obispo, USA Corresponding author: skdara@ucdavis.edu

Conventional pest management typically relies on chemical pesticides although entomopathogenic fungi-based formulations are available against a variety of arthropod pests. Multiple studies were conducted to promote the use of entomopathogenic fungi (EPF) in strawberry and vegetable production for both pest management and other purposes. A laboratory study that evaluated reduced rates of chemical pesticides along with Beauveria bassiana against the western tarnished plant bug, Lygus hesperus led to multiple strawberry field studies where combining or rotating EPF with chemical and other control strategies was an effective integrated pest management strategy. Another study demonstrated the compatibility of B. bassiana with several fungicides that are commonly used for disease control in strawberry. A third study showed that B. bassiana, Isaria fumosorosea, and Metarhizium brunneum have a positive impact on the growth and health of potted cabbage plants growing under water stress. These studies addressed some of the concerns of growers and demonstrated the efficacy of EPF, their compatibility with fungicides, and alternative uses in agriculture.

Sweetpotato weevil microbial control

8:45 •88•



Chad A. Keyser¹, Brooke Bissinger¹, Charles Pepe-Ranney¹, Jeffrey Davis², Milton O. Anyanga³, Sinnikka Smith¹, James Trimble³

¹AgBiome, Inc., Research Triangle Park, NC; ²Louisiana State University AgCenter, Baton Rouge, LA; ³National Agricultural Research Organization/National Crops Resources Research Institute, Kampala, Uganda Corresponding author: ckeyser@agbiome.com

AgBiome is identifying and developing a microbial solution that can be utilized to control sweetpotato weevils (Cylas spp.) in Sub-Saharan Africa. Sweetpotatoes are one of the most important agricultural crops worldwide and in Sub-Saharan Africa, are a key dietary staple that provide crucial nutrients for health. The sweetpotato weevil is the most serious insect pest of sweetpotatoes, reducing yields by 60-100% if untreated. Furthermore, they are difficult to control since all but the adult stage of the life cycle occurs within the tuberous roots. In the United States, control methods target adult weevils, employing frequent applications of chemical insecticides. However, in Sub-Saharan Africa the primary growers of sweetpotatoes are smallholder farmers who use little to no inputs, instead relying on cultural practices such as hand-weeding and hilling. Microbial control is an attractive solution, generally affording low or no exposure risk and potentially offering season-long control through inoculation of host plants. AgBiome is utilizing an innovative microbial capture and screening program which includes isolating, fully sequencing and testing bacterial isolates from the USA and Uganda for broad-spectrum coleopteran activity by screening against larval Colorado potato beetles and Western corn rootworms. Microbes with confirmed activity will then be tested against the US sweetpotato weevil, C. formicarius and later against the African species, C. brunneus and C. puncticollis. Effective control options that can easily be implemented by African sweetpotato farmers are desperately needed and an efficacious microbial control agent has potential to play a key role in protecting sweetpotato crops from the sweetpotato weevil and other important coleopteran pests.

DuPont Crop Biologicals: Harnessing beneficial microbial communities



Denny J. Bruck¹, Richard Broglie¹, Matt Ashby²

DuPont Pioneer, Trait Discovery and Optimization, ¹Johnston, IA and ²Hayward, CA corresponding author: Denny.Bruck@pioneer.com

Technological advances have enabled a deeper understanding of the complex interactions between crops, their microbial communities and the environment. The ability to deliver microorganisms or consortia of microorganisms as seed-applied products that optimize the root ecosystem systems creates new opportunities and challenges for enhancing crop productivity and sustainability in large acre field crops. One of the initial insect pest targets for the DuPont Crop Biologicals platform is the western corn rootworm, Diabrotica virgifera virgifera. Entomopathogenic fungal strains have been isolated, screened and advanced to the field. When applied to seed, these fungi significantly reduce root damage by D. virgifera virgifera and provide additive root protection to transgenic traits and chemical seed treatments.

Helping growers to get the best out of biopesticides: The UK AMBER project



Dave Chandler¹, Jude Bennison², Clare Butler Ellis³, Roma Gwynn⁴ Gill Prince¹, Rob Jacobson⁵, Mark Ramsden², Erika Wedgwood²

¹Warwick Crop Centre, University of Warwick UK; ²RSK ADAS, Cambridge UK; ³Silsoe Spray Applications Unit, Silsoe UK; ⁴Rationale Biopesticides, Duns, Scotland UK; ⁵Rob Jacobson Consultancy Ltd, Bramham UK. Corresponding author: dave.chandler@warwick.ac.uk

Globally, the commercial biopesticides industry is growing rapidly, with an increasing number of biopesticide products reaching the marketplace. In order for the biopesticides industry to make this growth sustainable, it is essential that farmers and growers understand how to use biopesticides in the most effective ways. In surveys we have conducted in the UK, we found that growers perceive biopesticides to be unreliable and difficult to use, and this is acting as a block to uptake. AMBER (Application and Management of Biopesticides for Efficacy and Reliability) is a 5 year project instigated and funded by UK growers of protected edible and ornamental crops designed to address this issue. The aim of AMBER is to have UK growers adopting new practices that have been demonstrated to improve the performance of individual biopesticide products within commercial integrated pest and disease management programmes. The project started in 2016 where we observed growers using selected microbial biopesticides on 5 different crop types as part of commercial crop production. This highlighted several opportunities for improving biopesticide performance, in particular by making changes to application practices based on better understanding of the optimum conditions required for good biopesticide performance. This included factors such as water volume, quantity of product, as well as improving awareness of the fundamentals of good application practice (choice of nozzles, tank cleaning etc.). Growers found some of the technical guidance provided by biopesticide companies confusing and difficult to understand. Our work also identified the need for simple models of the effect of biopesticides on pest growth, and better knowledge of effective doses and persistence of activity, to enable growers / agronomists to fine tune the timing and frequency of applications.

Seeing through the fog: Can we use LVM sprayers to improve application efficiency and performance of biopesticides?

9:30 •91•

Michael Brownbridge, Bernhardt Steinwender, Taro Saito

Vineland Research and Innovation Centre. 4890 Victoria Ave N. Vineland Sta., ON Canada LOR 2E0 Corresponding author: michael.brownbridge@vinelandresearch.com

Low volume and ultra low volume spraying (also known as fogging) is considered a more efficient method for pesticide application in greenhouses compared to classic hydraulic or mechanical techniques. Automated systems require less applicator time to treat a large area while concurrently minimizing risks of worker exposure. In addition, manufacturers of low volume equipment claim that the sprays cover plant surfaces more evenly,

reach areas that are usually missed (especially the underside of leaves), and reduce waste and potential contami-nation by avoiding run off. As living organisms, microbials (e.g., bacteria and fungi) are more sensitive to abiotic factors than chemicals. High temperature and pressure can negatively affect these microbes, yet these factors are integral to the transformation of microbial suspensions into spray droplets ranging from ~10-70 μm in diameter, depending on the spray system used. The utility of two low volume mist (LVM) sprayers for application of two fungal bioinsecticides Met52 EC (Metarhizium brunneum) and BotaniGard WP (Beauveria bassiana), and the biofungicide, Cease® (Bacillus subtilis), was assessed. Successful insect or disease control requires that sufficient viable conidia or spores reach the target pest, either at the time of application or via pick-up from the leaf surface. The degree to which the fogging process (cold and thermal) affects the viability of microbial biopesticides therefore has significant implications for efficacy. Today growers can choose from a range of machines for the application of microbial products. Results will show how different LVM sprayers affect the viability of conidia or spores contained in microbial biopesticides, and the importance of selecting the right equipment for their application.

Development of CO₂-releasing formulations for the control of soil-borne insect pests

Anant V. Patel¹, Pascal Humbert¹, Stefan Vidal², Michael Przyklenk³, Elisa Beitzen-Heineke³, Wilhelm Beitzen-Heineke³

¹ Bielefeld University of Applied Sciences, Faculty of Engineering Sciences and Mathematics, Bielefeld, Germany; ²Agricultural Entomology, Department for Crop Sciences, Georg-August University Goettingen, Goettingen, Germany;
³BIOCARE GmbH, Einbeck, Germany

Corresponding author: anant.patel@fh-bielefeld.de

Since effective synthetic chemicals used by conventional farmers in the past to control various soil-borne insect pests of worldwide relevance, e.g. wireworms or the western corn rootworm, have been phased out or are no longer marketed, the control options targeting these pests are strongly limited. Therefore, alternative control strategies based on biological control agents are urgently needed. Root-feeding herbivores generally use CO₂ gradients in soil to locate their host plants. Given the property of baker's yeast (Saccharomyces cerevisiae) to convert sugars into CO2, it finds application in biological pest control, especially in the form of so-called Attract-and-Kill co-formulations. However, one major obstacle is to extend the CO2 release over several weeks to obtain a long-lasting protection. S. cerevisiae metabolizes only mono- or disaccharides which cannot be retained by means being suitable for the encapsulation of biological control agents. A high molecular weight carbon source such as starch is retained by the polymer matrix, but S. cerevisiae is lacking the enzymatic equipment for degradation. To circumvent this problem, one promising approach is the coencapsulation of an amylolytic enzyme (Humbert, 2017). Amyloglucosidase was co-immobilized with high encapsulation efficiency of 86.5±16%. The CO₂ release was shown to be dependent on the concentration of the coencapsulated enzyme with a CO₂-production of 0.35 mL g⁻¹ h⁻¹ at 18 °C for a moderate concentration of 0.1 AGU/g. Besides the enzyme concentration, further factors influencing the CO₂ release from beads were revealed, e.g., incubation temperature, starch concentration, pH or bead diameter. In order to clarify the biochemical reactions taking place within the beads, scanning electron microscopy (SEM) and MALDI-ToF mass spectrometry imaging (MSI) were conducted. SEM micrographs proved the degradation of corn starch within the beads and MSI showed that S. cerevisiae cells are able to take up nearly all of the released glucose within the beads. The innovative CO₂releasing beads will pave the way towards novel attract-and-kill strategies in pest control. REF: Humbert, P., Vemmer, M., Giampà, M., Bednarz, H., Niehaus, K., & Patel, A. V. (2017). Co-encapsulation of amyloglucosidase with starch and Saccharomyces cerevisiae as basis for a long-lasting CO₂ release. World Journal of Microbiology and Biotechnology, 33(4), 71.

10:00-10:30 am

Coffee Break

TUESDAY, 10:30 am - 12:30 pm

Theater

Virus Division #2

Moderator: Peter J. Krell, Lorena Passarelli

Selected promoter expression analysis of Anticarsia gemmatalis multiple nucleopolyhedrovirus during infection of permissive, semipermissive and nonpermissive cell lines

10:30 •93•

Fabricio da Silva Morgado¹, Daniel Mendes Pereira Ardisson-Araújo², **Bergmann Morais Ribeiro**¹

¹ILabortório de Baculovirus, Departamento de Biologia Celular, Instituto de Ciências Biológicas, Universidade de Brasília, Brasília, Brasíli²Laboratório de Virologia de Insetos, Departmento de Bioquímica e Biologia Molecular, Universidade Federal de Santa Maria. Santa Maria. Brasíl

Corresponding author: bergmann.ribeiro@gmail.com

The activation of baculovirus gene promoter during infection of insect cells follows a transcriptionally controlled sequence of gene expression. Depending on the cell type, this sequence of promoter activation may be disrupted by antiviral cellular defense mechanisms and interfere with viral progeny formation. In this work, the activity of different promoters (*ie1*, *gp64*, *lef-1*, *vp39*, *p6.9* and *polh*) of the Anticarsia gemmatalis multiple nucleopolyhedrovirus isolate 2D (AgMNPV) was assessed during infection of permissive, semipermissive and nonpermissive cell lines in real-time using the firefly luciferase reporter gene. We have characterized in rich detail the AgMNPV promoters in permissive cell lines and revealed differential profiles of expression in cells with limited permissivity that correlate well with limitations in viral DNA replication. Semipermissive and nonpermissive cell lines presented delays and restrictions in late and very late promoter expression. Cells undergoing apoptosis did not inhibit late gene expression; however, viral progeny formation was severely affected.

Functional analysis of the N terminus of Autographa californica multiplenucleopolyhedrovirus DNA polymerase

94•

Guoqing Chen¹, Yang Fang¹, Wu Lijuan¹, Peter J. Krell², **Guozhong Feng**¹

¹State Key Laboratory of Rice Biology, China National Rice Research Institute, Hangzhou, China; ²Department of Molecular and Cellular Biology, University of Guelph, Guelph, ON, Canada. Corresponding author: fengguozhong@caas.cn

Autographa californica multiple nucleopolyhedrovirus (AcMNPV) DNA polymerase (DNApol) plays an important role in viral DNA replication, the N terminus (residues 1 to 186) is present in all baculovirus DNApols. However, its functional role has not yet been characterized. Here we report a functional analysis of the N terminus of AcMNPV DNApol. We truncated the N-terminus of DNApol to varying lengths and reinserted them into a nulldnapol AcMNPV bacmid with a green fluorescent protein (GFP) reporter. Although these truncation repair mutants rescued viral DNA replication and infectious virus production, the level of viral DNA replication and production of virus and occlusion bodies were compromised. Further bioinformatic analysis showed that the first 64 amino acids at the N termini contain a conserved α (-helix)- β (-sheet)- β - β secondary structure motif and four other conserved sequence motifs are located in further downstream between aa 67 to 186. Multiple alanine point substitutions in the $\alpha\beta\beta\beta$ secondary structural region or the other four sequence motifs in AcMNPV DNApol impaired viral DNA replication and infectious virus yield and occlusion body production. Taken together, our result demonstrated that N terminus of baculovirus DNApol is important in efficient viral DNA replication and production of both infectious virus and occlusion bodies.

Mutagenesis of the carboxyl region of AcMNPV lef-2 and its consequences for protein function

STU 11:00 •95•

Carina Bannach, Clare Allen, Linda A. King, Robert D. Possee

Department of Biological and Medical Science. Oxford Brookes University, Oxford, UK Corresponding author: caring.bannach-2015@brookes.ac.uk

The late expression factor 2 (lef-2) is an early viral gene involved in AcMNPV DNA synthesis and was originally identified by a transient in vitro replication assay. However, later work suggested that LEF-2 was additionally required for viral very late gene expression. Using BrdU mutagenesis, a virus designated very late deficiency 1 (vld1) was isolated, which was unable to express viral p10 and polh. The original mapping and sequencing of vld1 revealed a mutation at AcMNPV genome position 3620 within lef-2 that resulted in an amino acid change from aspartic acid to asparagine (LEF-2 $^{\rm D178N}$). This was thought to account for the altered phenotype of VLD1 comprising reduced budded virus production and the lack of very late gene expression. Recent resequencing of lef-2 within VLD1 identified a second mutation at nucleotide position 3548 that resulted in an alanine to threonine change (LEF-2^{A154T}). When we attempted to construct a recombinant AcMNPV with this mutation to examine its function it was impossible to isolate an infectious virus. The alanine 154 is conserved in most baculovirus LEF-2's so may have a critical role in its function. However, the same mutation within vld1 was permissive for virus replication. This result suggests that a combination of *lef-2* mutations (LEF-2^{A154T; D178N}) in VLD1 is acceptable for protein function but that LEF-2^{A154T} is not. Both of these mutations are located in the carboxyl region of LEF-2. We also examined the role of highly conserved cysteine residues in this portion of the protein and found variable requirements for their function in virus replication. Insect cells infected with these virus mutants showed different reductions in intracellular viral DNA levels as examined by qPCR, budded virus production and viral very late gene expression compared to wild type infected cells. The data indicated that different conserved amino acids at the LEF-2 C-terminus play a crucial role during the AcMNPV replication cycle.

A nucleocapsid assembly-essential cis-acting element in the Autographa californica nucleopolyhedrovirus ac83 gene

Zhihong Huang, Mengjia Pan, Silei Zhu, Hao Zhang, Wenbi Wu, Meijin Yuan, Kai Yang

State Key Laboratory of Biocontrol, Sun Yat-sen University. Guangzhou, 510275, China Corresponding author: yangkai@mail.sysu.edu.cn

The mechanism of baculovirus nucleocapsid assembly remains unclear. Previous studies have shown that deletion of the ac83 gene of Autographa californica multiple nucleopolyhedrovirus (AcMNPV) blocks viral nucleocapsid assembly. Interestingly, the ac83-encoded protein Ac83 was not detected as a component of the nucleocapsid in some studies, implying a particular role for ac83 in nucleocapsid assembly that may be independent of its protein product. To examine this possibility, Ac83 synthesis was disrupted by insertion of a chloramphenicol resistance gene into its coding sequence (with only nt 361 to 464 deleted) or by deleting its promoter and translation start codon. Both mutants produced progeny viruses normally, indicating that the Ac83 protein is not essential for nucleocapsid assembly. Subsequently, complementa-tion assays showed that the production of progeny viruses required the presence of ac83 in the AcMNPV genome instead of its presence in trans. Therefore, we reasoned that ac83 is involved in nucleocapsid assembly via an internal cis-acting element, which was named the nucleocapsid assembly-essential element (NAE). The NAE was identified only in alphabaculoviruses and have a conserved positional relationship with another essential cis-acting element (the conserved nonprotein-coding element) which was recently identified. The identification of the NAE may help to connect the data of viral cis-acting elements and related proteins in the baculovirus nucleocapsid assembly, which is important for elucidating DNA-protein interaction events during this process.

The nucleocapsid assembly-essential element of Autographa californica multiple nucleopolyhedrovirus is located in nt 1651 to 1850 of ac83

STU 11:30

Silei Zhu, Zhihong Huang, Wenbi Wu, Meijin Yuan, Kai Yang

State Key Laboratory of Biocontrol, Sun Yat-sen University, Guangzhou 510275, China Corresponding author: yangkai@mail.sysu.edu.cn

In the recent study, we found that the presence of ac83 gene in the Autographa californica multiple nucleopolyhedrovirus (AcMNPV) genome, rather than the synthesis of Ac83 protein, is essential for progeny virion production. A cis-acting element, the nucleocapsid assembly-essential element (NAE), was further identified in the ac83 coding sequence. Our previous study also demonstrated that viral replication was completely blocked by deletion of nt 1651 to 2011 of ac83, while repair of nt 1351 to 1800 of ac83 could partially rescue viral viability, indicating that the NAE is located in this region. In the present study, a series of ac83-truncated viruses were generated, which had truncated fragments of nt 1351 to 2011 of ac83. It turned out that the viability of these ac83-truncated viruses was essentially unaffected. Therefore, we reasoned that the NAE is approximately 200 bp in length, located in the overlapping region (nt 1651-1850 of ac83) of these truncated fragments. Sequence analyses revealed that the NAE is conserved among alphabaculo-viruses and characterized by multiple AT-rich regions. Additionally, a conserved motif (ATATACTAC or similar sequences) was observed in both the NAE and CNE, another cis-acting element. Whether this motif is related to their functions remains to be studied. Electrophoretic mobility shift assays (EMSA) detected protein(s) that bound to the NAE in infected cells rather than uninfected cells. The results of EMSA corroborated that the NAE is a cis-acting element and indicated the NAE interacts with some viral protein(s) or virus-induced cellular protein(s).

A global proteomic analysis of the substrates of a baculovirus sulfhydryl oxidase

Samantha A. Warnecke, Kathlyn L. Gomendoza, Elizabeth M. Martino, A. Lorena Passarelli

Division of Biology, Kansas State University, Manhattan Kansas, U.S.A. Corresponding author: lpassar@ksu.edu

Virions have a structural blueprint and an enclosed architecture required to protect the viral genome and transfer infection between cells. Changes in capsid or capsid-associated protein stability and/or conformation occur at specific stages during virus multiplication. Disulfide bonds within virion proteins can provide structural determinants required for virus replication. Baculoviruses are one of only a few types of viruses that encode a conserved sulfhydryl oxidase gene. This viral enzyme is able to oxidize protein thiol groups to disulfide bonds. The formation of disulfide bonds usually occurs in the endoplasmic reticulum, a site with an oxidizing milieu, as proteins are trafficked to the cell membrane. In contrast, oxidation reactions are not favored in the nucleus or cytoplasm of cells, where the environment is reducing. Autographa californica multiple nucleopolyhedrovirus (AcMNPV) encodes a functional sulfhydryl oxidase, Ac92. Deletion of ac92 affects both virion types that are produced during virus replication, resulting in noninfectious budded virus and singly, rather than multiply, enveloped occlusion-derived virions. Using an in vitro assay, we found proteins in the budded virus of AcMNPV that contain disulfide bonds. To identify the virion proteins containing disulfide bonds, we utilized an unbiased proteomic assay in which proteins containing thiol-reactive groups were alkylated and differentially separated using two-dimensional gel electrophoresis. Mass spectrometry was subsequently used to identify modified proteins. Continuing these studies will provide insight into the mechanisms of virus assembly and virion stability in the budded and occlusion-derived virions.

Autographa californica multiple nucleopolyhedrovirus ac51 gene encodes a nucleocapsid protein and is required for efficient egress of nucleocapsids to form budded virions 12:00 •99•

Jianxiang Qiu, Zhimin Tang, Yi Cai, Wenbi Wu, Meijin Yuan, Kai Yang

State Key Laboratory of Biocontrol, Sun Yat-sen University, Guangzhou, 510275, China Corresponding author: vanakai@mail.svsu.edu.cn

Alphabaculoviruses are lepidopteran-specific nucleopolyhedroviruses and the anterograde transport of their nucleocapsids is rarely known. In the present study, an alphabaculovirus unique gene, Autographa californica multiple nucleopoly-hedrovirus (AcMNPV) orf51 (ac51), was identified to be involved in anterograde transport of AcMNPV nucleocapsids. Our study indicated that ac51 is regulated by both early and late promoters and maintains its transcripts from the early to the late stage of infection. ac51 product (Ac51) was detectable from 12 hours post-infection (h p.i.) to 72 h p.i. Ac51 was distributed predominantly in the nuclei of infected cells and also locates surrounding the nuclear membrane in the cytoplasm. Moreover, Ac51 is a nucleocapsid protein of budded virions (BVs). Upon ac51 deletion, infectious BV production by 96 h p.t. was reduced by approximately 1,000fold compared with that of wild-type AcMNPV. Neither viral DNA synthesis nor viral gene expression were affected. Transmission electron microscopy showed that virogenic stroma formation, distinguishable nucleocapsid assembly and the formation of occlusion-derived virions and occlusion bodies were not impaired by ac51 deletion. However, in vAc51KOtransfected cells, the number of nucleocapsids in the nucleus was comparable with that of wild-type AcMNPV, while BV production in the supernatants in a virus life cycle was dramatically decreased, indicating a role for ac51 in the egress of nucleocapsids to form BVs. Taken together, our results support the conclusion that ac51, encoding a nucleocapsid proteins, is not involved in the nucleocapsid assembly, but plays an important role in the egress of nucleocapsids to form BVs.

Deciphering the genetic factor for morphology of granulovirus occlusion body



Keiko Tsuruta¹, Jörg T. Wennmann², Maki N. Inoue¹, Yasuhisa Kunimi¹, Johannes A. Jehle², Madoka Nakai¹

¹ Tokyo University of Agriculture and Technology, Tokyo, Japan; 2 Institute for Biological Control, Federal Research Center for Cultivated Plants, Julius Kühn Instiute, Darmstadt, Germany Corresponding author: madoka@cc.tuat.ac.jp

In 1999, we found a granulovirus (GV) that produced an unusual morphology of occlusion bodies (OBs) in Adoxophyes sp. (Lepidoptera: Tortricidae) larva in a tea field in Miyazaki Prefecture, Japan. This isolate is a morphological mutant of Adoxophyes orana granulovirus, AdorGV (AdorGV-M) with 99.7% nucleotide sequence identity the English isolate of AdorGV (AdorGV-E). Whereas AdorGV-E OBs have a typical ovo-cylindrical shape with a size of 0.3 x 0.5 μm , and each OB contains one virion, AdorGV-M has cuboidal-shaped OBs with a diameter of 0.8-2.2 $\mu m.$ In this study, we investigated the potential genetic factor responsible for difference of the OB morphology between the two isolates. Based on sequence comparison between AdorGV-E and AdorGV-M, five candidate genes were initially identified. Among them, pep-p10 was considered to be the most likely target. Because studying gene functions of AdorGV is hampered by the lack of susceptible cell lines and bacmid technology, we used Cydia pomonella granulovirus (CpGV) and Cp14R cell line as model to generate pep-p10 recombinants. A pep-p10 knockout mutant (bacCpGV $^{hsp\text{-}GFP}\Delta pep\text{-}p10$) was constructed from CpGV bacmid (bacCpGV^{hsp-GFP}) and used for cell transfection. The results revealed that both bacCpGV^{hsp-GFP} and the knockout mutant bacCpGV^{hsp-GFP}Δpep-p10 produced viable budded viruses (BV). The infected Cp14R cells were observed under transmission electron microscopy (TEM), and the results showed that both bacCpGV $^{\text{hsp-GFP}}$ and bacCpGV $^{\text{hsp-GFP}}\Delta \text{pep-p10}$ produced also OBs, however with completely different morphology. BacCpGV^{hsp-GF} produced typical normal-sized, ovo-cylindrical OBs, whereas $\mathsf{bacCpGV}^\mathsf{hsp}$ $^{r}\Delta$ pep-p10 produced OBs with a cuboidal shape and a diameter of 0.6-1.9 μm . We propose that deletion of $pep\mbox{-}p10$ resulted in the generation of

cuboidal OBs and that this gene is most likely involved in the OB morphology of GVs.

The Forum

NEMATODES DIVISION SYMPOSIUM

Nematode Omics

Organizer and Moderator: Patricia Stock

Looking for nematode biology in tera-bases of sequence data

10:30 •101•



Makedonka Mitreva^{1,2}

¹Division of Infectious Diseases, Department of Medicine: Washington University School of Medicine; St. Louis, Missouri; ²McDonnell Genome Institute, Washington University in St. Louis, St. Louis, Missouri Corresponding author: mmitreva@wustl.edu

There has been constant increase in the number of nematode genomes published (and in progress) which is a trend that correlates to ever-improving molecular techniques for obtaining biological material, performing DNA extraction and library construction, and predominantly to ever-evolving sequencing chemistry and instrumentation, as well as to decreased sequencing costs. These nematode genomes provide an invaluable resource to enable development of postgenomic tools to investigate the biology of parasitic nematode diseases and accelerate discovery for the development of anthelminthics, vaccines and diagnostics. However, identifying good candidate targets in high throughput data is challenging and requires an integrated multi-omics approach (including nematode genomics, transcriptomics, functional genomics, proteomics, interactomics etc), and innovative bioinformatic analysis. Here I present our progress on knowledge based drug target identification, prioritization and experimental validation exploring metabolic chokepoints as targets. We first reconstruct metabolic pathways for 56 nematode species, using ~700 unique enzymes encoded by their genomes. Comparative genomics identified phylogenetically conserved and restricted pathways that underline differences in life-cycle and mode of parasitism. A total of 185 enzymes are pan-phylum conserved, and 88 of them are conserved chokepoints. The prioritization of the chokepoints for experimental validation included level of conservation in nematodes and host, orthology in drug target databases, RNAi phenotype, expression profile across tissue and developmental stages, function in multiple pathways of interest, gene copy number, etc. The chokepoints were linked to inhibitors in drugbanks and subsequently, the predictions are being validated in multiple nematode species. Preliminary work on a broadly conserved chokepoint enzyme CPT (carnitine palmitoyl transferase) resulted in compiling of a small library of 22 CPT inhibitors that we screened against five parasitic nematode species spanning the phylum Nematoda. A worm motility assay identified 9 hits, including 4 with potential for broad applicability across clades since the are effective against gastrointestinal and filarial species. Parasite-specific sequence variations are now explored to increase selectivity to the parasites over the host ortholog. The prioritized drug targets and drug-like compounds have potential to expedite the discovery of new anthelmintic drugs with broad-spectrum efficacy.

Lateral gene transfer from bacteria to nematodes ... and beyond!

11:00 •102•

Julie C. Dunning Hotopp

Institute for Genome Sciences, Department of Microbiology and Immunology, Greenebaum Comprehensive Cancer Center, University of Maryland, Baltimore, MD, USA Corresponding author: jdhotopp@som.umaryland.edu

Lateral gene transfer (LGT) is the transfer of DNA between divergent organisms. In 2007, we demonstrated that bacterial DNA from Wolbachia endosymbionts is found in the genomes of numerous arthropods and filarial nematodes. Overall, LGT is increasingly described as occurring from bacteria to animals, but particularly Wolbachia and its arthropod and nematode hosts. At least 10.6% of the Wolbachia genome has been transferred to its Brugia malayi nematode host yielding >0.5 Mbp of Wolbachia sequence in the nematode's 89 Mbp genome. Most LGTs are gene fragments or otherwise degenerate sequences with respect to protein coding potential, and many

are present multiple times in the B. malayi genome. A handful of LGTs contain full length Wolbachia genes with the potential to encode full-length functional proteins. At least four LGTs have evidence of life stage-specific regulation of transcription at levels like other nematode transcripts, raising the possibility that they might be functional. In Drosophila ananassae, a remarkable 2% of the nuclear genome is of bacterial origin, including 20% of the known material on a *Drosophila* autosome. In combination, these results suggest that extensive amounts of bacterial DNA may reside in invertebrate nuclear genomes. Based on these findings, we hypothesized that there could even be integration of bacterial DNA in human somatic cells. Using publicly available sequence data from the human genome project, the 1000 Genomes Project, and The Cancer Genome Atlas (TCGA), we have presented evidence that bacterial DNA integrates into the human somatic genome through an RNA intermediate and that such integrations are detected more frequently in (a) tumors than normal samples, (b) RNA than DNA samples, and (c) the mitochondrial genome than the nuclear genome. As such, LGTs on invertebrate genomes may serve as a model for understanding insertional mutagenesis of somatic vertebrate genomes, including the human genome.

Transcriptional evidence of symbiont-modulated metabolism in the entomopathogenic nematode Steinernema carpocapsae (Nematoda: Steinernematidae) 11:30 •103•

BF Peterson¹, JG McMullen², MN Yousefelahi³, SP Stock^{1,4}

¹Center for Insect Science, University of Arizona^{; 2}Department of Entomology, Cornell University; ³ Chemistry and Biochemistry, University of Arizona; ⁴ Department of Entomology, University of Arizona Corresponding author: bfpeterson@email.arizona.edu

Steinernema carpocapsae nematodes, and their bacterial symbionts, Xenorhabdus nematophila, are lethal insect pathogens. Outside the insect host, symbiont-colonized infective juveniles (IJs) live in the harsh soil environment for extended periods of time without feeding. Therefore, nutrients accumulated before leaving the insect cadaver are the only food source available to IJs and their bacterial symbionts. We hypothesized that X. nematophila colonization causes a reallocation of these nutrients and impacts their utilization in IJs. To address this, we examined S. carpocapsae IJs from three rearing conditions: 1) in insecta with X. nematophila, 2) in vitro on lawns of X. nematophila, and 3) in vitro aposymbiotic on rich medium. Resulting IJs were subjected to standard RNAseq and differential gene expression analysis protocols. Our findings emphasize the influence that \boldsymbol{X} . nematophila has on the IJ metabolism; genes related to carbohydrate, lipid, and protein metabolism, were differentially regulated with symbiont colonization. We also measured total lipid, protein, and glycogen content across rearing conditions and assayed catabolism of potentially important disaccharides, like maltose. These results support our hypothesis that X. nematophila influences S. carpocapsae IJ metabolism, with the unanticipated finding that carbohydrate utilization may be a key factor for IJ longevity and symbiont provisioning.

The mechanisms of developmental plasticity: from switch genes and epigenetics to the interplay of organisms and their environment

12:00 •104•

Ralf J. Sommer¹

¹Max Planck Institute for Developmental Biology, Department for Integrative Evolutionary Biology, Corresponding author: ralf.sommer@tuebingen.mpg.de

Developmental plasticity is increasingly recognized as primary mechanism for the emergence of novelty. However, molecular mechanisms underlying this phenomenon remain elusive. The nematode Pristionchus pacificus exhibits developmental plasticity for its mouth-form and feeding strategies. Individuals develop one of two alternative mouth-forms, a predatory eurystomatous (Eu, wide-mouthed) or a bacteriovorous stenostomatous (St, narrow-mouthed) form. Using genetic screens, we have identified developmental switch genes that regulated plasticity. eud-1 mutants are all-St, whereas mutants in the downstream nuclear-hormone-receptor nhr-40 are all-Eu. More recent work indicates that eud-1 expression is under epigenetic control involving the histone acetyltransferase Isy-12, which acts through an antisense RNA at the eud-1 locus itself and up-regulates eud-1

expression. Here, we present our most recent molecular findings on i) the molecular mechanisms underlying plasticity, ii) the interaction of this regulatory network with the environment and iii) first insight into the involvement of small RNAs and Argonaute proteins.

Roosevelt Room

Fungi Division #2

Moderators: Nemat Keyhani, Drauzio E.N. Rangel

Light during mycelial growth produce conidia of Metarhizium robertsii with increased stress tolerance, germination speed, and virulence

10:30 •105•

Ariel de Souza Oliveira, **Drauzio Eduardo Naretto Rangel**

Instituto de Patologia Tropical e Saúde Pública. Universidade Federal de Goiás, Goiânia, GO 74605-050, Brazil. Corresponding author: drauzio@live.com

Light conditions during fungal growth are known to cause several physiological adaptations in conidia. In this study, conidia of the entomopathogenic fungi Metarhizium robertsii (ARSEF 2575) were produced on: 1) potato dextrose agar (PDA) medium in the dark; 2) PDA medium under white light (4.98 W m⁻²); 3) PDA medium under blue light (4.8 W m⁻²); 4) PDA medium under green light (2.2 W m-2); 5) PDA medium under red light (2.8 W m⁻²); and 6) minimum medium (MM = Czapek medium without sucrose) or MM supplemented with 3% lactose (MML) in the dark. The conidial production for each treatment as well as the speed of conidial germination, stress tolerance, and virulence to the insect *Tenebrio mollitor* (Colleoptera: Tenebrionidae) were studied. For stress tolerance, the conidial suspensions of each treatment were exposed from 0 (control) and 100 to 210 minutes (with increments of 10 minutes) to an irradiance of 0.60 W/m². After irradiation, the plates were incubated for 48 hours in the dark at 26 °C. For the osmotic stress, the conidial suspensions of each treatment were dropped on PDA medium supplemented with KCl concentrations of 0 (control) 1.0, 1.3, 1.4, 1.5, 1.9, and 2.1 M, and the germination was counted after 24 h. Conidia produced on MM in the dark were the most tolerant to both UV radiation and osmotic stress. Conidia produced under blue light were the second most tolerant to UV radiation, followed by the UV tolerances of conidia produced under visible light. Conidia produced under green or red lights were the least UV tolerant and less tolerant than conidia produced in the dark. For KCl tolerance, conidia produced under visible or blue light were the second most tolerant, followed by conidia produced under green light. Again, conidia produced under red light were the least tolerant to KCl. The fungus grown under blue light produced more conidia than the fungus grown in the dark. The conidial production of the fungus grown under white or red light was similar to the conidial production in the dark. The MML afforded the least conidial production. Conidia produced on MML or on PDA medium under white or blue light germinated faster than conidia produced on PDA medium in the dark or under red light. Conidia produced on MML or PDA medium under white light were also more virulent than conidia produced on any other treatment. In conclusion, the white and blue light treatments during mycelial growth produced conidia with increased stress tolerance, faster germination speed, and were more virulent to the insect, with similar result as conidia produced on MM or MML. Support: FAPEG, CNPq, FAPESP.

Over-expression of genes may reveal increased tolerance against UV-B radiation in Metarhizium spp. conidia produced on culture medium supplemented with riboflavin (vitamin B2)

STU 10:45 •106•

Ronaldo A. Pereira-Junior^{1,2*}, Carla Huarte-Bonnet^{1,2}, Flávia R. S. Paixão^{1,2}, Christian Luz¹, Drauzio E. N. Rangel¹, Donald W. Roberts³, Nicolás Pedrini², Éverton K. K. Fernandes¹

¹Instituto de Patologia Tropical e Saúde Pública Universidade Federal de Goiás, Goiânia, Brasil; ²Instituto de Investigaciones Bioquímicas de La Plata, Facultad de Ciencias Médicas, UNLP, La Plata, Argentina (CONICET-UNLP), Department of Biology, Utah State University, Logan, UT, USA. Corresponding author: pereirajunior.ra@outlook.com The success of entomopathogenic fungi is limited by their vulnerability to abiotic stresses, including ultraviolet radiation (UV) in sunlight, viz. UV-A and UV-B, which can significantly reduce the viability and delay germination of conidia. In an effort to protect conidia from solar irradiation, we evaluated the effect of adding riboflavin to culture medium for conidial production on the tolerance of these conidia to UV-B radiation, as well as the expression of photolyase (Phr), laccase (Lcc), and polyketide synthase (Pks) genes in Metarhizium acridum (ARSEF 324) and M. robertsii (ARSEF 2575). Conidia of both isolates were produced on potato dextrose medium without riboflavin (PDA) or PDA supplemented with riboflavin (PDA + Rb) at different concentrations and subsequently suspended in Tween 80[®] or riboflavin solution. Conidia were also produced on PDA medium supplemented with yeast extract (PDAY), which contains complex B vitamins. The conidia were exposed to UV-B (3.9, 5.46 or 6.24 kJ.m⁻²), visible light (16 Klux) for 2 hours, or to UV-B followed by visible light exposure. Relative UV-B tolerance was determined by comparing percent germination of conidia on the various media with and without UV exposure. Conidia from both Metarhizium isolates produced on PDA + Rb, regardless its concentration, or PDAY were considerably more tolerant to UV-B than conidia produced on PDA medium without supplement. The conidial tolerance of Metarhizium isolates to UV-B did not change when conidia were suspended in riboflavin solution, suggesting that these fungi need to produce and accumulate metabolites inside the conidia to confer photoprotection. The expression of MaLcc3 and MaPks2 for M. acridum, as well as MrPhr2, MrLcc1, MrLcc2, and MrLcc3 for M. robertsii was higher when the isolates were cultivated on PDA + Rb and exposed to UV-B followed by exposure to visible light, or exposed to only visible light, than conidia produced on PDA medium alone. The addition of riboflavin to substrates used for mass production of Metarhizium spp. and consequently enhancing the tolerance of conidia against UV-B radiation may be a viable approach to improving the effectiveness of these fungi in biological control programs.

RNAi-mediated knock down of insect immune-related genes in Tenebrio molitor increases entomopathogenic fungal virulence in Beauveria bassiana JEF-007



Mi Rong Lee, Yi-Ting Yang, Se Jin Lee,

Sihyeon Kim, Jong Cheol Kim, Jae Su Kim*

Department of Agricultural Biology, College of Agricultural & Life Sciences. Chonbuk National University, Jeonju 561-756, Korea Corresponding author: jskim10@jbnu.ac.kr

Toll signaling pathway is responsible for defense against Gram-positive bacteria and fungi. Gram-negative binding protein 3 (GNBP3) 1, $3-\beta$ -D-glucan pattern recognition proteins (GRP) has strong relationship for fungal cell wall component (β-1, 3-glucan), which can move prophenoloxidase (proPO) cascade and cause Toll signaling pathway. Myeloid differentiation factor 88 (MyD88) is essential adaptor protein used by signal transduction pathways of the Toll-like receptor (TLR). In this study, we monitored the response of TmGNBP3 and TmMyD88 genes of the insect immune system against Beauveria bassiana JEF-007 in Tenebrio molitor using RT-PCR. TmGNBP3 was up-regulated after fungal infection. To better understand the roles of Toll signaling pathway in T. molitor immune system, TmGNBP3 and TmMyD88 were knocked down by RNAi. The results of RNAi showed that target gene expressions were decreased at 2 days post-dsRNA injection, and dramatically reduced at 6 days post-dsRNA injection. Therefore, T. molitor was compromised by B. bassiana JEF-007 at 6 days post-dsRNA injection. Silencing of the TmMyD88 and TmGNBP3 resulted in reducing the resistance of the host to fungal infection. These results indicate that not only TmGNBP3 but also TmMyD88 is required in T. molitor for survival against B. bassiana JEF-007, and the knock down of these genes possibly contributes to the enhanced fungal insecticidal activity.

Drosophila usage of chemical cues in removing fungus Beauveria bassiana from the body surface

Aya Yanagawa¹, Marie-Ange Chabaud², Tomoya Imai¹, Toshimitsu Hata¹, Tsuyoshi Yoshimura¹, Frederic Marion-Poll^{3,4}

¹RISH, Kyoto University, Uji city, Japan; ² UMR Physiologie de l'Insecte : Signalisation et Communication, INRA Centre de Versailles, France; 3 UMR Evolution, Génomes, Comportement, Ecologie, CNRS, IRD, Univ Paris-Sud, Université Paris-Saclay, France; 4 AgroParisTech, France Corresponding author: ayanagawa@rish.kyoto-u.ac.jp

Grooming behavior serves to remove harmful microbes from an insect's body surface. Although this role has not been reported clearly in solitary insects yet, it is well known that solitary insects like Drosophila melanogaster dissipate the attached foreign organisms by self-grooming behavior. In this study, we examined whether grooming serves to protect flies by reducing the risk of fungal infection in D. melanogaster. First, we confirmed that fungal conidia were removed by grooming. Fruit flies were treated with harmful entomopathogenic fungus, Beauveria bassiana. Then the removal of fungal conidia from the insect's body surface through grooming was quantified through microscopic observation. Second, the role of insect perception in fungus removal was examined to learn how flies find those micro-sized organisms attached on their surface. Since size of conidia does not allow flies to find them visually, we studied a role of chemical cues using the wildtype fly Canton-S, the taste deficiency mutant poxn 70, and the olfactory deficiency mutant orco1. The results showed that flies remove fungal conidia from their body surfaces via grooming behavior, and indicated that it is olfactory cues rather than gustatory cues, which have a significant role in fungal removal in D. melanogaster.

The Drosophila model system for analyzing natural variation in resistance to Metarhizium spp.



Jonathan B. Wang, Hsiao-Ling Lu, Raymond J. St. Leger

Department of Entomology, University of Maryland, Maryland, USA Corresponding author: stleger@umd.edu

We have used broad and narrow host range Metarhizium spp, generalist and narrow host plant specialist Drosophila spp, 188 Drosophila melanogaster Genetic Reference Panel (DGRP) lines, and a panel of mutant D. melanogaster to explore the genetic basis of natural variation in Metarhizium host specialization and insect disease resistance. Features of interest include: 1) substantial individual variation in disease resistance between flies collected in the same habitat, with implications for the evolution of disease resistance; 2) strong genetic control over microenvironmental plasticity, particularly in fly species that have a wide environmental range; 3) males are typically more resistant than females; 4) little difference between sporulation capacity on resistant lines and susceptible lines, so there is no penalty for the fungus to kill quickly, and 5) variation between DGRP lines in resistance to M. anisopliae was correlated with resistance to Pseudomonas aeruginosa, oxidative stress sensitivity, sleep duration and number of nightly 'naps'. We identified a host of candidate genes associated with variation in disease resistance, many of which are known to interact physically and/or genetically which enabled us to place them in a biologically informative genetic network. Overall, our results suggest that fly's differ in their ability to control and/or tolerate replicating fungi during infection, which is achieved mostly through the coordinated interplay of morphological and physiological restraints, and phagocytic effectors that function in subtly different ways in different lines. However, the majority of polymorphisms with major effects on disease resistance were rare, suggesting a general cost to defense involving tradeoffs.

Host responses to entomopathogenic fungi

11:45 •111•

Wei Zhang^{1,2}, Yuxian Xia² and Nemat O. Keyhani^{1,2}

¹University of Florida, Dept. of Microbiology and Cell Science, Bldg 981, Gainesville, FL 32611, USA; $^{\rm 2}$ Chongqing University, School of Life Sciences, Chongqing, 400045, PR China Corresponding author: keyhani@ufl.edu

Insects employ a variety of defensive systems to deter and/or repond to potential microbial pathogens. Entompathogenic fungi have evolved measures to overcome host defenses, resulting in the selection of host countermeasures, ultimately leading to an arms race between the host and the pathogen. Litle, however, is known about specific host tissue responses to fungal infection. Transcriptomic analyses were performed to investigate the global gene expression responses of hemocytes, the fat body, and the central nervous system of the socially flexible locust, Locusta migratoria, during infection by the locust-specific fungal pathogen, Metarhizium acridum.

Marshall Room

Temporal patterns of gene expression were examined throughtout a time course of infection that included (a) spore attachment and consolidation (4 h), (b) germination on the insect cuticle surface (12 h), (c) appressorium, cuticle penetration and initial ingress into the hemocoel (24h/36h) and (d) mycosis (48h/72 h). Specific tissues were dissected during this time interval and RNA_Seq analyses performed. These data revealed that the fat body responds to fungal infection mainly through activation of genes involved in innate immune, energy metabolism and development. Hemocyte gene expression responses mainly affected genes involved in membrane regulation, activation of cellular immune responses, and release of humoral immune factors. Surprisingly, significant changes in central nervous system (CNS) tissues were seen throught the infection time course including at the earliest time point (4 h). Both neurological pathways and a significant number of immune related deifferentially expressed genes (DEGs) were identified in the CNS global gene expression response to fungal infection. The transcriptomics data were verified via qRT_PCR quantification of select genes.

A number of host genes are currently being examined via RNA interference knockdown in order to identify potential targets for increasing the efficacy of

Endophytic association of Metarhizium robertsii under varying C and N soil conditions and plant immune responses

entomopathogenic fungi to target insect hosts.

•112•



¹Larissa Barelli, ¹Sha Sha Hu, ²David Liscombe, ¹**Michael J Bidochka** ¹Dept. Biological Sciences,Brock University, St. Catharines, ON Canada L2S 3A1 Vineland Research and Innovation Centre, Lincoln, ON Canada LOR 2E0

Corresponding author: bidochka@brocku.ca

Many vascular plants are able to form close symbiotic associations with endophytic fungi. Metarhizium is a common soil fungus that is both a plant endophyte and an insect pathogen. Previously we have shown that the endophytic capability and insect pathogenicity of Metarhizium are coupled to provide an active method of nitrogen transfer to a host plant via fungal mycelia. Here we investigated nitrogen transfer from insects infected by Metarhizium robertsii under soil conditions with excess carbon (1% glucose) or nitrogen (1% ammonium nitrate) to host plants (haricot beans). Highest amounts of insect nitrogen were transferred to plants without soil treatment without C or N at 14 days in a soil microcosm containing M. robertsii and insects (Galleria mellonella). Interestingly highest fungal association with host plants was found in soil with excess carbon at 21 days. Also highest fungal amounts were in soil amended with excess carbon at 21 days. Other time points (0, 7, 14 and 28) were not statistically different with respect to fungal colonization. We are also interested in the question as to why plants allow beneficial fungi such as Metarhizium to colonize roots while pathogens elicit a strong immune response. We used the OPLS-DA method to tease out which compounds were driving the differences between plants colonized by Metarhizium as opposed to plants without Metarhizium. A Principal Component Analysis plot was used to illustrate global differences between samples. Overall, thousands of mass spectral features (compounds) were detected but out of all of these, Metarhizium seems to enhance precursor (4coumarate and tartaric acid) pools an accumulation of trans-coumarate, after day 7. We also observed jasmonate action- a hydroxyjasmonate (12-hydroxy or 11-hydroxy) accumulated at higher levels in Metarhizium colonized plants. No salicylic acid or derivatives were detected in these samples. The results help to understand better the endophytic interaction of Metarhizium in agroecosystems and plant immune responses to fungal endophytes.

Diseases of Beneficial Invertebrates Division #2

Moderators: David Bass, Colleen Burge

Health monitoring and disease management of endangered white abalone (Haliotis sorenseni) in a captive breeding program



Blvthe C. Marshman¹. Malina M. Loeher 1,2, Kristin M. Aquilino², Jim D. Moore^{1,2,3}

¹California Department of Fish and Wildlife; ²Bodega Marine Laboratory, University of California, Davis: ³Karen C. Drayer Wildlife Health Center, University of California, Davis Corresponding author: blythe.marshman@wildlife.ca.gov

White abalone (Haliotis sorenseni), a marine snail species native to waters off Southern California, significantly declined in the 1970s as a result of overfishing. In 2001, they were the first marine invertebrate to be federally listed as an endangered species and, since their listing, the population has continued to decline in the wild. The White Abalone Recovery Program, a NOAA-sponsored mission led by the Bodega Marine Laboratory (BML), is dedicated to restore the species throughout its native range through outplanting of captively reared progeny of wild-collected parents. One of the biggest challenges faced by this project is the susceptibility of the species to the rickettsiales-like prokaryote Candidatus Xenohaliotis californiensis (Xc), an intracellular bacterium that can cause a chronic degenerative disease called Withering Syndrome (WS). The WS disease, which can affect both wild and captive abalone, is exacerbated by elevated seawater temperatures and limited food sources. To help combat WS within captive breeding populations involved in the program, the CDFW Shellfish Health Lab performs routine testing on fecal samples using qPCR to detect emerging infections within populations at BML and partner facilities. To treat the disease, infected abalone undergo an oxytetracycline (OTC) antibiotic bath treatment that has been shown to not only eliminate the Xc infection, but also protect treated abalone from subsequent reinfection with Xc for at least four months post-treatment. The 21-day treatment, consisting of eight baths, has the advantage of not only administering a uniform dose to each abalone, but also allows for whole captive populations to be treated simultaneously. Shell-boring organisms, including bivalves, sponges, and polychaetes, exert additional stress on their abalone hosts and may facilitate opportunistic microbes that cause shell lesions. A shell waxing protocol was developed to rid the affected abalone of these pests and it has been effective in reducing the deleterious health impacts, including mortality, caused by lesions. Due to their efficacy while presenting minimal risk, we believe that both the OTC bath and shell waxing treatments can be used on captive abalone populations to aid in restoration efforts.

Candidatus Xenohaliotis californiensis viability and infectious dose in Haliotis rufescens



Nina S. Lottsfeldt, Mariah E. Weavil-Abueg, Lisa M. Crosson, Carolyn S. Friedman

School of Aquatic and Fishery Sciences, University of Washington, Seattle, Washington, USA Corresponding author: carolynf@uw.edu

Withering syndrome (WS) is a fatal disease that affects a number of NE Pacific abalone species (Haliotis spp.). The etiological agent is the Rickettsiales-like bacterium (RLO) Candidatus Xenohaliotis californiensis, an obligate intracellular parasite. Our study aimed to: (1) determine how long the WS-RLO remains viable in seawater under two temperature conditions, 14°C and 18°C, and (2) create a standardized protocol for exposing abalone in the laboratory with known concentrations of WS-RLO using infectious dose 50 (ID50) methodology. To determine WS-RLO viability, duplicate 500mL samples of effluent seawater from a California abalone farm were filtered daily for a total of 8 days. DNA and RNA were extracted from all filters and quantified via qPCR as a proxy for viability. To determine WS-RLO ID50, triplicate tanks of 20 uninfected red abalone were exposed for 3 hours to 1L of WS-RLO homogenate (infected abalone post-esophageal tissue and feces

suspended in sterile seawater) at four concentrations: 0, 10³, 10⁴, and 10⁵ WS-RLO DNA copies/mL. Abalone fecal samples were collected bi-weekly from each tank to monitor WS-RLO DNA loads and abalone post-esophageal tissues were sampled at day 61 post-exposure. Results from our viability experiment indicated WS-RLO DNA loads in seawater were highest at day 2 at both 14°C and 18°C then rapidly declined below the qPCR assay limit of detection by days 5 and 4, respectively. WS-RLO cDNA loads were 30-fold lower than WS-RLO DNA loads indicating that of the total WS-RLO DNA present, only a small proportion was potentially viable. Results from our ID50 experiment concluded that only 2.2-3.2 x 10³ WS-RLO DNA copies/mL are required to generate a 50% infection prevalence. These findings provide critical information to enable a better understanding of WS-RLO transmission dynamics including concerns over WS spillover and spillback between wild and farmed abalone populations, and can aid management decisions for the successful protection and restoration of threatened and endangered abalone species.

Complementary methods for the detection and characterisation of novel haplosporidian parasites of the edible mussel, Mytilus edulis

Georgia M. Ward^{1,2}, Stephen W. Feist¹, Ander Urrutia¹,

STU 11:00 •115•

¹Centre for the Environment, Fisheries and Aquaculture Science (Cefas), Weymouth Laboratory, UK DT4 8UB; ²Department of Life Sciences, Natural History Museum, London, UK, SW75BD

Corresponding author: georgia.ward@nhm.ac.uk

Stuart Ross¹, Matthew Green¹, David Bass^{1,2}

Haplosporida (Rhizaria, Cercozoa) is an order of protistan parasites of aquatic invertebrates, including molluscs, crustaceans and annelid worms. A number of haplosporidian species are known to be responsible for significant mortalities in commercially exploited oysters, including Haplosporidium nelsoni and H. costale (Crassostrea spp.) and Bonamia ostrea (Ostrea spp.). While these pathogens are well characterized within the host, parasite life cycle, modes of transmission and geographic distribution are much less clear. Similarly environmental DNA (eDNA) studies have hinted at significant uncharacterised genetic diversity within the order, however the relationships between these novel haplosporidian lineages and their hosts remains largely unexplored. This study uses the example of Minchinia sp., a novel parasite of the edible mussel Mytilus edulis, to demonstrate the efficacy of combining traditional histopathology with haplosporidian-targeted PCR screening, in situ hybridisation (ISH) and eDNA approaches to explore the prevalence, phylogenetics and life-cycle of haplosporidian parasites.

Development of biomarkers for Ostreid herpesvirus 1 resistance in Pacific oysters

Natalie D. Rivlin¹, Colleen A. Burge¹, Collin J. Closek², Carolyn S. Friedman³

¹Institute of Marine and Environmental Technology, University of Maryland Baltimore County, Baltimore, Maryland, USA; ²Center for Ocean Solutions, Stanford University, Palo Alto, California, USA: 3School of Aquatic and Fishery Sciences, University of Washington, Seattle, Washington, USA Corresponding author: nrivlin@umbc.edu

The Ostreid herpesvirus 1 (OsHV-1) has emerged as an infectious disease agent associated with mass mortalities of larval and seed Pacific oysters, Crassostrea gigas. Although OsHV-1 is a global pathogen of bivalve molluscs, it has been observed in only 2 adjacent bays in the US: Tomales Bay (TB) and Drakes Estero, California. In TB, OsHV-1 has caused severe (50-60%) losses of Pacific oyster seed nearly annually since the early 1990's, with mortality events recurring during the summer months. Seasonal elevated water temperatures are consistently associated with oyster mortalities in TB, and may trigger viral replication and/or transmission of OsHV-1 to naïve juvenile oysters. The present research investigates host susceptibility of Pacific oysters exposed to OsHV-1 and aims to develop biomarkers for OsHV-1 resistance. Individuals from two US west coast Pacific oyster families (hybrid pair-mated families) were exposed to OsHV-1 in TB over the course of a summer, and sampled periodically throughout mortality events. The two families exhibited differential mortality. Viral replication of OsHV-1 was quantified from whole-body tissue samples using qPCR, which revealed

differential copy number between the two families. Samples are currently being assessed using genetic and transcriptomic methods to screen for OsHV-1 resistant genes. Characterizing resistance traits will help to provide tools and solutions to the US and global oyster industries and will be informative to Pacific oyster breeding programs.

Hytrosavirus genetic diversity and eco-regional spread in Glossina species

STU 11:30 •117•

Meki, I.K.^{1, 2}, Kariithi, H.M.^{1, 3}, Vlak, J.M.², van Oers, M.M.^{2, 3} Parker, A. G.1, Vreysen M.J.B.1, Abd-Alla A.M.M.1,*

¹ Insect Pest Control Laboratory, Joint FAO/IAEA Programme of Nuclear Techniques in Food and Agriculture, Vienna, Austria; ² Laboratory of Virology, Wageningen University, The Netherlands. Biotechnology Research Institute, Kenya Agricultural & Livestock Research Organization, Nairobi, Kenya

Corresponding authors: a.m.m.abd-alla@iaea.org; monique.vanoers@wur.nl

Tsetse control via the sterile insect technique requires tsetse mass production, which can be hindered by infections with Glossina pallidipes Salivary gland hypertrophy virus (GpSGHV; Hytrosaviridae) that can cause collapse. Although GpSGHV asymptomatically infects many Glossina species, the rare SGH symptoms are preferentially triggered in *G. pallidipes*. Hypothetically, G. pallidipes significantly drives GpSGHV evolution that can lead to emergence of more virulent virus variants transmissible to multiple Glossina species. Here, we report GpSGHV diversity in seven tsetse species derived from 20 African geographical locations. Species identities were verified using optimized PCR-based technologies including the use of nuclear internal transcribed spacer sequence, microsatellite markers, and diagnosis of Wolbachia infection. Based on differences between GpSGHV-Uganda and GpSGHV-Ethiopia genomes, three genes and two variable number tandem repeat regions were PCR-amplified and sequenced to identify GpSGHV haplotypes and their phylogenetic relatedness. Of the identified 14 GpSGHV haplotypes, 13 were present in G. pallidipes from 13 of the 20 sampled locations. GpSGHV diversity was highest within the G. pallidipes populations; the reference haplotype was the most widely distributed (in 8 out 20 locations). These data are valuable in establishment of new tsetse colonies, and in prediction/ prevention of SGH outbreaks in facilities producing multiple Glossina species.

Impact of Glossina pallidipes salivary gland hypertrophy virus on the performance of heterologous host, G. f. fuscipes



Guler D. Uzel^{1, 2}, Andrew G. Parker¹, Marc J B. Vreysen¹, Robert L. Mach². Adlv M.M. Abd-Alla¹

¹Insect Pest Control Laboratory, Joint FAO/IAEA Division of Nuclear Techniques in Food and Agriculture, International Atomic Energy Agency, Vienna, Austria; ²Institute of Chemical, Environmental and Biological Engineering, Research Area Biochemical Technology, Vienna University of Technology, Vienna, Austria. Corresponding author: A.M.M.Abd-Alla@iaea.org

Tsetse flies (Diptera: Glossinidae) are the vector of African trypanosomosis, the cause of sleeping sickness in humans and nagana in animals. The lack of effective vaccine and the development of resistance by trypanosomes to the available drugs makes vector control the most effective approach for sustainable management of this disease. The sterile insect technique (SIT) is an effective method to suppress or eradicate tsetse flies in the frame of an area-wide integrated pest management programme. SIT relies on the mass production of the target insect and release of sterile males in the targeted area to compete with wild males in mating with wild females. Tsetse mass rearing is a challenge for SIT application and an understanding of the reproductive biology and associated pathogens is essential to improve mass rearing. Glossing fuscipes fuscipes is an important vector of sleeping sickness in central Africa. GpSGHV affects the fecundity, productivity and therefore performance and sustainability of G. pallidipes colonies. In this study, we evaluated the impact of GpSGHV on the performance of a G. f. fuscipes colony including productivity, mortality, survival, flight ability and insemination rates. The results indicate that GpSGHV infection has a significant impact on mortality, productivity, survival and mating ability but not on adult flight ability or insemination rates in G. f. fuscipes. These results explore the important role of GpSGHV in the sustainability of G. f. fuscipes

colonies and the need to implement measures to avoid virus infection to ensure the successful establishment of mass rearing of this species for SIT programmes.

Harmless sea snail parasite causes mass mortalities in numerous commercial scallop populations in the northern hemisphere

12:00

Árni Kristmundsson¹, Mark Andrew Freeman²

¹ Institute for Experimental Pathology at Keldur, University of Iceland, Keldnavegur 3, IS-112 Reykjavík,
Iceland, ² Ross University School of Veterinary Medicine, Basseterre, St. Kitts, West Indies

Corresponding author: arnik@hi.is

Phylum Apicomplexa comprises a group of unicellular, often highly pathogenic, obligate parasites exploiting either one host (monoxenous) or two hosts (heteroxenous) to complete a full reproductive cycle. Life cycles of species infecting molluscs are poorly understood, although both monoxenous and heteroxenous species are known. For decades, cyclical mass mortality events have occurred in various scallop populations, some of which have been recently attributed to apicomplexan infections. An apicomplexan, causing severe myodegeneration, recently reduced the Iceland scallop population by 90%, caused mass mortalities in queen scallop in Faroese waters and is the probable aetiology behind mass mortality events of sea scallops in the eastern USA and the Gulf of Alaska. We report the first dual mollusc life cycle for an apicomplexan: a species highly pathogenic in pectinid bivalves, but apathogenic when infecting the common whelk as Merocystis kathae. The sympatric distribution of the common whelk and the scallop in the North Atlantic makes transmission extremely effective, occurring via the gastrointestinal tract, by scavenging and predation in whelks and unselective filter feeding in scallops. Infective sporozoites from whelks utilize host haemocytes to reach muscular tissue, where asexual reproduction occurs. Phylogenetically, this apicomplexan is robustly placed within the Aggregatidae and its inclusion in analyses considerably increases resolution between other basal apicomplexans groups (gregarines), which raises concerns over the validity of some related taxa. Scallops seem able to regulate low-level infections of *Merocystis* as they exist in normal scallop populations between epizootics but disease outbreaks occur during prolonged or high levels of exposure from localised infected whelks. Hence, a targeted removal of whelks from valuable scallop grounds would be advantageous to minimize the occurrence of M. kathae epidemics and prevent damaging economic losses.

TUESDAY, 12:30 – 5:00 pm EXCURSIONS

Boxed lunches are provided for excursion participants. Buses for the San Diego Zoo and Midway excursions depart at 12:30 pm from the Price Center, and will drop off participants at the Birch Aquarium at 5:30 pm.

TUESDAY, 6:00 – 10:00 pm BARBEQUE at Birch Aquarium

Buses will leave the Village housing area at 5:30 pm for the Birch Aquarium for those not attending an excursion. Buses will return from the Aquarium as needed between 9 and 10 pm.

WEDNESDAY, 8:00 - 10:00 am

Theater

Bacteria Division #3

Moderators: Juan Ferré, Satomi Adegawa

Functional characterization of Vip3Ab1 and

8:00



Vip3Bc1 N-terminal and C-terminal domains

Marc Zack, Megan Sopko, Meghan Frey, Xiujuan Wang, Jennifer Arruda, Ted Letherer, Sek Yee Tan, and Ken Narva

Dow AgroSciences, 9330 Zionsville Road, Indianapolis, IN 46268

Corresonding author: mdzack@dow.com

Vegetative insecticidal proteins (VIPs) are soluble insecticidal proteins produced by Bacillus cereus and B. thuringiensis with demonstrated activity on lepidopteran insects. In general, it is accepted that Vip3 function by a mode of action different than that of the more-studied 3-domain crystal Bt family of insecticidal proteins. Therefore, the Vip3 family is valuable for development of insect-resistant crops such as corn and cotton. To date, 3 subsets of the VIP3 family have been identified; VIP3A, VIP3B, and VIP3C. We have recently shown that Vip3Bc1, a new member of the Vip3B family, has an insecticidal spectrum unique from that of Vip3A proteins. In addition, we have shown that Vip3Bc1, like Vip3Ab1, forms a ~340 kDa tetramer that is present before and after proteolysis. In the present work, we extend our understanding of Vip3 toxins by using chimeras of Vip3Ab1 and Vip3Bc1 to explore the relative roles of amino and carboxy terminal protein domains. We produced proteins in which we exchanged the N-terminal ~200 amino acids, named Vip3_AB and Vip3_BA, to denote the N-terminal changes. We then analyzed proteolytic stability, tetramer formation, and insecticidal activity. Vip3 AB was processed as expected by gut enzymes and formed tetramers before and after digestion. However, Vip3_BA was present primarily in monomeric form with some presence of tetramer. Monomers, but not tetramers, were rapidly proteolyzed by gut enzymes. Both chimeras lost lethal insecticidal activity. However, Vip3_BA caused significant growth inhibition of insects susceptible to Vip3Ab1 despite the majority of protein being present as a protease-sensitive monomer. Therefore, this work indicates a specific interaction between N-terminal and C-terminal domains of Vip3 proteins that is required for proteolytic stability and insecticidal activity. In addition, we have shown that the C-terminal portion of these proteins, often referred to as a core, directs specificity but is rapidly degraded and loses all activity on susceptible insects. These data provide novel insights towards Vip3 mechanism of action and insecticidal specificity that will support the development of new insecticidal traits.

Stability of the *Bacillus thuringiensis*Vip3Aa protein to protease digestion

8:15



Yolanda Bel, Núria Banyuls, Maissa Chakroun, Baltasar Escriche, Juan Ferré

ERi de Biotecnología y Biomedicina (BIOTECMED), Departament de Genètica, Universitat de València, 46100 Burjassot, España. Corresponding author: juan.ferre@uv.es

Vip3 proteins are secretable proteins from *Bacillus thuringiensis* whose mode of action follows similar steps as the Cry proteins. Once in the midgut of the insect, Vip3 proteins are activated. The activation of Vip3Aa has been shown to be necessary to form pores *in vitro*, and differences in the rate of activation have been related with differences in susceptibility amongst lepidopteran species. It had been described that the *in vitro* activation of Vip3A proteins with trypsin or insect midgut juice produces a major fragment of about 62–65 kDa, along with other fragments, mainly one of about 20 kDa that corresponds to the N-terminal region. In the present study, the activation process for the Vip3Aa16 protein has been closely examined in order to better understand the Vip3Aa protein stability and to shed light on its structure. At high concentrations of trypsin or midgut juice, if the action of proteases is not properly neutralized, the results of SDS-PAGE analysis may lead to think that the protoxin is completely processed. However, when the proteolytic reaction is efficiently stopped, the result reveals that the protoxin

is only cleaved at a primary cleavage site, regardless of the amount of trypsin used. The 66 kDa and the 19 kDa peptides generated by the proteases coeluted after gel filtration chromatography, indicating that they remain together after cleavage. The 66 kDa fragment was found to be extremely resistant to proteases. The trypsin treatment of the protoxin in the presence of SDS revealed the presence of secondary cleavage sites at S-509, and presumably at T-466 and V-372, rendering C-terminal fragments of approximately 29, 32, and 42 kDa, respectively. The fact that the predicted secondary structure of the Vip3Aa protein shows a cluster of beta sheets in the C-terminal region of the protein might be the reason behind the higher stability to proteases compared to the rest of the protein, which is mainly composed of alpha helices.

8:30 •121• Vip3Ca induces apoptosis in midgut epithelial cells from Spodoptera exigua and in cultured Spodoptera frugiperda (Sf21) cells

> Patricia Hernández-Martínez¹, Joaquín Gomis-Cebolla¹, Juan Ferré¹, **Baltasar Escriche**¹

¹ERi de Biotecnología y Biomedicina (BIOTECMED), Departamento de Genética, Universitat de València, 46100 Burjassot, España. Corresponding author: Baltasar.escriche@uv.es

Bacillus thuringiensis produces a wide range of insecticidal proteins which are active against a number of agriculturally pest species. Nowadays, products based on either crystalline insecticidal proteins (Cry proteins) and/or vegetative insecticidal proteins (Vip proteins) are used to control insect pests. Whereas the mode of action of Cry proteins has been extensively studied, little is known about the mechanisms of action of Vip3 proteins. In the present work, the mechanism responsible for Vip3Ca toxicity in both Spodoptera exigua larvae and in Spodoptera frugiperda cultured insect cells (Sf21) was studied. To elucidate the mechanism of cell death in the midgut epithelial cells, S. exigua larvae were exposed to different sublethal concentrations of Vip3Ca. Then, midguts were sectioned and stained using the DeadEndTM Fluorimentric TUNEL (terminal deoxynucleotidyl transferase dUTP nick end labeling) system. The results showed that in control larvae, and in the larvae exposed at the two lower concentrations used (10 and 100 ng/cm²), no TUNEL-positive cells were observed. However, TUNEL-positive cells were observed in the gut of the larvae intoxicated at the highest concentration used (10000 ng/cm²). These results suggest that the epithelial cell response is dose-dependent. In parallel, using the same methodology, we showed that the Vip3Ca protein can also induce apoptosis in Sf21 cells, suggesting that this in vitro system can be useful to further characterize the biochemical basis of the cell death.

Novel synergistic activity between Bacillus thuringiensis STU 8:45 •122• Cry9Aa and Vip3Aa toxins against Chilo suppressalis (Walker)

> **Zeyu Wang**^{1,2}, Longfa Fang¹, Zishan Zhou¹, Sabino Pacheco², Mario Soberón², Alejandra Bravo², Jie Zhang¹

¹State Key Laboratory for Biology of Plant Diseases and Insect Pests. Institute of Plant Protection, Chinese Academy of Agricultural Sciences, No. 2 Yuanmingvuan West Road, Haidian District, Beijing, 100193, China: ²Instituto de Biotecnología, Universidad Nacional Autónoma de México, Cuernavaca, Apdo. Postal 510-3, Morelos, 62250, Mexico Corresponding authors: jzhang@ippcaas.cn; bravo@ibt.unam.mx

A novel synergistic interaction among two different proteins produced by Bacillus thuringiensis (Bt) is described in this work. The Cry9Aa, a threedomain Cry toxin, and Vip3Aa, a vegetative insecticidal toxin, exhibit highly significant synergism against Chilo suppressalis with synergy factor of 35.7. These two proteins bound specifically to BBMV of C. suppressalis, and do not share binding sites since there is no-binding competition between them. We demonstrate that both Cry9Aa and Vip3Aa interact with each other by Ligand-blot, ELISA and Pull-down assays. This interaction showed an affinity value (Kd) of 40.1 nM. The specific interaction sites between these proteins were identified by analyzing binding of Cry9Aa to overlapping fragments of Vip3Aa and by analyzing binding of Vip3Aa to individual domains of Cry9Aa heterologous expressed in E. coli cells. To further narrow the binding sites of Cry9Aa and Vip3Aa we are using cellulose membrane-bound peptides. Ongoing research is focused to functional studies involving site-directmutagenesis followed by synergistic bioassay. The results indicate, for the

first time, that the binding between Cry9Aa and Vip3Aa could be the basis of their synergism against C. suppressalis. Our data contribute to further understanding of synergy activity among Bt toxins and will be highly relevant to propose novel combinations of toxins to be used in the Pyramids scheme that would help to effectively control this pest and delay development of resistance.

A genomic and proteomic approach in the mining of new insecticidal proteins from Bacillus thuringiensis

STU 9:00 •123•



Gomis-Cebolla, J¹, Ricietto, APS¹, Ferré, J¹.

¹ERI of Biotechnology and Biomedicine (BIOTECMED), Department of Genetics, Universitat de València, Burjassot, Spain; ² Departamento de Biologia Geral, Universidade Estadual de Londrina, Londrina, Paraná, Brazil Corresponding author: juan.ferre@uv.es

The next generation sequencing (NGS) allows rapid sequencing of whole genomes with profitable cost-effective ratios and in combination with the development of a large number of freeware NGS tools can enable the rapid detection of insecticidal protein genes at the genome level. Nine samples were selected from Spanish collections of Bacillus thuringiensis (Bt) based in their vip1 and vip2 gene content (Hernández-Rodríguez et al.,2009 J. Appl. Microbiol. 107:219-225). As a first step, we determined which type of vip1 and vip2 genes contain every Bt isolate and we selected those Bt isolates with low similarity to the described vip1 and vip2 genes (E-SE10.2 and O-V84.2). With these Bt isolates we performed a combined genomic and proteomic approach. These two Bt isolates were sequenced to determine the full gene content and then a proteomic analysis of the secreted and crystal insecticidal proteins was conducted to validate the automatic genomic annotation of Rast Server. As a result, we obtained a set of vegetative proteins, specifically two couples of Vip-like proteins (Vip2Ac-like_2/Vip4-like and Vp2Ac-like_3/Vip4Aa1-like_1). If insect bioassays prove that they hold insecticidal activity, these proteins could be considered as first representatives of new binary toxins, Vip6Aa-Vip5Aa and Vip6Ba-Vip5Ba, respectively.

Implication of the Bacillus cereus & B. thuringiensis siderophore Bacillibactin and its Siderophore Bonding Protein (SBP) FeuA in growth and virulence under iron restricted conditions

Laurent Consentino Christophe Buisson, Agnès Rejasse, Christina Nielsen-Leroux

INRA, UMR 1319- Micalis-AgroParisTech, Domaine de Vilvert, Jouv en Josas, France Corresponding author: Christina.nielsen-leroux@inra.fr

The ability of B. cereus (Bc) and B. thuringiensis to colonize various mammals and insects is linked to the presence of several adaptation factors, one of which is the capacity to acquire iron. Iron is an essential for several cellular functions and thus for all living organisms. Meanwhile free iron is toxic and is bound to molecules like hemoglobin, ferritin, transferrin etc. Previously, an in vivo screen of B. cereus led to the identification of a novel protein, IlsA, which is specifically expressed in the insect hemocoel and under iron restrictive conditions in vitro. It was further shown that IIsA is localized on the surface of B. cereus and affinity tests revealed that IIsA interacts with both hemoglobin and host ferritin. Furthermore, we showed that B. cereus siderophores (bacillibactin, encoded by the operon entA-dhbBCF) is particularly important for ferritin iron uptake and for full virulence in G. mellonella and may favor IlsA-related iron uptake (Segond et al. 2014). Siderophores are small molecules with strong affinity for ferric iron. To enter the bacteria the siderophore/Iron complex generally binds to a Siderophore Binding Proteins (SBP) at the surface of the cell. To get further insight into the role of the SPB FeuA of B. cereus in iron uptake from ferritin a Bc ATCC 14579 strain mutant ΔfeuA and a ΔfeuAΔentA double mutant along with complemented strains were constructed and analysed in vitro and in vivo. The results indicate that the ΔfeuA mutant is affected in vitro almost similarly to the bacillibactin entA-mutant with horse-spleen ferritin as the sole iron source. Addition of extracellular bacillibactin restored the growth kinetics of only the DentA mutant. Interestingly, in vivo infection (injection of exponentially growth bacteria into the hemocoel) of G. mellonella shows that the feuA mutant and particularly the double mutant ΔfeuAΔentA were as virulent as the willdtype strain. These results may indicate that in vivo the

double mutant could be stimulated to express other virulence /adaptation factors or may be less sensitive to insect defense mechanisms. These hypotheses are currently under investigation.

A revolution in global health: mass-producible purified natural Bt crystals as a drug product



Yan Hu, Ambily Abraham, Jason Noon, Zeynep Mirza, Hanchen Li Florentina Rus David Gazzola Devsy Tatianan Pinto Rodriguez, Gary Ostroff, Raffi V. Aroian

> University of Massachusetts Medical School. Program in Molecular Medicine, Worcester, MA, USA; Corresponding author: raffi.aroian@umassmed.edu

Soil-transmitted helminths (hookworms, whipworms, Ascaris) are nematodes that live in the human intestine and are the most common parasites on earth. They are major contributors to childhood stunting and poverty worldwide. Low efficacy and emerging resistance to current chemical drugs mandates development of new therapeutics. We have demonstrated that spore-crystal lysates expressing the three-domain Bt crystal protein Cry5B can cure hookworm and Ascaris infections in animal models of human disease and significantly impact whipworms. This makes Bt and Cry5B potential major players on the global health stage. The primary challenge for the development of a Bt-based therapeutic is the identification of an appropriate API (Active Pharmaceutical Ingredient) containing Cry5B. Cry5B spore-crystal lysates, although mass producible, face a difficult road as an FDA approvable drug given the phylogenetic proximity of Bt to Bacillus cereus and the difficulty in defining drug substance specifications for a crude mixture of live spores, bacterial lysate contaminants and Cry5B as the API. In the course of defining what a suitable Bt Cry protein drug substance might look like, we ascertained that an ideal solution might be purified natural Bt crystals that lack spores and Bt contaminants. However, heretofore we knew of no such means to purify Bt crystals free of spores and contaminants by a pharmaceutically scalable, inexpensive process that retains full bio-activity. Such purified crystals could be formulated for oral delivery in roomtemperature stable enteric capsules to treat STH infections. Here, we describe the discovery and development of a method to purify naturally formed protein crystals from Bt or other heterologous microbial protein production systems that contain one or more Cry proteins free from spores and other bacterial contaminants (>95% pure at the protein level). The simple process is inexpensive and compatible with industrial and pharmaceutical production methods. Tests in vitro and in vivo against parasitic nematodes with Purified Cry5B Crystals (PCC) demonstrate that PCC is highly bioactive, even more so than Cry5B spore-crystal lysates. The discovery of PCC not only opens up the path for development of Cry protein drugs, but also potentially novel and potent applications of purified Bt crystal insecticides.

Novel insecticidal proteins from plants for crop protection against major lepidopteran pests



Jennifer Barry¹, Lu Liu², Eric Schepers¹, Amy Lum², Janet Rice¹, Nasser Yalpani¹, Ryan Gerber³, Nuria Jimenez¹, Fikru Haile¹, Xiuli Qi¹, Adane Kassa¹, Matt Heckert², Weiping Xie², Scott H. Diehn¹, Virginia C. Crane¹, Howard Damude⁴, Carol Pilcher¹, Russ Booth¹, Mark Nelson¹, Albert L. Lu¹, Timothy M. Nowatzki¹, Gusui Wu¹

¹DuPont Pioneer, Johnston IA, USA; ²DuPont Pioneer, Hayward CA, USA; ³AgBiome Inc., Research Triangle Park NC, USA; 3 Jackson ImmunoResearch Inc., West Grove PA, USA Corresponding author: jennifer.barry@pioneer.com

Many lepidopteran insect species are economically important pests of crops worldwide. The loss can be in billions of US dollars annually due to reduced crop productivity and increased pest management costs. Transgenic crops developed with Bacillus thuringiensis (Bt) insecticidal protein encoding genes have been widely adopted and they offer benefits to both farmers and the environment by increasing productivity and reducing reliance on conventional pesticides. However, rapid and widespread adoption of these transgenic crops has increased the risk of evolution of pest resistance to Bt proteins and in certain instances it has resulted in reduced effectiveness. We report here the discovery of a new family of potent insecticidal proteins from plant species. These proteins show activity against lepidopteran insect pests

including corn earworm (CEW) (Helicoverpa zea), fall armyworm (FAW) (Spodoptera frugiperda), soybean looper (SBL) (Chrysodeixis includens) and velvetbean caterpillar (VBC) (Anticarsia gemmatalis). Transgenic soybean (Glycine max L. Merill) and corn (Zea mays L.) plants expressing those proteins show protection from feeding damage by these insect pests. This family of novel proteins is useful for developing new transgenic crops for lepidopteran pest control.

The Forum

MICROSPORIDIA DIVISION SYMPOSIUM

The past and future frontiers in microsporidiology (Retrospective look at Microsporidia research from the first meeting 50 years ago)

Organizer: Yuliya (Julia) Sokolova

Moderators: Leellen Solter and Susan Bjornson

Highlights of research on microsporidia from aquatic hosts in North America



James J. Becnel

Center for Medical, Agricultural and Veterinary Entomology, US Department of Agriculture, Agricultural Research Service, Gainesville, Florida 32608 Corresponding author: james.becnel@ars.usda.gov

The earliest record of microsporidia from North America was probably Glugea stepani from a fish in 1901. There were few additional reports of microsporidia until Kudo, a pioneer in microsporidian research in North America, published more than a dozen significant papers on mostly aquatic microsporidia from 1920-1925. Kudo (1924) listed 21 species of microsporidia that had been described from North America with the great majority (18) from aquatic hosts. The aquatic hosts were mostly mosquitoes and blackflies but also included parasites of mayflies and microcrustacea. Over the next 30 years progress was slow with only about 13 new species described from insects in North America with only 4 of these from aquatic hosts. During this time, Sprague, a student of Kudo, did publish a number of descriptions of microsporidia from marine and freshwater Crustacea. As part of the movement beginning in the late 1950's to investigate biological control options for pest arthropods, microbial control efforts led to many new reports of microsporidia in host from North America. For the aquatic host, the focus was mainly on mosquitoes beginning with the California Department of Public Health's Fresno laboratory. This was followed by U. S. Department of Agriculture, Agricultural Research Service Laboratories formed in Lake Charles, Louisiana and Gainesville, Florida and eventually at many University programs. The Society for Invertebrate Pathology became a focal point for researchers on microsporidia and this association was formalized with the establishment of the Microsporidia Division in 1969. Much was learned about microsporidian biology and taxonomy in aquatic hosts in North America led by researchers such as Kellen, Chapman, Anthony, Hazard, Sprague, Undeen and a number of students that have carried on the research. Highlights will be presented on the many advancements made on microsporidia found in aquatic hosts by this relatively small but dedicated group of microsporidiologists.

Microsporidia in marine invertebrates

8:20

Kelly Bateman, Grant Stentiford

European Union Reference Laboratory (EURL) for Crustacean Disease, Cefas, Barrack Road, The Nothe, Weymouth, Dorset, DT4 8UB, UK. Corresponding author: Kellv.bateman@cefas.co.uk

Microsporidia are a diverse parasite phylum infecting multiple host groups and causing important diseases in aquatic organisms. Almost half of the known microsporidian genera infect aquatic hosts with approximately 50 genera in arthropods and at least 21 genera found in aquatic non-arthropod invertebrates, protists and hyperparasites of aquatic hosts. To date, most of the focus has been on hosts of commercial or ecological significance (e.g., farmed or fished decapod crustaceans) meaning that Microsporidia remain vastly under-reported in our less enigmatic hosts. Nevertheless, access to materials collected from hosts from a wide range of aquatic environments

and the development of pathological and molecular diagnostics is continually expanding our understanding of the phylum and, providing exposing an everincreasing diversity in pathogen morphology, lifecycle and parasitic strategy. Whilst the majority of taxa were erected on the basis of morphological descriptions of various life cycle stages (meronts, sporonts, sporoblasts and spores), more recent taxonomic assemblages have been based upon phylogenetic data, normally relating to sequencing of the partial SSU rRNA gene. SSU-based phylogenies appear to perform well for placement of new genera (of which there are apparently many) within the phylum. However, where species are more closely related, concatenated phylogenies, based upon multiple gene sequences is proving useful for taxonomic clarity. The increasing availability of whole genome sequences for members of the Microsporidia is assisting construction of concatenated phylogenies based upon provision of gene sequences from across the parasite genome. Overall, whilst our understanding of diversity in the aquatic microsporidians has steadily expanded with the application of molecular diagnostic tools, we predict an era of accelerated discovery based upon use of these tools on a wider range of host and environmental matrices. However, recent work combing morphological characteristics with phylogentic data has demonstrated plasticity in morphological traits making them unreliable metrics for the purpose of classification.

The (relatively) early history of microsporidian researchers in North America

Joseph Maddox

Illinois Natural History Survey, Prairie Research Institute, University of Illinois, Champaign, IL, USA (retired) Corresponding author: jmaddox2405@comcast.net

An informal review of the activities of microsporidian researchers with an emphasis on events before 2000. Questions (and some answers) about the phylogeny and classification of microsporidia, the role of microsporidia as biological control agents, and the formation of the Microsporidia Division of the SIP will be discussed.

The path to registration of a microbial pesticide



John E. Henry

Retired, Corresponding author: jehenrydlake@gmail.com

This narrative describes the initiation and execution of a project on biological control of grasshoppers with an intent of providing non-chemical approaches to managing the densities of grasshoppers. The project led to the selection and development of the pathogenic protozoan, Nosema locustae, as a tool for applied use. It presents the criteria used to select this organism and reviews the extensive testing that established that applications of the organism affected both short term and long term host population regulation. With the involvement of the US Environmental Protection Agency (EPA) the organism was registered as the active ingredient in a microbial pesticide.

History and highlights of microsporidia research in Russia

9:20 •131•

Yuliya Y. Sokolova^{1,2}

¹Louisiana State University, School of Veterinary Medicine, Baton Rouge, LA, USA; ²Institute of Cytology, Russian Academy of Sciences, St.Petersburg, Russia Corresponding author: yysokolova@gmail.com

The focused research on Microsporidia (M) in Russia counts from mid-60ies, when Irma Issi defended her PhD dissertation The microsporidiosis of Cabbage White and other insects, and its biological importance, and established a Microsporidia research group, Issi's lab, within the Laboratory for Microbiological Control, Institute for Plant Protection, St. Petersburg, Russia. This lab has been running by her for more than 50 years now, and became an incubator for young microsporidiologists: >30 PhD students from all over the former Soviet Union have passed through the lab and graduated under Prof. Issi's supervision. The research have been focused on: (1) host population control by M and possible application of M in microbiological control; (2) interactions of M with the insect host: impact on immunity and physiology; (3) host-parasite relations at the cellular and subcellular levels; (4) taxonomy of M. One of the brightest research achievements of the lab

was a long-term (over 25 years) observation on the epizooty of Nosema mesnili in Pieris brassicae that demonstrated how the microspridium took the control over the host population. Peaks and declines in host and parasite densities were accompanied by alterations in the microsporidium host cycle and switches of transmission routes. Unfortunately this research was published only in Russian. Dr.Issi's monograph Microsporidia as a Phylum of Parasitic Protists (1986) focused on the taxonomy, systematics and origin of M, was translated to English by Dr.Lipa and printed in 1991 by the request of SIP Microsporidia Division. Irma's studies were enthusiastically continued by her graduates, Vladimir Voronin, Petr Kilotchitsky (both specialized in microsporidia from freshwater hosts), Tamara Pankova (M from East-Siberian host), Viatcheslav Dolgikh (gave a new breath to the lab by establishing advanced biochemical, cell and molecular biology approaches), and myself (life cycles, taxonomy and cell biology of M), to mention only a few. The next generation of Dr.Issi's prodigious grandkids, Yuriy Tokarev, Elena Nassonova and Anastasiya Simakova, have already established themselves as international scholars. I will also focus on the three major recent highlights of Russian microsporidiology: structure and functional of secretory compartment of M, biochemistry of host-parasite relations, and studies on metchnikovellids.

Microsporidiosis: Perspective from human infections

9:40 •132•

Louis M. Weiss^{1,2}

¹Department of Pathology, Division of Parasitology, Albert Einstein College of Medicine, Bronx, New York, USA: 2Department of Medicine. Division of Infectious Diseases, Albert Einstein College of Medicine, Bronx, New York, USA

Corresponding author: louis.weiss@einstein.yu.edu

Microsporidiosis, infection by members of the phylum Microsporidia, affects both invertebrate and vertebrate hosts. Phylogenetic studies indicate that these pathogens are related to the Fungi, most likely as a sister group with the Cryptomycota. Given the wide host range of these organisms it is not surprising that they are pathogens of humans. These organisms were initially reposted in humans in 1959 with the description of a child with encephalitis, but until the HIV epidemic human infections were only sporadically reported. Gastrointestinal infections due to Enterocytozoon bieneusi and Encephalitozoon intestinalis were associated with chronic diarrhea and wasting in patients with AIDS. The number of reported cases increased exponentially until the widespread use of combination antiretroviral therapy (cART) resulted in a decline in infection in this population. There are over 200 microsporidian genera, of which the following have been reported to cause infection in humans: Nosema (N. corneum renamed Vittaforma corneae; N. algerae reclassified as Brachiola algerae and then Anncaliia algerae), Pleistophora, Encephalitozoon, Enterocytozoon, Septata (reclassified as Encephalitozoon), Trachipleistophora, Brachiola (reclassified as Anncaliia), Anncaliia, Tubuli-nosema, Endoreticulatus and Microsporidium, Infection in humans has now been documented in both immune compromised patients (including those with transplantations) and immune competent patients. These infections have involved almost all organ systems causing diseases such as encephalitis, myositis, sinusitis, keratitis, and conjunctivitis. As our understanding of these ubiquitous pathogens increases our recognition of their role as human pathogens and our ability to diagnose these infections is

THIS SYMPOSIUM CONTINUES IN THIS ROOM AFTER THE COFFEE BREAK

Microsporidia associated with bumble bees 10:30 •133• in the southern Neotropical region



Santiago Plischuk, Marina Haramboure, Carlos E. Lange

Centro de Estudios Parasitológicos y de Vectores (CCT La Plata CONICET-UNLP-CICPBA), La Plata, Argentina Corresponding author: santiago@cepave.edu.ar

With 40 years of history in Argentina and long-term studies restricted to a few host insect orders, Microsporidiology as a whole is still a young discipline in the country. Initial research was prompted by experimental use of the grasshopper biocontrol agent Paranosema locustae. Other sustained efforts

studied microsporidia associated with black flies, mosquitoes, fire ants, and more recently with honey bees and bumble bees. Bumble bees (Bombus spp.) play a key role as highly efficient pollinators in a variety of natural and man-made ecosystems. Of the approximately 250 species known worldwide only nine native species have been reported to inhabit Argentina, Chile, and Uruguay which seem to depict a somewhat low diversity for southern South America when compared to the Palearctic and Nearctic regions. Two additional Palearctic species were introduced late in the XX century in Chile, became naturalized, and spread to Argentina across the southern Andes. Less than a decade ago knowledge of microsporidia associated with bumble bees in the southern Neotropic was nonexistent. Since then surveys started and the occurrence of at least three microsporidia was recorded. The predominantly fat tissue pathogen Tubulinosema pampeana was discovered in native Bombus atratus and B. bellicosus in the Pampas, the enteric Nosema ceranae was registered in natives B. atratus, B. bellicosus, B. morio and B. brasiliensis in the Pampas and North of the country, and systemic Nosema bombi in exotics B. terrestris and B. ruderatus in western Patagonia. Since B. terrestris is extremely ubiquitous and has already reached the northern Patagonian Atlantic coast it is likely that its range expansion will continue further possibly allowing new microsporidia-host associations of unpredictable outcomes. The presentation will review the status of both microsporidia and hosts, emphasizing the importance of immediate intensification of surveys before B. terrestris reaches a wider distribution.

Microsporidia infection in C. elegans: How an obligate intracellular parasite makes itself at home 10:50 •134•



Emily Troemel

University of California, San Diego La Jolla, CA, USA Corresponding author: etroemel@ucsd.edu

Our lab studies Nematocida parisii, which is a natural pathogen of the genetically tractable nematode C. elegans. We have found that N. parisii restructures host cells through causing the host intestine to become a syncytium, and by hijacking host exocytosis for spore exit. These and other strategies have enabled this obligate intracellular parasite to 'make itself at home' in the host organism.

Phylogeny of Microsporidia: impact of molecular approaches 11:10 •135•



Charles R. Vossbrinck¹, Yuri Tokarev²

¹Department of Environmental Sciences, The Connecticut Agricultural Experiment Station New Haven, CT, USA; ²Institute for Plant Protection, St.Petersburg, Russia Corresponding author: Charles.Vossbrinck@ct.gov

Prior to the use of molecular characters to study phylogeny taxa were described based on phenotypic character states. As single-celled microorganisms, defining homology between characters is difficult and it appears that the Microsporidia may be gaining and loosing character states rapidly resulting in polyphyletic taxa. In addition there is no fossil data for microsporidia to clarify any relationships. We will briefly review former classification systems for the microsporidia. We suggest developing a microsporidial classification based upon a molecular phylogeny and placing the character states on the tree in a cladistic fashion.

Roosevelt Room

Microbial Control Division #2

Moderators: Wendy Gelernter, Travis Glare

Toxin-mediated control of insect host population by the endosymbiotic bacteria Spiroplasma poulsonii





G. Garcia-Arraez, F. Masson, J.C. Paredes, B. Lemaitre

Global Health Institute, School of Life Science, Ecole Polytechnique Federale de Lausanne (EPFL), Lausanne, Switzerland. $Corresponding\ authors:\ mario.garcia arraez @epfl.ch;\ bruno.lema itre @epfl.ch$

Spiroplasma is a group of widespread arthropod-associated bacteria. They exhibit a wide range of interactions with their hosts, from parasitism to mutualism. All species lack cell wall components, which make them invisible to the immune system of their insect hosts. Spiroplasma poulsonii (S. poulsonii) is an endosymbiotic bacterium that infects specifically the fruit fly, Drosophila melanogaster. S. poulsonii is found extracellularly in the hemolymph but can colonize the germline during oogenesis to be vertically transmitted from mother to offspring. S. poulsonii can disrupt the sex ratio of its host by inducing male killing, which consists in the specific death of all male embryos. While male-killing has been reported in many species, the molecular mechanisms underlying this phenotype are still unknown. We have isolated a new S. poulsonii variant called blind-killer that kills only 25% of the offspring of young female flies increasing to 70% for the offspring of two-weeks old females. Interestingly, the sex ratio of the offspring is not affected by the infection, since both male and female embryos die in similar proportions. In order to gain insight into the blind-killing phenotype, we have sequenced and compared the genome and transcriptome of both male-killer and blind-killer strains. Intriguingly, a family of toxin encoding genes, called Ribosome Inactivating Proteins (RIPs), were found to be differently expressed between the two strains. To date, Spiroplasma RIPs have been involved in the protection exerted by this endosymbionts against entomopathogenic nematodes in *Drosophila neotestacea*. It was proposed that these RIP toxins can specifically cleave the ribosomal 28S RNA of the nematodes. We are now investigating the role RIPs in male-killing.

DNA-based genotyping for Cry1Fa resistance in field populations of Spodoptera frugiperda



STU 8:15 •137 •



Rahul Banerjee¹, Lucas Hietala², Robert Meagher³, Rod Nagoshi³, James Hasler⁴, Kenneth Narva⁴, and Juan Luis Jurat-Fuentes^{1,2}

¹Genome Science and Technology Program, University of Tennessee, Knoxville, TN 37996, USA; ²Department of Entomology and Plant Pathology, University of Tennessee, Knoxville, TN 37996, USA; ³Behavior and Biocontrol Unit, USDA-ARS, Gainesville, FL 32608, USA: 4Dow AgroSciences, Indianapolis, IN 46268, USA Corresponding author: rbanerje@vols.utk.edu

Populations of the fall armyworm (Spodoptera frugiperda) in North and South America have developed field-evolved resistance to transgenic corn producing the Cry1Fa insecticidal protein from Bacillus thuringiensis. Our group identified mutations in the SfABCC2 gene that are genetically linked with Cry1Fa resistance in Puerto Rico and Florida and developed genotyping assays to detect these alleles in fall armyworm samples. We report the use of these assays to screen for the Puerto Rico and Florida alleles in field collected samples of fall armyworm, including archived and recent collections. Our results allow the detection of resistance evolution in Puerto Rico, testing its spread to southeastern US, and the detection of additional field-evolved resistance alleles. These findings represent the first report of successful molecular monitoring for field-evolved resistance to transgenic Bt crops and facilitate our understanding of resistance dispersal and frequency in field population of S. frugiperda

Pathogen growth rate is constrained by host diet





Robert Holdbrook¹, Catherine E. Reavey¹, Joanna L. Randall¹, Yamini Tummala¹, Sheena C. Cotter², Stephen J. Simpson³, Kenneth Wilson¹

¹ Lancaster Environment Centre, Lancaster University, Lancaster LA1 4YQ, UK; ² School of Life Sciences, University of Lincoln, Brayford Pool, Lincoln LN6 7TS, UK; ³ Charles Perkins Centre, The University of Sydney, NSW 2006, Australia Corresponding author: rhold91@gmail.com

The use of bacteria and other pathogens as biological pesticides against insect crop pests is growing due to environmental concerns over traditional chemical pesticides. However, we have only a rudimentary understanding of how factors such as host nutrition impact on the efficacy of biopesticides. The Egyptian cotton leafworm, Spodoptera littoralis, is a major pest throughout much of Africa. Several studies have shown that the outcome of host-pathogen interactions in this and related species varies with dietary macronutrient composition, with individuals on a higher-protein diet inhibiting disease progression for longer. Using the gram-negative entomopathogenic bacterium Xenorhabdus nematophila, we investigated the nutrition dynamics in the host-pathogen relationship from the perspective of the parasite. Synthetic haemolymphs (bloods) were designed to replicate the in vivo host nutritional state when fed on a range of diets varying in their macronutrient compositions. We found that the in vitro

population growth profiles of X. nematophila cultivated in these synthetic bloods varied in a con-sistent manner, reflecting our in vivo findings; bacterial replication rate decreased inversely with solution protein content. This implies that the insect's preference for dietary protein may not only boost immune function, but may directly inhibit pathogen proliferation, further increasing the host's chances of survival.

Poor growth of Bacillus thuringiensis in larval cadavers

8:45 •139•

Alejandra Zamudio-Ramírez, Javier Luévano-Borroel, Rosalina García-Suárez, A. Nadin Lule-Chávez, Luis A. Verduzco-Rosas, J. Francisco Castillo-Esparza, Mónica García-Montelongo, Sebastián E. González-Villarreal, Leandro G. Ordoñez-Acevedo, Jorge E. Ibarra.

> CINVESTAV-Irapuato, Irapuato, Gto., Mexico Corresponding author: jorge.ibarra@cinvestav.mx

Entomopathogens may be important biotic factors in the regulation of insect population densities in nature and agro-ecosystems. They share important features such as virulence, infectivity, co-evolution with their host, ability to cause epizootics, transmissibility, and a strong relationship with their host's biology. Many show some degree of specificity to their hosts, and their horizontal transmission is based on their ability to massively grow in the host so the cadavers become into inocula to other hosts. Although the most successful entomopatho-gen used in insect control is Bacillus thuringiensis, it does not show some of the typical features of a natural entomopathogen. This report shows that B.t. is deficient in its ability to grow massively in its host, hindering its ability for horizontal transmission. B.t. was quantified in cadavers of Manduca sexta killed by this bacterium, from 0 to up to 12 days after death, and in three different settings: cadavers left on the artificial diet, cadavers removed from the artificial diet after death, and cadavers left on tomato leaves. Cadavers left on the artificial diet showed an increasing number of B.t. that stabilized at about 1 million CFU per cadaver. Cadavers removed after death from the artificial diet decreased rapidly the B.t. numbers to stabilize at about 1,400 CFU per cadaver. When cadavers were left on tomato leaves, the number of B.t. CFU decreased to about 500 CFU per cadaver. In none of these settings, B.t. overgrew saprophytic bacteria in the cadavers. In the experiment with cadavers left on tomato leaves, proportions of Bt:saprophytic bacteria were 2.4:1000 at 0 days after death (DAD); 1.4:1000 at 3 DAD, and 0.3:1000 at 6 DAD. These results clearly indicate a poor growth of B.t. in larvae killed by itself, especially when tested under near-field conditions. These results also may answer some doubts about the ability of B.t. to cause epizootics in nature.

Formulation, field evaluation and commercialization of coleoptera toxic Bacillus thuringiensis



H.M. Mahadeva swamy, R. Asokan

Bio-Pesticide Laboratory, Division of Biotechnology, ICAR-Indian Institute of Horticultural Research, Hesaraghatta Lake Post, Bengaluru 560089 Corresponding author: clintonbio@gmail.com

A formulation with high efficacy and low cost would add to the characteristic features of B. thuringiensis. The native isolate of B. thuringiensis having better Coleopteran insecticidal activity were used for the formulation and field evaluation. Primarily used the amarantus stalk and corn stalk extract as a source for mass production Bt. Both stalk extracts supported growth of native Bt strains. Corn stalk extract yielded 1.12x10⁻⁹ cells in JK Bt IIHR isolate. Whereas Amarantus stalk extract yielded 1.01X10⁻⁹ cells in AND Bt IIHR isolate and 9.58X10⁻⁹ cells/ml in MD native Bt isolate. In the present study, granular, talc and liquid formulations of B. thuringiensis were prepared. The cultures were purified by restreaking on LB agar plates several times and maintained on LB agar slants with 20% glycerol at 4ºC. Liquid formulations of native Bt was prepared by using five different stabilizers, viz., glycerol, groundnut oil, mustard oil, mineral oil, and sunflower oil. Granular formulation was done using sodium bentonite granules. Whereas wettable powder formulation was done using talc powder. Liquid formulation bioassay were conducted against the adults of brinjal ash weevil Myllocerus subfasciatus and M. discolour (Coleoptera: Curculionidae). Granular and talc formulation bioassay were carried out against the larvae of Arecanut white root grubs (Leucopholis lepidophora B. & Leucopholis burmeisteri) and white

grub (Holotrichia serrata) Coleoptera: Scarabaeidae. Formulations were very effective against the tested insects. Successfully conducted the field trial against Arecanut white root grubs and brinjal ash weevil. The Bt based formulation offer advantages over harmful chemical insecticides being target specific, biodegradable and economical.

Screening of four soil bacteria for secondary metabolites (SMs) 9:15 •141• against pulse beetle for post-harvest preservation of green gram

9:30

Shripad M. Upasani

Department of Zoology Arts, Commerce and Science College Dharangaon 425105 MH, India Corresponding author: shripad.upasani@gmail.com

Green gram (Vigna radiata (L) Wilczek) is major constituent of human diet in India. Production is not meeting the demand, hence grains has to protect biologically in storage saving 25% grain being lost every year. Four bacteria namely Bacillus farraginis NAIMCCB-00813, Lysinibacillus sphaericus NAIMCC-B-00111, Stenotrophomonas acetaminiphila NAIMCC-B-00473 and Paenibacillus polymyxa NAIMCC-B-00864 were tested for their secondary metabolites having pesticidal activity. Among four bacteria B. farraginis and Stenotrophomonas acetaminiphila shown well growth and yielded comparatively high amount of secondary metabolites. Pesticidal activity of all crude secondary metabolites checked against Callosobruchus chinensis Linn., which is cultured on Green gram (Vigna radiata (L) Wilczek); in previous study at School of Life Sciences, North Maharashtra University Jalgaon, it has been found to be most susceptible pulses to the infestation of Called Land Land Carraginis shown 95% adult mortality at 6 mg ml in 2 h, Lysinibacillus sphaericus shown 66% adult mortality at 8 mg ml $^{-1}$ in 7 h. Paenibacillus polymyxa shown

acetaminiphila shown 97% adult mortality at 3 mg ml in 1.5 h. Also only 1%, 2%, 10% and 17% adults has been seen emerged in green gram treated by B. farraginis, Stenotrophomonas acetaminiphila, Lysinibacillus sphaericus and Paenibacillus polymyxa secondary metabolites respectively in ovicidal activity. As Bacillus farraginis and Stenotrophomonas acetaminiphila found to be the significant bacteria for management of Callosobruchus chinensis efforts has been made to identify their active ingredients by UVVIS spectroscopy, TLC, HPLC, FTIR and HRLCMS. It has been seen that SMs extracted from bacteria are volatile and their action on beetle is fumigant. Solid formulations have been shown best results in protecting grains while kept in bags of green gram in the simulated grainary conditions.

Molecular Identification, characterization and implementation of Spodoptera litura associated NPV for the management of major lepidopterist insect pests of major crop in Pakistan

> Jam Nazeer Ahmad^{1,2}, Robert L Harrison^{3,} Muhammad Jafir¹ Ishita Ahuja⁴, Atle Bones⁴, Samina Jam Nazeer Ahmad^{1,2}

¹Intergrated Genomics Cellular Developmental and Biotechnology Lab, Department of Entomology, University of Agriculture Faisalabad, Pakistan; ²Plant Stress Physiology and Molecular Biology Lab, Department of Botany, University of Agriculture Faisalabad, Pakistan; ³Invasive Insect Biocontrol & Behavior Laboratory, USDA, ARS, BARC, Baltimore Ave. Beltsville ⁴Department of Biology, Norwegian University of Science and Technology (NTNU), NO-7491 Trondheim, Norway Corresponding author: iamnazire@vahoo.com

The microbial control of major insect pest may serve as a potential replacement of insecticides when insect pests gain resistance against synthetic pesticides. In the present study, for the first time, native Nucleopolyhydroviruses infected insects were collected from fields of one crop growing region of Pakistan and brought to laboratory for identification and laboratory trials. The experiments were conducted to investigate the effect of native Nucleopolyhedroses virus (NPV) isolated locally from field populations of army worm (Spodoptera litura -Lepidoptera: Noctuidae). For molecular study, DNA was isolated from NPV infected insects and used for Polymerase chain reaction by using Lef8, Lef9 and Polh primers pairs, RFLP and then sequencing. Our results indicated that our native NPV were closely related to Spodoptera litura associated NPV (splNPV) already identified from other countries. For laboratory trials, three dose rates of NPV (1 x 10'; 1 x

10⁸; and 1 x 10⁹ POB mL⁻¹) were applied against 2nd and 4th instar larvae spodoptera litura and Helicoverpa armigera reared in our laboratory. The data was recorded after every 24 h for mortality, pupation, adult emergence and egg eclosion for both insects' larval instars. All the bioassays were carried out at 25±2°C, 75% r.h. and L16: D8 h photoperiod. The medium and higher dose rate of NPV for Spodoptera and higher dose rate for Helicoverpa population exhibited good effect against various stages of insects. Our results showed that the army worm population was highly susceptible and life stages were severely affected against identified spINPV. We also found maximum mortality of Helicoverpa population at high dose rate. Moreover, it may be suggested that SpINPV can be equally tested against some other lepidopterist pests. It can also be combined with other microbial mixtures and insecticides that can prove as a potential Integrated Pest Management (IPM) strategy against S. litura and Helicoverpa armigera in crop growing areas of Pakistan.

Eicosanoids act in insect/pathogen interactions: cloning and characterization of a phospholiplase A2 from Heliothis virescens

9:45

David Stanley¹, Cynthia Goodman¹, Joseph A. Ringbauer, Jr¹, Tamra Reall¹, Yongguyn Kim²

¹USDA/ARS Biological Control of Insects Research Laboratory, Columbia MO 65203, Department of Bioresource Sciences, ²Andong National University, Andong, Korea Corresponding author: David.Stanley@ars.usda.gov

Phospholipase A₂s (PLA₂s) make up a large superfamily of enzymes responsible for hydrolyzing fatty acids from the sn-2 position of phospholipids. They are very well appreciated in the biomedical literature and understudied in insect species. PLA2s act in insect venoms and in a range of physiological functions, including digestion and as the first step in the biosynthesis of prostaglandins and other eicosanoids, which are key mediators of cellular and humoral insect immune reactions to pathogens and other microbes. Here, we report on cloning, expression, and characterization of a recombinant PLA2 from H. virescens tissues. We also characterized PLA2s in two insect cell lines, a Heliothis zea (HzAM1) and a H. virescens (HvAM1) line. The three PLA₂ preparations were sensitive to the usual biophysical parameters, source protein concentration, reaction time, temperature and pH. Reactions in the presence of selective secretory and intracellular PLA₂ inhibitors indicate most of the activity in the cell lines is due to cellular PLA₂s. The cloned gene encodes a 202-amino acid protein that includes a signal peptide, a calcium-binding domain, and a catalytic site. Phylogenetic analysis shows it is a new PLA2 that clusters into the secretory PLA2s. Because PLA2 is the first step in eicosanoid biosynthesis, we infer that PLA2s are crucial elements in insect/pathogen relationships.

10:00-10:30 am

Coffee Break

WEDNESDAY, 10:30 am - 12:30 pm

Theater

Virus Division #3

Moderators: Katsuhiko Ito, Ikbal Agah Ince

10:30 •142• Ascovirus P64 homologs: A novel family of large cationic proteins that condense viral genomic DNA for encapsidation

> **Dennis K. Bideshi**^{1, 2}, Tatsinda Spears³, Heba Zaghloul⁴, Yeping Tan¹, Yves Bigot⁵, Brian A. Federici^{1,3,4}

¹Dept. of Entomology, University of California, Riverside, California, 92521; ²California Baptist University, Dept. of Biological Sciences, Riverside, California; ³Graduate Programs in Cell, Molecular and Developmental Biology; ⁴Microbiology, University of California, Riverside, California; ⁵UMR CNRS7247, INRA Val de Loire, 37380 Nouzilly, France Corresponding author: dbideshi@calbaptist.edu

Most eukaryotic dsDNA viruses use small virus or host protamine-like proteins or histones to condense their genomic DNAs (gDNAs) during virogenesis. These arginine- and lysine-rich proteins are highly basic and typically less than ~20 kDa. Ascoviruses are large dsDNA (~125-186 kbp) viruses that cause a chronic and fatal disease in lepidopteran larvae. At present, little is known about the molecular basis for condensation and encapsidation of their large gDNAs. Previous proteomic and functional studies showed that Spodoptera frugiperda ascovirus (SfAV-1a) virions lack small cationic proteins, but instead contain a relatively large novel viruscoded DNA-binding protein (P64; 64 kDa, pI = 12.2) proposed to condense its gDNA during virion maturation. The N-terminal domain of P64 contains four virus-specific 2-cysteine adaptor motifs, whereas the C-terminal domain has 14 modular repeats rich in arginine and serine that potentially mediate gDNA sequestration. In this study, we used centrifugation, electrophoretic mobility shift assays and transmission electron microscopy to demonstrate P64's basic C-terminal domain condenses SfAV-1a gDNA. We further show that P64 homologs occur in other ascoviruses. Collectively, our data indicate P64 homologs are a novel family of viral gDNA condensing proteins.

Transcriptome analysis of the Spodoptera frugiperda ascovirus in vivo provides insights into how its apoptosis inhibitors and caspase promote increased synthesis of viral vesicles and virion progeny

Heba Zaghloul¹⁴, Robert Hice¹, Peter Arensburger³, Brian A. Federici¹²

¹Interdepartmental Graduate Program in Microbiology and Institute for Integrative Genome Biology, ²Department of Entomology, University of California, Riverside, Riverside, CA 92521; ³California State Polytechnic University, Pomona, Department of Biological Sciences, 3801 West Temple Avenue, Pomona CA 9176:. ⁴Department of Botany and Microbiology, Faculty of Science, Alexandria University, Egypt, Corresponding author: hzagh001@ucr.edu

STU 10:45 •143•

Ascoviruses are ds DNA insect viruses that primarily attack caterpillars of the family Noctuidae. Ascovirus virions are large (400 x 150 nm) and typically transmitted on the ovipositor of endoparasitic wasps during egg laying. However, the most unique characteristic of ascoviruses is their novel cytopathology, whereby after infection the nucleus lyses, after which the cell cleaves into a cluster of vesicles in which virions continue to assemble after the vesicles move from infected tissues into the hemolymph. A virusencoded caspase induces this unique cytopathology, but other genes involved in pathogenesis and viral vesicle formation remain unknown. Suitable cell lines that reproduce typical ascovirus vesicles in vitro have not been developed. Thus, we used strand-specific RNA-Seq analysis to study transcription of the type species, Spodoptera frugiperda ascovirus-1a (SfAV-1a), in vivo during infection of S. frugiperda 3rd instars. We focused on 44 core ascovirus genes and 26 others putatively involved with viral vesicle membrane formation. Differential expression patterns revealed three temporal classes, namely, early, late and very late. Three genes that inhibit apoptosis (IAP-like) were expressed early, whereas the caspase gene was expressed very late. This correlates well with apoptotic events associated with viral vesicle formation. A Diedel-homolog gene, the only known virokine, was also highly expressed, implying this protein may help SfAV-1a evade its host's innate immune system. Interestingly, 15 bi-and poly-cistronic messages were also identified in the SfAV-1a transcriptome, indicating possible usage of non-canonical translation mechanisms by ascoviruses.

Expression profile of nsd-2 gene encoding the putative Bombyx mori bidensovirus receptor after virus infection 11:00 •144•

Katsuhiko Ito¹, Takeshi Fijii¹, Takeshi Yokoyama¹,Keiko Kadono-Okuda²

¹Department of Science of Biological Production, Tokyo University of Agriculture and Technology: ²Institute of Agrobiological Sciences, National Agriculture and Food Research Organization Corresponding author: katsuito@cc.tuat.ac.jp

Bombyx mori bidensovirus (BmBDV) is a pathogen that replicates only in the midgut columnar cells and causes fatal disease in the silkworm. The resistance to BmBDV is determined by a single gene, nsd-2 (resistance gene), which is charac-terized as non-susceptibility irrespective of the viral dose.

Previously we have identified nsd-2 by positional cloning using B. mori genome information and found that this gene encodes a putative amino acid transporter which may work as a receptor for BmBDV. In this study, to better understand the relationship between BmBDV and the putative virus receptor, we performed an expression analysis of viral transcripts and receptor, +ⁿ (susceptibility gene), in the midgut after virus infection. We first fed an artificial diet smeared with a BmBDV suspension to newly ecdysed fourth instar larvae (susceptible strain for BmBDV), and investi-gated the expression pattern of the BmBDV-derived transcript in the midgut every day. The quantitative RT-PCR analysis showed the unique expression pattern; the expression of the viral transcript increased soon after viral infection and then decreased until the end of the fourth instar, and then the viral transcript increased again after becoming the fifth instar. Next, we compared the expression pattern of + nsd-2 between the midgut of the virus-infected and uninfected silkworm. The qRT-PCR analysis revealed that the expression levels of +^{nsd-2} drastically decreased from the fifth instar in the midgut of the virus-infected larvae. On the other hand, there was no change of the expression levels of $+^{nsd-2}$ in the midgut of the virus-uninfected larvae. Additionally, we investigated the expression patterns of other genes whose expression in the midgut have been already reported, but similar differences were not detected. These results suggested that BmBDV might specifically regulate the expression level of the virus receptor, $+^{nsd-2}$, during infection. We believe that a study of the relationship between virus and receptor will contribute to elucidation of the BmBDV infection mechanism.

The transcription strategy of Bombyx mori Bidensovirus and a characterization of the viral structural proteins

11:15 •145•

Qian Yu, Peng Lü, Yali Xing, Qin Yao

Institute of Life Sciences, Jiangsu University, Zhenjiang, China Corresponding author: yaoqin@ujs.edu.cn

Bombyx mori bidensiovirus (BmBDV) is a species of Bidensovirus that has been was placed into a new genus within the new family Bidnaviridae by the International Committee on Taxonomy of Viruses. BmBDV causes fatal flacherie disease in silkworms, which causes large losses to the sericulture industry. BmBDV contains two sets of complementary linear single-stranded DNAs of approximately 6.5 kb (viral DNA 1, VD1) and 6.0 kb (viral DNA 2, VD2). VD1 and VD2 are encapsidated in separate icosahedral non-enveloped capsids, which are similar in size and shape. However, the complete mRNA procession profile remains unclear. In this work, we investigated the expression strategies of BmBDV non-structural proteins and structural proteins. Total RNA was obtained from the midgut of infected silk worm at 5th instar. RACE methods were used to identify the 5' and 3' transcription ends. Meanwhile, a total of six structural proteins were separated by twodimensional electrophoresis and shown to be encoded by the BmBDV VP gene via mass spectrometry. The transmission electron microscopy results showed that co-expression of the BmBDV VP and SP structural proteins in Spodoptera frugiperda sf9 cells resulted in the formation of 22-24 nm viruslike particles. Furthermore, a mutation of the major structural proteinencoding VP gene, in which the second in-frame ATG codon was mutated to GCG, abrogated the production of several structural proteins, indicating that this strategy of expressing BmBDV VP is dependent on a leaky scanning translation mechanism.

Cloning and rescue of the genome of Bombyx mori bidensovirus and characterization of a recombinant virus

STU 11:30 •146•

Panpan Zhana, Di Miao, Qin Yao

Institute of Life Sciences, Jiangsu University, Zhenjiang, China Corresponding author: yaogin@ujs.edu.cn

Bombyx mori bidensovirus (BmBDV) is the first segmented ssDNA virus identified in insect, which was classified into the Bidnaviridae family by International Committee on Taxonomy of Viruses (ICTV) in 2012. In this study, we constructed a full-length genomic clone of BmBDV which contains the complete terminal structure. A recombinant BmBDV that expressed the green fluorescence protein (GFP) gene was constructed. Then, the insect cell xx was transfected with the linearized genome using continuous culture and

expanded cell culture methods. Western blotting analysis detected the expression of the major structural protein, VP, and green fluorescence was observed in infected cells. Furthermore, re-infected silkworm larvae showed typical densonucleosis symptoms, and more importantly, the direct visualization of green fluorescence allowed us to detect recombinant plasmids that were expressed in the silkworm midgut. These results suggest that infectious virus particles have been rescued from the infected BmN cells after co-transfection of the linear genome. We also found that BmBDV cannot transform mammalian cells, which means that BmBDV is safe to mammals. Overall, this study showed BmBDV has the potential to be used as biological pesticides, and also provided a platform for researching viral replication mechanisms, viral gene functions, and viral infection mechanisms.

Study the interaction between Bombyx mori bidensovirus STU 11:45 •147• structural protein and the host protein + nsd-2



Yahui Zhang, Peng Lü, Qin Yao

Institute of Life Sciences, Jiangsu University, Zhenjiang, China Corresponding author: yaoqin@ujs.edu.cn

Bombyx mori bidensovirus is major pathogen that causes chronic densonucleosis disease of silk worm. In 2012, the International Committee on the Taxonomy of Viruses (ICTV) established a new family, Bidnaviridae, and designated BmBDV as the type species in the new genus Bidensovirus. VP as the major structural protein of BmBDV could decide the host specificity by structural protein interaction with the host receptor. The minor capsid protein P133 closely related to host recognition and the virus into the host. The lacking of the + nsd-2 gene encoded trans-membrane protein could make the silkworm resistance from BmBDV infection, showing the + nsd-2 gene may contain the recognition site of BmBDV. However, the mechanisms of the interaction between BmBDV and membrane protein are still unclear. In this study, a BmN cell strain stably expressing the host protein + nsd-2 has been successfully established. The expression plasmids pIB-HA-VP and pIB-FLAG-P133 were then transfected into the BmN($+^{nsd-2}$) cell strain, respectively. The interaction between BmBDV structural protein and the host protein + nsd-2 was analyzed by Co-IP and pull-down technics. Co-IP results revealed that structural protein VP of BmBDV could interact with the host + nsd-2 protein. Morevoer, these results reveal a proof of concept for the infecting mechanism of the host cell by BmBDV at the molecular level.

Heliothis zea Nudivirus-1 miRNAs promote latent infection via epigenetic regulation

STU 12:00 •148•

Pei-Chi Wu1, Yueh-Lung Wu1

Department of Entomology, National Taiwan University, Taiwan Corresponding author: runwu@ntu.edu.tw

Heliothis zea nudivirus-1 (HzNV-1) is a7n unique insect virus that can induce both lytic and latent infections in many insect cell lines. During latent infection, the persistency-associated gene 1 (pag1) expresses several microRNAs (miRNAs) and is the only detectable HzNV-1 gene. Two of the miRNAs derived from pag1 have been found to suppress and silence HindIII-I 1 (hhi1), an early gene of HzNV-1. Hhi1 induces strong apoptosis and increases the lytic to latent ratio. Pag1 reduces the apoptosis caused by hhi1 and further promotes the establishment of latent infections. Previous reports have shown that persistent infection of herpes simplex virus (HSV) is highly associated with epigenetic regulation of the chromatin structure and is regulated by latent-specific miRNAs. Since pag1 is a non-coding transcript, it could regulate host chromatin structures through miRNAs to further establish latent infections. In this study, we found that histone modification associated enzymes changed upon pag1 transfection. We then tested the histone acetylation/methylation protein expression levels through Chromatin Immunoprecipitation (ChIP) assay to investigate the histone modifications of insect hosts. This study not only provides a better understanding of the HzNV-1 infection mechanism, but also the relationship between viral miRNAs and their hosts.

Functional analysis of non-polyadenylated invertebrate iridovirus mRNAs: The role of a CAUUA-containing hairpin 12:15 •149•

Discovery of new insecticidal traits

10:45 •151

Ikbal Agah Ince¹, Zeynep Kanlıdere¹, Ayca Zeynep İlter Akülke¹, Arzu Özgen^{1,2}

¹Department of Medical Microbiology School of Medicine, Acibadem University, 34752, Atasehir, Istanbul, Turkey; ²Vocational School of Health, Istanbul Gelisim University, 34310, Avcılar, Istanbul, Turkey Corresponding author: agah.ince@acibadem.edu.tr

It is observed that the transcripts of Invertebrate iridescent virus 6 (IIV6) are not polyadenylated, in line with the absence of canonical polyA motifs (AATAAA) downstream of the open reading frames (ORFs) in the genome. Previously, we used ligation-based amplification of cDNA ends (LACE) to determine the 3' ends of IIV6 the transcripts of the virion protein genes from infected Drosophila Schneider 2 (S2) cells. Many of the transcripts ended directly after a CAUUA motif (CATTA in cDNA). In silico analysis showed that the complementary motifs CATTA and TAATG were strongly enriched in the 100 base pair regions downstream of the IIV6 ORFs in comparison to their presence in the whole viral genome. When we have investigated the hairpin formation at 3'end of individual ORFs in silico among the related species suggests that a 3' hairpin sequence, often with a CAUUA motif, is a conserved feature in invertebrate-infecting iridoviruses (IIV3, IIV 9 and IIV31). The CAUUA motif as part of the stem-loop structure is likely to determine the pre-mRNA cleavage, or alternatively the direct termination point. We hypothesized that this motif may play a role in transcript 3'end formation. For the functional analysis of the hairpin structure, we selected two CATTA containing virion protein genes, 274L and 261R, to unravel the role of the conserved CAUUA motif in 3' hairpin structure formation using site directed mutagenesis. Each 3' UTR region is sub-cloned into pIZ vector into the downstream region of firefly luciferase gene as a phusion driven by Opie2 promotor. Single or double mutations introduced to CATTA motif on the 3' UTRs. We have examined whether these mutations on the CATTA motif affects viral 3' end formation or not.

Roosevelt Room

Microbial Control Division #3

Moderator: Trevor Jackson, Stefan Jaronski

Global patterns of field-evolved resistance to Bt crops:Successes and failures

10:30 •150•



Bruce E. Tabashnik 1, Jeffrey A. Fabrick 2, Yves Carrière 1

¹University of Arizona, Department of Entomology, Tucson, AZ, USA; ²USDA ARS, U.S. Arid Land Agricultural Research Center, Maricopa, AZ, USA Corresponding author: brucet@cals.arizona.edu

From 1996 to 2016, farmers worldwide planted a cumulative total of >830 million hectares of genetically engineered corn, cotton, and soybean that produce insecticidal proteins from the bacterium Bacillus thuringiensis (Bt). These Bt crops kill some key insect pests, yet are not toxic to most other organisms including people. Bt crops can suppress pests, reduce insecticide sprays, and increase farmer profits. However, their benefits are diminished or lost when pests evolve resistance. Here we review data monitoring resistance to nine Bt proteins in 15 major pest species targeted by Bt crops on six continents. We report a surge since 2005 in cases of resistance with practical consequences for pest management in the field. This surge in practical resistance is associated with cross-resistance, increased cumulative exposure to Bt crops, and increased monitoring. In addition, practical resistance to Bt crops is associated with non-recessive inheritance of resistance and a scarcity of refuges of host plants that do not produce Bt proteins. To maximize the benefits of Bt crops, we encourage implementation of large refuges of non-Bt host plants together with alternative control tactics, particularly when inheritance of resistance is not recessive and alleles conferring resistance are not rare.

Rebekah Kelly, Vadim Beilinson, Mary Kroner and AgBiome team AgBiome Inc., 108 T.W. Alexander Dr. Bldg 1, RTP, NC, USA

Corresponding author: rkelly@agbiome.com

Insect pests are a serious threat to agricultural production, and insecticidal proteins are among the most promising tools for limiting the damage they cause. Since the commercialization of biotech crops in 1994, however, only a small number of insecticidal proteins have made it to market. Moreover, emerging pest resistance to the current products points to a pressing need for new and unique modes of action. AgBiome is working to fill that need by discovering and screening plant-associated microbes to provide new solutions for insects, nematodes, and disease. We have built a large and diverse proprietary strain collection of over 40,000 fully sequenced microbial isolates. These microbial isolates are the foundation of our Biological and Trait discovery programs. Using our innovative microbial-capture and geneidentification technology, we have discovered more than 3,000 potential insecticidal genes to test in our high-throughput Lepidopteran, Coleopteran, and Hemipteran assays. Currently, we have identified over 20 active genes that are being evaluated for new modes of action and efficacy in plants.

Exploiting mosquito biology with transgenic Metarhizium pingshaense

STU 11:00 •152•

Brian Lovett¹, Etienne Bilgo², Abdoulaye Diabate², Raymond St. Leger¹

¹University of Maryland, College Park, Maryland, USA; ²Institut de Recherche en Sciences de la Santé/Centre Muraz, Bobo-Dioulasso, Burkina Faso Corresponding author: lovettbr@umd.edu

We compared arthropod genes encoding insect-specific sodium, potassium and calcium channel blockers for their ability to improve the efficacy of Metarhizium against insecticide-resistant anophelines. Some of our most potent strains, engineered to express multiple toxins, are able to kill mosquitoes with a single spore. A strain expressing the EPA approved Hybrid (Ca++/K+ channel blocker), was studied for efficacy in a MosquitoSphere in a malaria endemic region of Burkina Faso. Compared to the wild-type fungus, Met-Hybrid killed anopheline mosquitoes in half the time and at much lower spore doses in semi-field conditions, which increased the percent of lethally infected mosquitoes and the effective persistence of the pathogen. In the MosquitoSphere, we can reliably use males to deliver a fatal dose of transgenic fungus to female mosquitoes during mating: revealing an avenue of application that would easily integrate with established vector control techniques. Importantly, Met-Hybrid had an outsized reduction on blood feeding in infected mosquitoes compared to wild-type fungus. These lethal and pre-lethal effects of Met-Hybrid prevent >90% of disease transmission in just 5 days, surpassing the WHO threshold for successful vector control agents. This NIH funded, international effort represents an important step in the progression of transgenic mosquito control technologies into field application.

Functional response of the predator Chrysoperla carnea STU 11:15 •153• (Stephens) in an endophytic entomopathogenic fungus aphid management system

Natalia González-Mas, María Cuenca-Medina, Enrique Quesada-Moraga Departamento de Ciencias y Recursos Agricolas y Forestales, Universidad de Córdoba Corresponding author: cr2qumoe@uco.es

The recently discover endophytic behavior of entomopathogenic fungi (EF) have led to complementary uses in horticulture complementary to the welldeveloped spray applications of commercially available mycoinsecticides. Fungal suspension spraying, seed dressing and even soil drenching with EF conidia may lead to endophytic colonization, whereas the possible effect of such endophytic colonization targeting the pest on the trophic cascades remains unknown. The trophic cascade effects of endophytic colonization of melon leaves targeting Aphis gossypii on attraction, settling and predatory capacity of the generalist predator Chrysoperla carnea have been investigated by offering Beauveria bassiana challenged aphids, either by direct spray with a fungal suspension or by feeding on endophytically colonized melon plants. Results shown that the proportion of Chrysoperla carnea larvae that prefer fungal colonized plants (70%) was higher when

compared with control (30%) three hours after the preys were offered (χ^2 =0.025). However no differences were observed in the average number of herbivorous prey eaten by the predators (P<0.05). Our results demonstrate that fungal colonization has an impact on natural enemy's behavior attracting them to entomopathogenic fungal colonized plants which could improve their efficacy when these two techniques of biological control are applied simultaneously. Aphid control strategies based on the use of endophytically colonized plants are apparently safe for predators, whereas further research is needed to evaluate the overall success of IPM strategies based on the simultaneous use of these biocontrol agents.

Potential of entomopathogenic nematodes Rhabditida: Steinernematidae, Steinernema carpocapsae and Heterorhabditidae, Heterorhabditis bacteriophora for control of Planococcus citri (Risso) (Hemiptera: Pseudococcidae) in Georgia

11:30 •154•

Nona Mikaia

Tbilisi, Georgia; Sokhumi State University Department of Natural Sciences and Health Care Corresponding author: nonamikaia@gmail.com

Planococcus citri, the citrus mealybug, is the most important species of mealybug known to infest citrus in West Georgia. Various laboratory bioassays were conducted to determine the potential of entomopathogenic nematodes to control P. citri. Adult female P. citri were screened for susceptibility to two introduced from Germany nematode species. Planococcus citri was found to be most susceptible to Steinernema carpocapsae and Heterorhabditis bacteriophora, causing 92% and 71% mortality, respectively. The development of nematodes after infecting adult female P. citri showed both H. bacteriophora and S. carpocapsae were able to complete their life cycles inside the host. Further bioassays illustrated a linear relationship between mealybug mortality and the concentration of nematodes applied, with the highest level of control using a concentration of 150 infective juveniles (IJs)/insect. As nematodes would be used as an aboveground application to control P. citri in citrus orchards, available water is a major limiting factor. Insecticidal activity proved to be dependent on the available surface moisture after nematode application. The water activity (aw) bioassay indicated that S. carpocapsae to be two times more tolerant to lower levels of free water, compared to H. bacteriophora. After application, nematodes have a limited time frame in which to locate and infect hosts, as the level of available free water gradually decreases, as trees dry out. Steinernema carpocapsae proved able to locate and infect P. citri quicker than H. bacteriophora. Nematode activity was not significantly affected when exposed to 20°C and 25°C. IJs were able to infect P. citri at an exposure time as short as half an hour. Results also showed that the first 2 to 3 h application is the most decisive time for establishing successful infection of mealybugs by potential use of nematodes for the control of P. citri.

A physiological time model to predict the virulence of entomopathogenic fungi under conditions of fluctuating temperature



Declan Perry, Dave Chandler

Warwick Crop Centre, University of Warwick UK Corresponding author: D.Perry@warwick.ac.uk

Temperature determines the speed of kill of entomopathogenic fungi (EPF) used as biopesticides. Unfortunately, little research has been done to understand and predict how biopesticide efficacy is affected by the varying temperature conditions that are typical of many field crop environments, and this is preventing us making best use of fungal biopesticides in IPM. Moreover, because candidate EPF strains are often selected in screens done at only one temperature, we may also be failing to identify 'winning' strains that are best adapted to variable field conditions. As part of a wider project to investigate EPF for the control of the diamondback moth (Plutella xylostella L.) (DBM), we have investigated thermal- and physiological-time models to understand and predict EPF virulence under varying temperatures. A number of non-linear statistical models used to describe biological processes in ectotherms were fitted to data on the effect of temperature on: (i) DBM larval development rate; (ii) growth and germination of different EPF

strains pathogenic to DBM larvae; and (iii) fungal virulence to DBM larvae (measured as speed of kill and lethal dose). Of these, we found that a particular model described by Briere (1999) gave the best fit overall and allowed the cardinal temperatures to be identified with greater precision than before now. A plot of the relationship between the optimum temperature for virulence vs. the optimum for germination was not significantly different from zero (linear regression, P = 0.13). Fungal isolates differed in their thermal profiles for growth, germination and virulence, and we were able to use the model to select EPF strains whose thermal virulence 'profile' closely matched that for DBM development. Next, a day-degree model was developed, using mortality data at fixed temperatures, to predict DBM mortality at various variable temperature regimes. A simple DD model based on a sigmoidal equation was accurate in predicting mortality during a variable temperature regime over seven days. These results help to understand and address the sources of variability when taking EPF from the lab to field and should be a valuable tool for biopesticide development.

Plant cell-wall degrading enzymes improve endophytic STU 12:00 •156• establishment of Metarhizium brunneum in potato plants

Vivien Krell¹, Desireé Jakobs-Schoenwandt¹, Stefan Vidal², Anant V. Patel¹

¹ Bielefeld University of Applied Sciences, Faculty of Engineering Sciences and Mathematics, Bielefeld, Germany; ²Georg-August-University Goettingen, Department of Crop, Sciences/Agricultural Entomology, Goettingen, Germany Corresponding author: anant.patel@fh-bielefeld.de

Biocontrol with endophytic entomopathogenic fungi is a promising option to protect plants systemically from insect herbivores. Yet, current applications are limited especially by low plant colonization. Inspired by penetration mechanisms of plant pathogenic fungi, we aimed at enhancing potato plant penetration and colonization by the endophytic entomopathogenic Metarhizium brunneum strain Cb15 through supplementation of plant cell-wall degrading enzymes pectinase and cellulase or corresponding substrates into beads containing mycelial biomass. We found that after bead application to potato tubers and incubation at 18-23 °C with a LD cycle of 16:8 for 21 days, M. brunneum was re-isolated from surface-sterilized roots, tubers, and shoots in 75.0 ± 9.4 %, 33.3 ± 9.8 %, and 29.2 ± 15.1 % of samples, respectively. Additional verification of re-isolated M. brunneum was conducted with gPCR. Pectinolytic and cellulolytic enzymes were successfully induced by addition of corresponding substrates, but activity levels were low and no correlation between enzymatic activity and fungal penetration was found. However, incorporation of cellulase into beads led to a substantial increase in plant penetration by 25.0 % in roots, 54.2 % in tubers, and 16.6 % in shoots. This was accompanied by a 3.0-fold enhanced spore formation on the surface of beads to 1.91 x 10⁸ ± 0.26 x 10⁸ per bead. Finally, a stronger root development of treated plants was observed indicating a fertilizing effect mediated by the formulation. This study will pave the way to understanding and eventually use of enzymes as penetration aid for endophytes.

Biological control of ambrosia beetles that vector laurel wilt fungus in avocado using entomopathogenic fungi

Pasco B. Avery¹, Daniel Carrillo², Rita H. Duncan¹,

12:15 01570

Alison Lukowsky², Verónica B. Fuentes¹, Cecilia Gámez^{1, 3}, Ronald D. Cave¹

¹University of Florida, IFAS, Indian River Research and Education Center, Ft. Pierce, Florida, USA: ²University of Florida, IFAS, Tropical Research and Education Center, Homestead, Florida, USA: ³Escuela Agrícola Panamericana El Zamorano, Honduras Corresponding author: pbavery@ufl.edu

Laurel wilt (LW) is a disease threatening the avocado industry in Florida. The causative agent of the disease is a fungus vectored by ambrosia beetles that bore into the trees. Until recently, management strategies for the vectors of the laurel wilt fungus relied solely on chemical control and sanitation practices. Beneficial entomopathogenic fungi (EPF) are the most common and prevalent natural enemies of pathogen vectors. Laboratory experiments demonstrated that commercial strains of EPF can increase mortality of the

primary vector of the pathogen, *Xyleborus glabratus*, and alternative vectors, *Xylosandrus crassiusculus, Xyleborus volvulus, and X. bispinatus* (Coleoptera: Curculionidae: Scolytinae). Field experiments revealed that EPF do not prevent beetles from boring into the trees. However, EPF-infected beetles die inside the trees without reproducing, providing control similar to that by chemical insecticides but persisting longer on the bark of avocado trees. To improve efficacy, EPF must be integrated into the management strategy for LW and other avocado pests, which includes the use of several fungicides, insecticides, and adjuvants. We determined antagonistic and synergistic relationships among EPF and agrochemicals used by avocado growers. Several producers have incorporated this biocontrol strategy to manage ambrosia beetles and other pests of avocado. The main limitation of EPF is they do not control ambrosia beetles already breeding inside trees. The search for improved delivery systems that could increase efficacy of EPF will be discussed.

WEDNESDAY, 12:30 – 1:30 pm LUNCH BREAK

Roosevelt Room

Nematode Division - Business Meeting

The Forum

Student Workshop on Science Communication

The road to more effective science communication

12:30 •158•

Bruce Lieberman

¹ Bruce Lieberman, Science Writer & Editor

Presenter: bruce.lieberman@yahoo.com; www.blieberman.com

In this workshop, Bruce Lieberman will review how scientists can better communicate the significance, the details, and the impact of their work to a lay audience. He'll share examples of clear and concise science writing, by scientists and journalists alike, and offer tips on how researchers can make their work more understandable and compelling to anyone. Bruce will also discuss how scientists can effectively interact with media professionals, whether reaching out to news organizations through their institution's public affairs office or calling a reporter or editor directly. He'll also cover some of the changes underway in the media business, how these changes impact a scientist's ability to share their work with the public, and how a scientist can reach out directly to people through social media. As part of the workshop, Bruce will conduct a short exercise during which a few volunteers will be asked to describe what they're working on now; the group will then review these brief presentations. Finally, all attendees will leave the workshop with handouts on what's been covered, including practical tips they can take with them, written examples of great science writing, and web links to other valuable resources on science communication.

Bruce Lieberman is a freelance science writer and editor with more than 25 years of experience in the news business. He worked as a reporter at daily newspapers for many years before becoming an independent writer and editor in 2010. Bruce has written about astronomy, climate change, and biology for national publications and online. He has also worked on science writing projects for universities, biomedical labs, and foundations. As an editor, Bruce has worked with state and regional government agencies throughout California, and he has consulted with them to improve written communications. Bruce earned his Master's Degree at Columbia University's Graduate School of Journalism in 1998, and his Bachelor's Degree in Rhetoric from the University of California at Berkeley in 1988.

WEDNESDAY, 1:30 - 3:30 pm

Theater

Virus Division #4

Moderators: Monique van Oers, Johannes Jehle

SIP EARLY CAREER AWARD WINNER

Insane in the brain: how baculoviruses manipulate brain function and behavior in caterpillars

.12 2

Yue Han¹, Hanneke Suijkerbuijk^{1,2}, Hans M. Smid², Monique M. van Oers¹, **Vera I.D. Ros**¹

¹Laboratory of Virology, Wageningen University,
Droevendaalsesteeg 1, 6708 PB, Wageningen, The Netherlands;
²Laboratory of Entomology, Wageningen University,
Droevendaalsesteeg 1, 6708 PB, Wageningen, The Netherlands

Corresponding author: vera.ros@wur.nl

Modification of host behaviour is a widely adopted strategy of viruses to enhance their own transmission. The examples of behavioural manipulation are rapidly accumulating, covering a broad spectrum of viruses and hosts. Nevertheless, surprisingly little is known on the underlying causative physiological, neuronal, hormonal or molecular mechanisms. A typical case of behavioural manipulation is found in insects infected with baculoviruses. Infected caterpillars show enhanced mobility and climb to the top of plants or the forest canopy prior to death, a phenomenon known as 'tree-top disease'. As a consequence, the virus is spread over a larger area, thereby increasing the chance to infect a new caterpillar. The baculovirus-insect system provides an excellent platform to study viral manipulation of insect host behaviour. It allows the comparative analysis between wildtype viruses and single gene knock-out mutants. I will review the current knowledge on the mechanism of baculovirus-induced behavioural manipulation, including an overview of viral genes found to modify host behaviour and how these may affect host pathways. I will show data on virus spread in the larval brain and discuss how the virus might manipulate host behaviour by affecting the host central nervous system.

MAURO MARTIGNONI AWARD WI<u>NNER</u>

Timely trigger of zombie behaviour: the importance STU 2:00 •160• of light exposure timing in baculovirus-induced tree-top disease

Yue Han¹, Stineke van Houte^{1,2}, Monique M. van Oers¹, Vera I.D. Ros¹

Laboratory of Virology, Wageningen University, Droevendaalsesteeg 1,
6708 PB Wageningen, the Netherlands; ² Centre for Ecology and Conservation,
Biosciences, University of Exeter, Penryn, Cornwall TR10 9FE, UK

Corresponding author: vue.han@wur.nl

Host behavioural manipulation is a common strategy used by parasites to enhance their survival and/or transmission. Baculoviruses induce hyperactivity and tree-top disease (pre-death climbing behaviour) in their caterpillar hosts. However, little is known about the underlying mechanisms of this behavioural manipulation. Previously, we showed that the baculovirus Spodoptera exigua multiple nucleopolyhedrovirus (SeMNPV) induced tree-top disease at 3 days post infection in third instar S. exigua larvae and that light from above plays a key role in triggering this behaviour. Here we further investigated the temporal requirements for the presence of light to trigger this behaviour and found that light from above was needed between 43-50 hours post infection to induce tree-top disease. Infected larvae that were not exposed to light from above in this period finally died at low positions. Exposure of larvae to light prior to this period did not affect the final positions where they died. We hypothesize that virus achieve this via invading host central nervous system. Currently, we are comparing transcriptomes from larval heads between manipulated and non-manipulated groups to identify host/viral molecules that potentially are involved in this process. Overall we conclude that light in a particular time frame is needed to trigger SeMNPV-induced tree-top disease in S. exigua larvae.

The baculovirus *per os* infectivity factors form a complex to resist proteolytic degradation by larva-derived alkaline proteases

STU 2:15 •161•

proteases Bob Boogaard¹, Jan W.M. van Lent¹, David A. Theilmann²,

Martin A. Erlandson³, Monique M. van Oers¹

1Laboratory of Virology, Wageningen University and Research, the Netherlands; 2Summerland Research and Development Centre, Agriculture and Agri-Food Canada, Summerland, British Columbia, Canada; 3Saskatoon Research and Development Centre, Agriculture and Agri-Food Canada, Saskatoon, SK, Canada Corresponding author: bob.boogaard@wur.nl

Baculoviruses orally infect caterpillars in the form of occlusion-derived viruses (ODVs). The ODV envelope contains a number of proteins essential for oral infectivity, called per os infectivity factors (PIFs). PIF1, PIF2, PIF3 and PIF4 form a stable complex. The P74 protein (PIF0) associates loosely with this complex, but is not needed for its formation in contrast to the other constituents. We show here that PIF-proteins are prone to proteolytic degradation when they are not part of an intact complex. When wildtype ODVs released from larva-derived occlusion bodies (OBs) were analysed, both the complex and PIF monomers were found. However, in ODVs from a p74-deletion mutant, only the complex was found and no PIF monomers. But when these ODVs were released from OBs isolated from cultured insect cells or heat-treated larva-derived OBs, the PIF monomers were found again, suggesting that larva-derived alkaline proteases degraded the monomers. Disappearance of PIF monomers was also observed in larva-derived pif2-, pif3-, pif4- and pif6-deletion mutants. The PIF monomers were found again in these mutants when the OBs were heat-treated prior to ODV release, just as previously observed. So baculoviruses might protect their PIFs against insectderived alkaline proteases by complex formation, to safeguard oral infectivity.

Silencing chitin deacetylase, insect intestinal mucin and response to pathogen genes in *Spodoptera littoralis* (Lepidoptera:Noctuidae): Increases and decreases in the biological activity of baculoviruses

Umut Toprak^{1,2}, Serife Bayram¹, Dwayne Hegedus²

2:30 •162•

¹Ankara University, Molecular Entomology Lab. Faculty of Agriculture, Dept. of Plant Protection, Ankara, Turkey; ²Agriculture and Agri-Food Canada, Saskatoon Research Centre, Saskatoon, SK Canada Corresponding author: utoprak@agri.ankara.edu.tr

Baculoviruses are invertebrate-specific viruses with great potential for use as a biopesticide. Upon ingestion of the occlusion bodies by an insect, they dissolve and virions are released in the midgut. In order of virions to initiate an infection in midgut epithelial cells, they must first pass the peritrophic matrix (PM), an inner porous tube serving as a barrier against pathogen invasion within the midgut. PM is composed of chitin microfibrils and multiple proteins. Previous studies showed that several proteins such as chitin deacetylases (CDAs), insect intestinal mucins (IIMs) and response to pathogen (REPAT) interfered with the baculovirus infections. We recently showed that various CDAs, IIMs and REPATs are tightly associated with the PM in Spodoptera littoralis (Lepidoptera: Noctuidae), a destructive pest of vegetables and field crops in Mediterranean Region Countries. In order to understand the potential role of these proteins in the PM, we attempted to silence genes encoding a CDA (SpliCDA823), an IIM (SpliIIM3594) and a REPAT (SpliREPAT872) and then checked the status of the baculovirus infections. Silencing of SpliIIM3594 induced the mortality, while silencing of SpliCDA823 reduced the mortality. No effect on the baculoviral activity was detected upon silencing the SpliREPAT872; however, this silencing was transient. This study showed that baculovirus infections interfere with insect mucins and chitin deacetylases by possibly disruption of the PM. This study was supported by The Scientific and Technological Research Council of Turkey (Project #: 1120305).

Dissecting the host cell endocytic trafficking pathway STU 2:45 •163• of baculovirus by single-virus tracking and live-cell imaging

Fujun Qin^{1,2}, Congrui Xu^{1,2}, Shili Yang^{1,2}, Gaobo Zhang^{1,2}, Chengfeng Lei¹, Jia Hu¹, Hanzhong Wang¹, Xiulian Sun¹

¹Wuhan Institute of Virology, Chinese Academy of Sciences, Wuhan 430071, P.R. China; ²University of Chinese Academy of Sciences, Beijing 100049, P.R. China Corresponding author: sunxl@wh.iov.cn

The Baculoviruses are enveloped DNA viruses that have been widely applied for insect control, eukaryotic expression, and gene therapy. Based on the use of limited inhibitors of endocytosis, previous study showed that the budded viruses (BVs) form of Autographa californica multicapsid nucleopolyhedrovirus (AcMNPV) entered into insect cells mainly through clathrinmediated uptake. However, the mechanism and the dynamics related to BV particles entering and trafficking through the endocytic network in the host cells remain unclear. In this study, in order to investigate the cell entry pathway, several Sf9 cell lines expressing distinct endocytic markers and recombinant baculoviruses with labeling tags were constructed. Singleparticle virus tracking revealed that most of BVs (82%) randomly diffused on cell surface before being delivered to a preformed clathrin coated pit (CCP). After internalization into cytoplasm, BVs were primarily transported to Rab5positive early endosomes, in which the nucleocapsids of BV particles (78%) were released into cytoplasm. Subsequently, a minority of viruses (22%) progressed to mature into intermediate endosomes and late endosomes, in which membrane fusion was also detected. Moreover, actin facilitated the processes of scission of CCP from the plasma membrane, and propelled clathrin coated vesicular (CCV) away from the membrane, whereas microtubules are required for the transport of virus-bearing endocytic carrier vesicle (ECV) to the perinuclear region. In summary, this is the first study that investigates the host cell entry process of baculovirus BVs at the single particle level in live cells, and describes the mechanistic and kinetic insights of the infection of BV. These findings will contribute to a better understanding of the infection pathway of baculoviruses.

Comparative study of fast-killing versus slow-killing SfAV isolates

STU 3:00 •164•

Hiroki Ishii¹, Eiko Arai¹, Ikbal Agah Ince², Xiao-Wen Cheng³, Dennis Bideshi^{4,5}, Maki N. Inoue¹, Brian Federici⁴, Madoka Nakai¹

¹Institute of Agriculture, Tokyo University of Agriculture and Technology, Fuchu,
Tokyo, Japan; ²Department of Medical Microbiology School of Medicine,
Acibadem University, 34752 Atasehir, İstanbul, Turkey; ³Department of Microbiology,
32 Pearson Hall, Miami University, Oxford, Ohio 45056, USA;
⁴Department of Entomology, University of California, Riverside, Riverside,
California 92521, USA; ⁵California Baptist University, Riverside California 92405, USA
Corresponding author: madoka@cc.tuat.ac.jp

Ascoviruses (AVs) are double-stranded DNA viruses with circular genomes. They primarily attack lepidopteran larvae. AVs are transmitted by female parasitic wasps via oviposition and cause a chronic disease that terminates mortality. It is likely that a prolonged infectious period results in a higher rate of transmission by parasitoids that vector these viruses. However, we found that an atypical fast-killing AV (Spodoptera frugiperda ascovirus-X: SfAV-X) kills host rapidly after infection. The 50% survival time (ST_{50}) of SfAV-X infected 4th instar *Spodoptera litura* larvae was 2.6 dpi, while ST_{50} of the slow-killing AV (SfAV-F), which is an isolate closely related to SfAV-X, was 22.0 dpi. Our poster at SIP2016 showed that the genome replication speed of SfAV-X was significantly faster than that of SfAV-F. However, it showed SfAV-X infected larvae died as soon as genome replication plateaued (the interval was 0.9 dpi), while SfAV-F infected larvae died long-after SfAV-F genome replication plateaued (interval was 18.1 dpi). So the fast-killing phenotype results from genome replication speed and other factors. In this study, to identify other factors that cause the fast-killing phenotype, we compared tissue tropism and AV proteins between SfAV-X and SfAV-F. Firstly, we observed virions in tissues by Transmission Electron Microscopy (TEM). Because it is known that the tissue tropism of SfAV-1a, the slow-killing AV, is limited primarily to the fat body, we hypothesized that the fast-killing AV could infect other tissues resulting in a fast-killing infection. However, our observations showed SfAV-X only infected the fat body, so broader tissue tropism was not the cause of fast-killing. Secondly, we compared AV

hemolymph proteins of SfAV-X and SfAV-F after viral genome replication plateaued. This experiment identified 51 AV proteins in hemolymph of infected larvae, 7 of which were synthesized at a significantly higher rate in SfAV-X than in SfAV-F. Thus, our current hypothesis is that one or more of these 7 proteins are responsible for the faster killing speed of SfAV-X.

Microplitis bicoloratus bracovirus regulates NF-κB signaling pathway

3:15 •165•



Dong-Shuai Yu¹*, Ya-Bin Chen¹*, Ming Li¹*, Ming-Jun Yang², Yang Yang, Jian-Sheng Hu, Kai-Jun Luo

¹School of Life Sciences, Key Laboratory for Animal Genetic Diversity and Evolution of High Education in Yunnan Province, Yunnan University, Kunming, 650091, P.R. China; 2Shanghai-MOST Key Laboratory of Health and Disease Genomic, Chinese National Human Genome Center at Shanghai. Shanghai, P.R. China, 201203

Corresponding author: kaijun_luo@ynu.edu.cn

Polydnaviruses (PDVs) play a critical role in altering host gene expression to induce immunosuppression. However, PDV gene(s) to affect host genes remain largely unclear. Here, the whole-genome sequencing of Microplitis bicoloratus bracovirus (MbBV), which is known as an apoptosis inducer, was performed. The MbBV genome showed 17 putative double stranded DNA circles and 179 fragments with a total size of 336,336 bp encoding 116 open reading frames (ORFs). Based on conserved domains, 9 gene families were identified, in which IkB-like viral ankyrin (vank) family included 28 members and was one of the largest family. Among 116 ORFs, 13 MbBV genes were expressed in the MbBV-induced apoptosis hemocytes and further analyzed. Three vank genes (vank86, vank92, vank101) were expressed in hemocytes collected from Spodoptera litura larvae parasitized by M. bicoloratus, in which host NF-κB/IκBs including relish, dorsal, and cactus, were also persistently expressed. When Spli221 cells were infected with MbBV viral particles, mRNA levels of host and viral NF-κB/IκB genes were persistent and also varied in the virus-induced pre-apoptosis cell Spli221 from 1 to 5 hours post infection; then both presented a time-dependent expression manner in virus-induced apoptotic cells. Upon these analyses, we found that viral IKBlike transcription could not inhibit host NF-kB/lkB expression, suggesting that they might be able to regulate the different transcriptional mechanisms.

The Forum

Bacteria Division #4

Moderators: Ming Sun, Marianne Carey

Two mosquitocidal Clostridium bifermentans strains contain a Cry toxin and a Clostridium Neurotoxin-like loci 1:30 •166•

Estefania Contreras, Swati Chawla, Nadia Qureshi, Jianwu Chen, Sarjeet Gill

Department of Cell Biology and Neuroscience. University of California Riverside. California, USA Corresponding author: estefani@ucr.edu

Mosquito-borne diseases have been significantly impacted human civilization despite of centuries of intensive control effort. Diseases such as Dengue, Yellow fever, Chikungunya or Zika which are transmitted most commonly by an infected Aedes female mosquito, Filariasis and West Nile fever by Culex, and Malaria by Anopheles mosquito species, still remain serious public health problems with constant threats of re-emergence. Bacillus thuringiensis israelensis and Lysinibacillus sphaericus formulations have become the predominant non-chemical means for Culex and Aedes mosquito control. However, new biological insecticides with different mode of actions are needed in order to broaden susceptibility range and avoid mosquito resistance. In our work, we compared the toxicities of two Clostridium bifermentans mosquitocidal strains and Bacillus thuringiensis israelensis against Aedes aegypti, Anopheles gambiae and Anopheles stephensi. Clostridium bifermentans malaysia was the most toxic strain to Anopheles gambiae. In order to find its toxic components, we sequenced the genome of the two C. bifermentans mosquitocidal strains, a non mosquitocidal C. bifermentans wild type strain and a non toxic C. bifermentans malaysia mutant that we developed using gamma irradiation. Our sequencing data showed that C. bifermentans mosquitocidal strains contain a plasmid containing two toxin loci. The first one is composed by Cry16A, Cry17A and

two hemolysins which are toxic as a complex to Aedes aegypti, but not to Anopheles mosquitoes. Interestingly, the second toxin locus has similarity to Clostridium Neurotoxin loci, which we renamed Ctox locus.

Brevibacillus laterosporus insecticidal toxin genes and their expression during pathogenesis

1:45 •167•

Luca Ruiu¹

¹Dipartimento di Agraria, Sezione Entomologia, University of Sassari, Italy Corresponding author: lucaruiu@uniss.it

The endospore-former Brevibacillus laterosporus, morphologically featured by a typical canoe-shaped parasporal body, is an insect pathogen acting by ingestion through a toxin-mediated mechanism involving specific interactions with the host midgut epithelium. Genome sequencing of some B. laterosporus strains showing insecticidal activity against different targets, led to the identification of several putative toxins and virulence factors, including mosquitocidal toxins, vegetative insecticidal proteins, chitinases, and several polyketides and nonribosomal peptides. In addition, proteins of the spore coat canoe-shaped parasporal body complex (SC-CSPB) were shown to be involved in pathogenesis, thus representing additional virulence factors. Our recent genome sequencing and annotation of strain UNISS 18, a well documented pathogen of mosquitoes and flies, highlighted a variety of putative insecticidal toxin genes. Following a comparison with other B. laterosporus genomes, we conducted in vitro and in vivo gene expression and proteomic analyses that revealed which proteins may have a leading role in the pathogenic process. This study was financially supported by Italian Ministry of Education, University and Research (MIUR) (PRIN project 2015 BIOPIC, 2015BABFCF).

Identification of a genetic locus encoding a Serratia entomophila Penicillin-Binding Protein region associated to toxicity towards insect larvae

2:00 •1680

M. Eugenia Nuñez-Valdez¹, Victor Cruz-López¹ Mauricio Díaz-Sánchez¹, Zitlhally Rodríguez-Segura¹, Jeanwu Chen², Sarjeet Gill²

Centro de Investigación en Dinámica Celular Universidad Autónoma del Estado de Morelos, Cuernavaca, Morelos, Mexico and Depts. of Cell Biology and Neuroscience², University of California, Riverside, California, USA Corresponding author: eugenia@uaem.mx

The bacterium S. entomophila strain Mor4.1 (SeMor4.1; Enterobacteriaceae) is a entomopathogenic bacteria isolated in Mexico from larvae of the soil dwelling pest Phyllophaga spp (Coleoptera: Scarabaeidae). Insecticidal activity against larvae of Phyllophaga spp, Anomala sp and larvae of the lepidopteran Manduca sexta has been observed by injecting either, the bacteria or the cell free culture broths, suggesting that Mor4.1 secretes toxin-like factors with a wide spectrum of action at the level of the hemocoel. In order to identify SeMor4.1 toxic factors a SeMor4.1 genome library was prepared in Escherichia coli. Five insecticidal clones were selected by injection bio-assays. Further subcloning of the C8 clone render the clone E9 encoding an amino terminal region of a putative Penicillin-Binding Protein 1b (PBP) inserted in the pBluescriptSK+. Cell-free culture broth from E9 was evaluated (50µg protein/larva) by injection bio-assays in larvae of the Coleoptera Phyllophaga blanchardi and the Lepidoptera Spodoptera frugiperda and Galleria mellonella. Results showed that E9 caused 70%, 90% and 65% mortality after three days from the onset of bioassay in P. blanchardi, S. frugiperda and G. melonella larvae, respectively. No significant mortality was observed for the control larvae. No toxic activity was observed by oral inoculation. The characterization of SeMor4.1 toxins is important to determine their potential for the control of insect pests. Acknowledgments: We thank CONACYT (CB-2014-243027) for supporting this work.

A novel type six secretion system in the insect-pathogenic bacterium Xenorhabdus bovienii

2:15 •169•

 $\it Rebecca \, McQuade^1$, Christine Bradshaw 1 , S. Patricia $\it Stock^{1,2}$

¹Center for Insect Science, University of Arizona, Tucson, USA;
²Department of Entomology, University of Arizona, Tucson, USA

Corresponding author: rmcquade@email.arizona.edu

Xenorhabdus bovienii (XB) has a fascinating dual lifecycle- as a beneficial symbiont of nematodes and a potent pathogen of soil-dwelling insect larvae. Steinernema nematodes harbor XB in a specialized intestinal receptacle and vector XB into insects, releasing the bacteria into the insect body cavity. where they produce a variety of insecticidal toxins and antimicrobials. We recently identified a novel type six secretion system (T6SS) in the genome of XB. Because T6SSs allow bacteria to directly manipulate eukaryotic host cells or other bacteria, we speculate that the novel T6SS contributes to XB's interactions with its invertebrate partners or the normal flora of host insects. Using gRT-PCR, we found that the T6SS is highly expressed in XB in a medium mimicking insect hemolymph. To identify potential T6-secreted proteins, we generated a mutant XB strain lacking Hcp, a key structural component of the T6SS, and compared its secretion profile to that of wild type XB. Using 1D gel electrophoresis coupled with mass spectrometry, we identified several proteins in the wild type supernatant that are absent in the Δhcp supernatant, including one with domains characteristic of T6-secreted effectors. We are currently exploring the functions of these proteins and the contribution of the novel T6SS to the complex XB lifecycle.

Influence of exogenous pathogens on the expression of *BmToll9-2* gene in larval *Bombyx mori*

•170•

Jisheng Liu^{1*}, Wenli Liao¹, Mingli Zhao¹, Guy Smagghe², Luc Swevers³

¹School of Life Sciences, Guangzhou University, Guangzhou, China;

²Department of Crop Protection, Faculty of Bioscience Engineering,
Ghent University, Ghent, Belgium; ³Insect Molecular Genetics and Biotechnology,
Institute of Biosciences & Applications, National Centre for Scientific

Research Demokritos, Athens, Greece

Corresponding author: iishena.liu@azhu.edu.cn

Aim: Toll-like receptors are important members in Toll signaling pathway in insects and play an important role in activating the innate immune response to invading pathogenic microorganisms. This study aims to explore the influence of exogenous pathogens on the expression of the Toll-like receptor gene BmToll9-2 in larvae of the silkworm, Bombyx mori. Methods: Exogenous pathogens lipopolysaccharide (LPS) and Escherichia coli were respectively injected into the body cavity of the 5th instar larvae to induce their immune response, and the expression levels of BmToll9-2 in the midgut, fat body and epidermis at different time after injection were detected by real-time quantitative PCR. Results: The real-time quantitative PCR results showed that injection of LPS could induce the expression of BmToll9-2 in the midgut of the 5th instar larvae, and the best induction effect was at 6 h after injection. Injection of E. coli could also induce the expression of BmToll9-2 in the midgut of the 5th instar larvae, and the best induction effect was at 3 and 6 h after injection. Injection of LPS and E.coli could also induce the expression of BmToll9-2 in the fat body and epidermis, and the best induction effect was at 24 h after injection. Conclusion: Expression of BmToll9-2 in B. mori larvae is up-regulated after injection of LPS and E. coli, suggesting that BmToll9-2 receptor may be involved in the recognition process of Toll-like receptors to

Symbionts in waiting: the dynamics of incipient endosymbiont complementation and replacement in minimal bacterial communities of psyllids

exogenous pathogens such as LPS and E. coli in insects.

2:45

Jennifer L. Morrow, Aidan A.G. Hall, Markus Riegler

Hawkesbury Institute for the Environment, Western Sydney
University, Locked Bag 1797, Penrith, NSW 2751, Australia
Corresponding author: m.riegler@westernsydney.edu.au

Obligate bacterial primary (P-) endosymbionts that are maternally inherited and codiverge with hosts are widespread across insect lineages with nutritionally restricted diets. Secondary (S-) endosymbionts are mostly facultative but in some hosts they complement P-endosymbiont function and

therefore become obligate. Phylogenetic evidence exists for host switching and replacement of S-endosymbionts. The community dynamics that precede endosymbiont replacement and complementation have been little studied across host species, yet they are fundamental to the evolution of endosymbiosis. We performed bacterial 16S rRNA gene amplicon sequencing of 25 psyllid species (Hemiptera, Psylloidea) across different developmental stages and ecological niches by focusing on the characterisation of the bacteria other than the universally present P-endosymbiont Carsonella (Gammaproteobacteria). Most species harboured only one dominant representative of diverse gammaproteo-bacterial S-endosymbionts that was consistently detected across all host individuals and populations (Arsenophonus in eight species, Sodalis or Sodalis-like bacteria in four species, unclassified Enterobacteriaceae in eight species). The identity of this dominant obligate S-endosymbiont varied across closely related host species. Unexpectedly, five psyllid species had two or three co-occurring endosymbiont species other than Carsonella within all host individuals, including a Rickettsiella-like bacterium (Gammaproteobacteria) in one psyllid species. Based on standard and quantitative PCR, all psyllids carried Carsonella, at higher titres than their dominant S-endosymbionts. Some psyllids also had Alphaproteobacteria (Lariskella, Rickettsia, Wolbachia) at varying prevalence. Incidence of other bacteria, including known plant pathogens and potential entomopathogens was low. Ecological niche of gallforming, lerp-forming and free-living psyllid species did not impact endosymbiont communities. Two flush-feeding psyllid species had population-specific differences, and this was attributable to the higher endosymbiont diversity in native ranges and the absence of some endosymbionts in invasive ranges. Our data support the hypothesis of strict vertical transmission of minimal core communities of bacteria in psyllids. We also found evidence for S-endosymbiont replacement across closely related psyllid species. Multiple dominant S-endosymbionts present in some host species, including at low titre, constitute potential examples of incipient endosymbiont complementation or replacement. Our multiple comparisons of deep-sequenced minimal insect bacterial communities exposed the dynamics involved in shaping insect endosymbiosis.

Invertebrate pathology in the absence of infection: The role of gut bacteria in mosquito development



Michael R. Strand, Kerri L. Coon, Luca Valzania, David A. McKinney, Kevin J. Vogel, Mark R. Strand

Department of Entomology, University of Georgia, Athens, GA, USA
Corresponding author: mrstrand@uga.edu

Mosquitoes host communities of microbes in their digestive tract that consist primarily of bacteria. Several mosquito species including *Aedes aegypti* do not develop beyond the first instar in the absence of a gut microbiota. In contrast, several species of bacteria, including *Escherichia coli*, rescue development of axenic larvae into adults. The molecular mechanisms underlying bacteria-dependent growth in contrast are unknown. Here we designed a genetic screen around *E. coli* that identified high affinity cytochrome *bd* oxidase as an essential bacterial gene product for mosquito development. Bioassays showed that bacteria in non-sterile larvae and gnotobiotic larvae inoculated with wild-type *E. coli* reduced midgut oxygen levels below 5%, whereas larvae inoculated with *E. coli* mutants defective for cytochrome *bd* oxidase did not. Experiments further supported that hypoxia stimulates growth and ecdysone-induced molting. Altogether, our results identify aerobic respiration by bacteria as a previously unknown but essential process for mosquito development.

Roosevelt Room

Fungi Division #3

Moderators: Jae Su Kim, Richard Humber

Developing a strain improvement system for the entomopathogenic fungus Beauveria bassiana: a way to get better biocontrol agents?

STU 1:30 •174•

Laura Reyes, Dave Chandler

School of Life Sciences, University of Warwick, Wellesbourne Campus, Warwick, CV35 9EF, UK Corresponding Author: L.E.Reyes-Haro@warwick.ac.uk

Biocontrol agents (BCAs) based on entomopathogenic fungi (EPF) are playing an increasing in Integrated Pest Management programmes. At present, the commercially available EPF all consist of wild type strains isolated from nature, however, there is potential to breed more effective strains by recombining wild types with complementary characteristics. The aim of this research is to develop a system to improve strains of the entomopathogenic fungus Beauveria bassiana through genetic recombination. A group of 50 Beauveria strains were genotyped using multi-locus sequencing and mating gene analysis, and then phenotyped with respect to their virulence, thermal biology, tolerance of UV light, and spore production. Five strains were selected with complementary phenotypes and potentially compatible mating types for parasexual recombination studies. Spontaneously generated nitrate non-utilizing (nit) mutants have been produced from these strains using a potassium chlorate-amended selective medium and their vegetative compatibility is currently being investigated prior to parasexual recombination studies. Eight fungal strains have also been selected to investigate the potential for inducing sexual recombination by pairing complementary mating types on different media (oatmeal agar, malt extract agar, and Czapek Dox agar +/- biotin) known to induce ascospore formation in other anamorphic ascomycetes. Work is currently underway to develop parasexual and sexual recombination systems for selected strains with complementary phenotypes. The long-term aim is to produce recombinant strains that are more efficient as biocontrol agents.

Regulation of gene expression in Beauveria bassiana strain infecting Riptortus pedestris, bean bug



Se Jin Lee, Sihyeon Kim, Mi Rong Lee, Jae Su Kim*

Department of Agricultural Biology, College of Agricultural & Life Sciences, Chonbuk National University, Jeonju 561-756, Korea Corresponding author: iskim10@ibnu.ac.kr

Beauveria bassiana (Bb), a major species of entomopathogenic fungi, has been known as highly virulent against a variety of insect pests. So far, many studies have focused on its insecticidal activity and virulence, but little consideration was given to the molecular mechanisms to figure out how it works in pathogenesis. Herein, to understand the transcriptional interaction between Bb and bean bug (Riptortus pedestris), the genomic DNA isolated from a Bb strain JEF-007 was subjected to whole genome sequencing using Pacbio sequencing, and transcriptomes of the infecting fungal strain and fungi-infected bean bug were analyzed. The genomic DNA of Bb JEF-007 was 36.5 Mb long and predicted to have 39 scaffolds and 10,857 genes. A total contigs of non-infected/infected bean bug and non-infecting/infecting fungal strain were 141,722 and 37,503, respectively. Differential expressed gene (DEG) analysis showed that 2,302 and 2,381 genes were up- and downregulated respectively in the infecting Bb JEF-007 and virulence-related genes such as halogenase, cytoxic lectin, bacterial-like toxin and proteins related to cell wall formation were validated to be highly up-regulated. DEG analysis of the infected bean bugs showed that 3.588 and 3.296 genes were up- and down-regulated, respectively, and cuticle protein, cysteine-rich secreted protein and pigment-dispersing factor precursor were highly upregulated possibly to survive against the fungal infection. This work can provide a deep insight on molecular mechanisms of infecting Bb and how the insect responds to the fungal infection in transcriptional level.

Alkane-grown Beauveria bassiana: Mycelial pellets formation, oxidative stress induction and cell surface alterations

STU 2:00 01760

Carla Huarte-Bonnet, Juan C. Ponce,

Marianela Santana, Eduardo Prieto, Nicolás Pedrini

Instituto de Investigaciones Bioquímicas de La Plata (CCT La Plata CONICET-UNLP), Facultad de Ciencias Médicas.

La Plata, Argentina

Corresponding author: carlahb@hotmail.es

Entomopathogenic fungi attack insect hosts via attachment to the epicuticle, usually composed by aliphatic hydrocarbons. B. bassiana is able to grow on insect-like alkanes improving its virulence. In this study we described some physiological and molecular processes involved in fungal growth, nutritional stress response and cell surface alterations found in alkane-grown conidia. Fungi were observed by transmission electron microscopy (TEM) and atomic force microscopy (AFM), and their hydrophobicity was measured on the cell surface. Additionally, the expression pattern of several genes associated with oxidative stress, hydrophobicity and peroxisomal biogenesis was analyzed by qPCR. We found a novel type of growth in alkane-cultured B. bassiana, similar to mycelial pellets described in other filamentous fungi grown in alkane-free media. Pellets were able to germinate in media without a carbon source and produce viable conidia. TEM showed that pellets were formed by hyphae cumulates with an apparent surface thickening. Cell surface appeared to be more hydrophobic and exhibited different surface topographies as was observed by AFM. We also found a significant induction in several genes encoding for catalases, superoxide dismutases, hydrophobins and peroxins. Additional studies are being conducted to better understand the relationship between alkane growth adaptation and fungal cell changes, in order to improve the efficacy of fungal penetration through the cuticle and thus enhance virulence against insect pests.

Entomopathogenic fungal granules for biological control of Protaetia brevitarsis seulensis larvae



Sihyeon Kim, Se Jin Lee, Jong Cheol Kim, Mi Rong Lee, Jae Su Kim

Department of Agricultural Biology, College of Agricultural & Life Sciences, Chonbuk National University, Jeonju 54896, Republic of Korea Corresponding author: jskim10@jbnu.ac.kr (J. S. Kim)

Many turfgrass pests, such as Ectinohoplia rufipes, Exomala orientalis and Popillia auadriauttata, cause serious damage to grass fields in Korea, Given the residual side-effects of synthetic pesticides in turfgrass soil, more safe control agents, for example entomopathogenic fungi, need to be developed. In this study, Protaetia brevitarsis seulensis as an alternative insect was used to screen out highly virulent entomopathogenic fungi. In a virulence assay where the insect larvae were exposed to the fungal mass of nine genus of entomopathogenic fungi on cultured dishes, Beauveria, Metacordyceps and Metarhizium isolates showed high virulence against P. brevitarsis seulensis. One Metathizium isolate showed ca. 60% virulence 7 days after the exposure. Compared to the contact-exposure, the spray treatment of Metathizium isolate showed relatively lower virulence against the larvae. In addition, the selected isolate showed insect stage-dependent virulence in laboratory conditions; high virulence against eggs and 1st larvae but low virulence against 2nd and 3rd larvae. A semi-filed assay was conducted to investigate the potential of soil application of the selected isolate. These results provide a basic information for the control of P. brevitarsis seulensis using entomopathogenic fungi.

Metarhizium brunneum (Ascomycota: Hypocreales) treatments are safe for the generalist predator Chrysoperla carnea (Stephens) (Neuroptera: Chrysopidae) 2:30 •179

Inmaculada Garrido-Jurado¹, Álex Ríos-Moreno¹, Enrique Quesada-Moraga¹ ¹Department of Agricultural and Forestry Sciences, ETSIAM, University of Cordoba, C4Building, Campus of Rabanales, 14071 Cordoba, Spain. Corresponding author: g72gajui@uco.es

Metarhizium sp. has shown great success in the control of insect pests as a biocontrol alternative to hazardous chemical pesticides. This genus can secrete secondary metabolites such as destruxin A, which information on their fate in the food chain and their risk to human and animal health is

scarce. In the present work, predator-prey bioassays were performed to evaluate the behavior and survival of Chrysoperla carnea (Stephens) larvae when feeding on larvae of the polyphagous pest Spodoptera littoralis challenged by M. brunneum BIPESCO5 and EAMa 01/58-Su strains. In addition ecotoxicological studies based on HPLC-MS were performed to monitor the fate of destruxin A in the prey-predator system. The maximum concentration of destruxin A produced by BIPESCO5 and EAMa 01/58-Su strains in infected S. littoralis larvae was approximately 0.014 and 0.031 µg/g, respectively, whereas the metabolite was not detected in C. carnea larvae consuming M. brunneum-challenged S. littoralis larvae. Furthermore, C. carnea larvae preferred consuming healthy prey versus M. brunneumchallenged prey as revealed by both the higher predator ratio feeding on S. littoralis control larvae and the higher per capita number of prey control larvae consumed by the predator compared to the M. brunneum-challenged larvae. Hence, the M. brunneum treatments against S. littoralis larvae were safe for C. carnea due to both the lack of fungus-related mortality in the predator and the lack of movement of destruxin A from the prey to the predator.

Ecological biocontrol as a strategy to develop fungal biopesticides for successful pest management (e-Biopesticide)

Jae Su Kim¹, Se Jin Lee¹, Jong Cheol Kim¹, Sihyeon Kim¹, Mi Rong Lee¹, So Eun Park¹, Seok Ju Lee¹, Taek Su Shin², Tae Hoon Kim², Pan Jung Ha², Tae Hyun Park²

2:45 •180•

¹Department of Agricultural Biology, College of Agricultural & Life Sciences
Chonbuk National University, Jeonju 561-756, Korea; ²Crop Protection
R&D Center, Farm Hannong (LG Chemical Affiliated Co.), Nonsan 39-23, Korea
Corresponding author: jskim10@jbnu.ac.kr

Now we are trying to deal with problems in pest management, such as ecological toxicity of synthetic pesticides, increasing pest resistance and stronger pesticide regulation in many countries. As one of the solutions, many global companies have been merging and acquiring some biopesticide companies, Pasteuria, Agraquest, Prohphya, Itaforte, and Becker Underwood. Some collaborative works have been producing biopesticides with enhanced quality. In the R&D of microbial insecticides, bacterial and viral insecticides received many interests and outstanding commercial products have been developed. However little consideration has been given to the fungal insecticides, although this group has much higher potential in controlling hemipteran and thysanopteran insects by the hyphal penetration through the integument. High production cost of fungal insecticides might be a technical barrier. Application of the fungal insecticides to unfavorable conditions might result in lower performance in pest management. Herein this work, I suggest ecological biocontrol considering long-term colonization in nature rather than quick pest control. Additionally other important technical aspects need to be strongly considered, such as economic downstream process, effective control and environmentally safe, so finally proposing 4e-biopesticide. In terms of ecological biocontrol, entomopathogenic fungal pesticides can be applied to the soil and water in agricultural fields, water in urban cities and hydric areas in forest, where the use of synthetic pesticides is not easily accepted and getting disapproved before long. We have some achievements in the management of thrips and other pests. With this field-oriented works, our group is also working on fungal molecular biology to elucidate the mode of actions and interaction between insects and entomopathogenic fungi. This ecological biocontrol can be a strong background for successful development of fungal biopesticides.

Isolation, morphological and molecular characterization of entomopathogenic fungi as a potential control strategy against Lobesia botrana in Argentinean vine growing areas

Rodrigo J. López Plantey^{1,3,4}, Daciana Papura², Andrés E. Riquelme³,
Antonella Balloni³, Pablo H. Pizzuolo^{1,4}, Joana J. Boiteux^{1,4},
Denis Thierry², Gabriela S. Lucero^{1,4}

¹ Instituto de Biología Agrícola de Mendoza (CCT Mendoza CONICET-UNCuyo), Facultad de Agronomía, Mendoza, Argentina; ²UMR-SAVE, INRA/Bordeaux Sciences Agro, Villenave-d'Ornon, France; ³Cátedra de Zoología Agrícola, Facultad de Ciencas Agrarias, UNCuyo, Mendoza, Argentina; ⁴Cátedra de Fitopatología, Facultad de Ciencias Agrarias, UNCuyo, Medoza, Argentina

**Corresponding author: rlopezplantey@mendoza-conicet.gob.ar

The appearance of new pests in a culture requires actions to prevent establish-ment, spreading internal or across international borders. The European grapevine moth Lobesia botrana was recently introduced in Argentina in 2010, when it was declared as quarantine pest. The current methods for controlling grape moth's populations include conventional chemical ovicides or larvicides, but also alter-native methods as mating disruption and Bt toxin. The use of entomopathogenic fungi as a control strategy of Lepidoptera is known from ancient times. Argen-tinean wine production is mainly concentrated in the western area of the country, next to the Cordillera de los Andes with a high specificity in the local environ-mental conditions. The aim of present work was to isolate and characterize natives entomopathogenic fungi from Argentinean grapevine environment, which could be propose as biological control agents against L. botrana. Soils from vineyards over 15 years old were sampled across 1200km on the Argentinean provinces of Mendoza, San Juan, Catamarca, La Rioja, Salta, Río Negro and Neuquén. A total of 45 samples were prepared and conserved at laboratory. Petri dishes with prepared soils samples were used according to the insect bait method (3 repetition per sample), where 15 larvae L4/L5 of L. botrana were placed for 7 days at controlled temperature (25°C), photoperiod (16:8) and humidity (>60%). Larvae with fungus symptoms were separated and cultivated until fungus isolation and purification. A total of 32 strains of entomopathogenic fungi were isolated which were characterized by dichotomous key and molecular techniques using a partial sequence of nuclear protein-encoding gene, translation elongation factor 1-alpha. Three genders were identified by morphological characterization and all the strains were placed by BLAST® in one of these three species: Beauveria bassiana, Metarhizium robertsii and Paecilomyces lilacinus. The results help to start the physiological and pathogenic studies of these strains against L. botrana and their use as biological control.

3:30-4:00 pm Refreshments Break

WEDNESDAY, 4:00 – 6:00 pm POSTERS

West Ballroom

All presenting poster authors should be present at or near their posters.

All posters must be removed by 6:30 pm on Wednesday

VIRUS DIVISION POSTERS

Genomics of alphabaculovirus isolates infecting *Malacosoma disstria* in North America

Vir01 •181•

Martin A. Erlandson¹, Doug Baldwin¹, Andrew Keddie²,
Georae Rohrmann³. David A. Theilmann⁴

¹Saskatoon Research and Development Centre, Agriculture and Agri-Food Canada, Saskatoon, SK, Canada; ²Department of Biological, Sciences, University of Alberta, Edmonton, Alberta, Canada; ³Department of Microbiology, Oregon State University, Corvallis, OR, USA; ⁴Summerland Research and Development Centre, Agriculture and Agri-Food Canada, Summerland, BC, Canada

 ${\it Corresponding\ author:\ martin.erlandson@agr.gc.ca}$

The forest tent caterpillar (FTC), Malacosoma disstria, is a major defoliator of deciduous trees in Canada and the USA and is characterized by cyclic population outbreaks. The alphabaculovirus species infecting FTC, Malacosoma disstria nucleopolyhedrovirus (MadiNPV), is one of the major natural regulators of FTC population cycles. We have sequenced the

Vir02 •182•

Vir03 •183•

complete genomes of two strains, pp3A and pp11A, from the MadiNPV-A92 isolate. The MadiNPV-pp3A and -11A genomes are closely related with an average predicted ORF identity of 99% and have 41% G+C content, and are 135,623 and 136,414 bp, respectively, and contain 135 and 139 ORFs, respectively. Phylogenetic analysis of polyhedrin, lef-8 and lef-9 sequences, generated using degenerative primers, of NPVs isolated from FTC, as well as sequences from the NCBI database of MaamNPV (M. americanum - eastern tent caterpillar, North America), MacaNPV (M. californicum - western tent caterpillar, North America), and ManeNPV (M. neustria - lackey moth, Eurasia) indicated that the MadiNPV isolates constitute a distinct alphabaculovirus species that is most closely related to ManeNPV. Our previous studies indicated that budded virus of MadiNPV pp3A and pp11A isolates had differential replication dynamics in three cell lines from FTC. However, the only major differences in the gene content of the MadiNPV isolates was the presence of 3 additional ORFs of unknown function in pp11A and a large 910 aa ORF in pp3A of unknown function that is represented by 2 truncated ORFs in pp11A. Whether these ORFs play a role in the differences in virus replication dynamics noted in the various host cell lines will require further study.

Oddities in the genome of the alphabaculovirus infecting Malacosoma californicum pluviale

Elisabeth A. Herniou¹, Julien Thézé^{1,2}, Aurélien Chateigner¹, Shannon Escasa³, Martin Erlandson⁴, David Thielmann⁵, Jennifer S. Cory^{3,6}

¹ Institut de Recherche sur la Biologie de l'Insecte, UMR CNRS 7261, Université de Tours, Faculté des Sciences et Techniques, Tours, France; ² Department of Zoology, University of Oxford, Oxford OX1 3SY, UK; ³ Laboratory for Molecular Virology, Great Lakes Forestry Centre, NRC, Canada: 4 Saskatoon Research and Development Centre, Agriculture and Agri-Food Canada, Saskatoon, SK, Canada; 5 Summerland Research and Development Centre, Agriculture and Agri-Food Canada, Summerland, BC, Canada; ⁶ Department of Biological Sciences, Simon Fraser University, Burnaby, Canada Corresponding author: elisabeth.herniou@univ-tours.fr

The Western tent caterpillar Malacosoma californicum pluviale is well known for its cyclic population dynamics associated with its specific alphabaculovirus McpINPV. Over the years considerable effort has been put into determining the complete genome of McpINPV, but so far assembly has remained unsatisfactory. Here, based on combined 454 and Illumina data, we were able to assemble a circular consensus sequence of 135,827 bp and comprising 144 ORFs. One of the striking features of this genome is the disruption of the cluster of 4 genes that is conserved in all other baculovirus genomes completely sequenced to date. A second characteristic of McpINPV concerns its homologous repeat regions. The genome of McpINPV contains 12 HRs formed of large palindromic repeats and ranging in size from 233 to 928 bp. Whole genome phylogenetic analyses showed McpINPV belong to the group II of the genus Alphabaculovirus but is distant from other baculovirus genomes.

Genomic comparisons among Phthorimeae operculella granuloviruses isolated from different hosts in field

Gloria P. Barrera¹, Juliana Gómez¹, Mariano N. Belaich², Pablo D. Ghiringhelli², Carlos Espinel¹, Laura Villamizar³

¹Corporación Colombiana de Investigación Agropecuaria, Corpoica, Bogotá, Colombia: ²Laboratorio de Ingeniería Genética y Biología celular y Molecular. Universidad Nacional de Quilmes, Provincia de Buenos Aires, Argentina; AgResearch Ltd. Lincoln Research Centre. Christchurch 8140, New Zealand Corresponding author: gbarrera@corpoica.org.co

Some baculoviruses infect different hosts as a mechanism to increase their genetic diversity and to improve their fitness and maintenance in nature. However, most species from Betabaculovirus genus are characterized by showing a reduced host range. In the case of Phthorimaeae operculella granulovirus (PhopGV), previous works showed that infects different Gelechiidae's insects including Tuta absoluta (Meyrick) and Tecia solanivora (Povolny) with different biological activity. With the goal to contribute in the understanding of genetic causes associated with the infection of PhopGV in different hosts, two field isolates recovered from T. absoluta (TuabGV) and T. solanivora (TesoGV) were studied. To this, both genomes were completely sequenced and compared between them and with respect to other PhopGV

previously reported. Thus, a high degree of conservation in sequence and gene organization was observed being the main differences small insertions and deletions located in some open reading frames (ORF) and other noncoding regions. Only four ORFs showed variability among all genomes, corresponding to PhopGV 027 (envelope fusion protein), 041 (metalloproteinase), 046 and 092 (unknown function) ORFs. On the other hand, 24 ORFs were identical between TesoGV and TuabGV but they showed differences with respect to the PhopGV orthologs, while only five ORFs showed differences between TesoGV and TuabGV. When controlled bioassays of PhopGV, TuabGV and TesoGV were carried out, all isolates showed the highest performance against their original hosts, however TesoGV reduce the insecticidal activity against alternative host while PhopGV maintain high fitness in all. The proper biological interpretation of these variations may assist in understanding how baculoviruses support mutations that help them to increase their chances of success in nature. In this sense, they could be associated to the etiological cause of some phenotypical changes observed, thus transforming pangenomic studies into useful tools to nurture the field of functional genomics in Baculoviridae.

A new Group II alphabaculovirus isolated from Spodoptera ornithogalli



Gloria P. Barrera¹, Mariano N. Belaich², Pablo D. Ghiringhelli², Judith Guevara¹, Laura Villamizar¹,

¹Corporación Colombiana de Investigación Agropecuaria, Corpoica, Bogotá, Colombia. ²Laboratorio de Ingeniería Genética y Biología celular y Molecular, Universidad Nacional de Quilmes, Provincia de Buenos Aires, Argentina. ³AgResearch Ltd. Lincoln Research Centre. Christchurch 8140, New Zealand Corresponding author: gbarrera@corpoica.org.co

The yellow striped armyworm Spodoptera ornithogalli (Guenée) (Lepidoptera: Noctuidae) is a polyphytophagous insect widely distributed in America since Canada to Argentina. The larvae feed on various plants of agriculture importance such as alfalfa, tomato, tobacco and onion, among others. Particularly, S. ornithogalli can be a serious pest on transgenic cotton and ornamental flower crops in Colombia. In order to explore bioinsecticide agents to control populations of this insect, one field baculovirus isolate from Spodoptera ornithogalli larvae was characterized. Electron micrographs revealed the presence of Occlusion Bodies (OBs) with a mean diameter of 1.51 µm containing several rod-shaped nucleocapsids per virion. The virus genome was isolated and sequenced using Illumina technology. The phylogenetic tree based on translated proteins showed a closer relationships with alphabaculoviruses from insects belonging to Noctuidae family, S. litura, S. exigua and S. frugiperda. Based on the criterion for baculovirus species demarcation, nucleotide distances of concatenated polh, lef-8 and lef-9 genes were calculated by Kimura 2-parameter model. The nearest distance with respect to the other species [corresponding to Spodoptera litura nucleopolyhedrovirus (SpliNPV-II; GenBank NC_011616)] was greater than 0.05 indicating that SporMNPV is a new species belonging to the Group II of Alphabaculovirus genus of Baculoviridae family, thus enriching the available knowledge about Spodoptera spp. baculoviruses occurring in nature.

Characterization of baculovirus pathogenic to cassava hornworm (ErelGV) from Cruzeiro do Sul, Acre, Brazil

Vir05 •185•



William Sihler¹, **Márcio Martinello Sanches**¹, Rosana Falcão¹, Murilo Fanzolin², Joelma Lima Vidal Estrela², Marlinda Lobo de Souza¹

> ¹Embrapa Recursos Genéticos e Biotecnologia, Brasília, Brasil; ²Embrapa Acre.Rio Branco, Brasil Corresponding author: marlinda.souza@embrapa.br

Baculoviruses are rod-shaped enveloped viruses with circular doublestranded DNA, with the majority of species infecting insects of the order Lepidoptera. Due to its potential as pest control agents in agriculture and forests, they have been successfully used as bioinsecticides. The hornworm (Erinnyis ello) is an important pest of cassava and rubber tree with high capacity of migration. In this work it is presented the characterization of a baculovirus with occurrence in Erinnvis ello populations in area of small cassava farms in Cruzeiro do Sul, State of Acre, Brazil (Latitude -7,55086; Longitude -72,72570). The viral particles were purified from larvae with symptoms of infection through ultracentrifugation in sucrose gradient. After

treatment with uranyl acetate 2%, the particles were visualized using a JEOL 1011 transmission electron microscope. The virus with ovicylindirical occlusion body, which presented only one virion per envelope and size less than 0.5 µm, was identified as member of genus Betabaculovirus (formerly Granulovirus). The viral DNA was extracted through phenol/chloroform cycles, digested with different restriction enzymes and submitted to agarose gel electrophoresis containing ethidium bromide. The analysis of restriction patterns obtained with the enzymes Bam HI and Hind III revealed a total of seven and three DNA fragments, respectively. As a result of the cleavage with Pst I, eight molar fragments and three submolar fragments were identified. Finally, after digestion with Eco RI 21 fragments were observed, with some submolar bands being also present. The comparison of the DNA restriction profile from ErelGV isolated in Acre, North Brazil, with the pattern described for ErelGV collected in Santa Catarina State, South Brazil, showed a very high similarity between them.

First report of iflavirus infection in Spodoptera littoralis (Lepidoptera:Noctuidae)

Vir06 •186•

Umut Toprak

Ankara University, Molecular Entomology Lab., Faculty of Agriculture, Dept. of Plant Protection, Ankara, Turkey ${\it Corresponding\ author: utoprak@agri.ankara.edu.tr}$

Iflaviruses are single-stranded RNA viruses with a cytoplasmic viral replication from the family of Iflaviridae (order Picornavirales) that primarily infect insects. Most iflaviruses cause sublethal infections and are difficult to be detected by symptomatic observations; while some could be lethal to the honeybee and silkworm hosts. Iflaviruses have been detected from Halvomorpha halvs, Heliconius erato, Lymantria dispar, Spodoptera exigua. Bombyx mori and Apis mellifera; however, little is known on this family of viruses. In the current study, an iflavirus was identified through the occurrence of expression sequence tags (ESTs) from the larval midgut of Spodoptera littoralis (Lepidoptera:Noctuidae) and was denoted Spodoptera littoralis iflavirus 1 (SpliIV1). The identification was based on a cDNA encoding the polyprotein. BLAST analysis revealed an identity value of 97% with the polyprotein of Spodoptera exigua iflavirus 2. The SpliIV1 polyprotein cDNA is 4883 b in length, encoding a polypeptide protein of 1522 aminoacid with a predicted molecular weight of 171 kDa. The Spodoptera littoralis iflavirus 1 is the first iflavirus described in S. littoralis. However, more studies are required to understand the pathology and activity of the SpliIV1 in insect hosts.

A new virus infected mud vrab revealed by Cryo-EM

Vir07 •187 •

Qinfen Zhang, Yuanzhu Gao, Qianqian Wang, Jiamiao Huang, Shaoping Weng, Jianguo He

State Key Lab for Biocontrol, School of Life Sciences. Sun Yat-sen University, Guangzhou, China Corresponding author: Lsszaf@mail.svsu.edu.cn

The mud crab (Scylla serrata), an economically important species, is cultivated commercially in many countries, including China, India and Australia. However, viral diseases have had serious negative economic effects on the mud crab, limiting its growth. Previously, the mud crab reovirus (MCRV) and mud crab dicistrovirus (MCDV) have been isolated from mud crabs that had obvious symptoms of sleeping disease (SD). Cryo-Electron Microscopy (Cryo-EM) single particle techniques have been applied to study the high-resolution structures of these viruses. Better than 4 angstrom three dimensional (3-D) structure of the MCDV and MCRV were obtained. Very surprising, we found a new virus in the two-dimensional (2-D) classifications in MCDV sample, although the size and shape in 2-D are very similar to that of MCDV. And therefore it is very difficult to found there are two species virus through the density gradient centrifugation and conventional electron microscopy techniques. Further studies on both 3-D structure, genome and protein analysis reveal that the new virus should be a new member of the Nodaviridae. We named the new virus as the mud crab nodavirus (MCNV). The structure of the MCNV shows a RNA-containing capsid shell with an inner diameter of approximately 227 Å and an outer diameter of about 300 Å. The most prominent structural feature is a total of 30 protrusions located on the icosahedral 2-fold axes, which is usually on 3-fold axes in other

Nodavirudae members. These protrusions are approximately 36 Å tall and dimeric with two linking hinges. In contrast to the well-resolved capsid shell with icosahedral symmetry, the protrusions reveal highly flexible conformation as evidenced by the relatively weaker densities. This significant flexibility may be resulting from the long hinges connecting protrusions and capsid shell. The protrusions may play important roles in host reorganization.

Characterization of a persistent baculovirus infection established in an insect cell line





Raquel Arinto-Garcia¹, Sarah Irons¹, Louise Hughes¹, Chris Hawes¹, Linda King¹, Robert Possee²

Dept. Biological & Medical Sciences, Oxford Brookes University, Oxford UK; ²Oxford Expression Technologies Ltd, Oxford, UK Corresponding author: Raquel.arinto.garcia-2015@brookes.ac.uk

Baculoviruses such as Autographa californica nucleopolyhedrovirus (AcMNPV) are double-stranded DNA viruses that exclusively infect insects, leading to death of the larval host. Nevertheless, many insects harbour persistent, non-lethal virus infections throughout their life stages. Recently, a clonal culture of a persistent infection was achieved when infecting Trichoplusia ni cells (Hi5®) with a AcMNPV p10 deletion mutant. This cell line (C20) has been maintained continuously for 6 years and constitutes an important model to study persistent virus infection in vitro. Furthermore, the C20 cell line also provides a mechanism to study the superinfection exclusion phenomenon, as it is largely refractory to infection with a second virus. This study aims to understand the balance between the host insect cell and the virus, which allows the persistent infection to be maintained. In addition, we intend to investigate the barrier(s) preventing baculovirus superinfection. Characterization of the C20 cell line revealed that the cells retain viability for longer than the parental cell line, being able to survive for 40 days before sub culture. The persistent infection is maintained at a very low level although the infectious virus titre fluctuates considerably within and between different cultures. Using a recombinant virus with eGFP fused to VP39, the baculovirus major capsid protein, confocal laser scanning microscopy was used to follow virus uptake and subsequent infection to detect the super infection blockage. This study, combined with scanning electron microscopy on early infection time points, suggested the secondary infection of C20 cells is blocked on the adsorption or uptake step. Further studies are examining this question using recombinant viruses with eGFP expression regulated by early and late promoters. Understanding the bio-logical relationships responsible for the maintenance of C20 cell line may help us, in the longer term, to understand the finely constructed relation-ship between virus and host that leads to either lytic or persistent infection.

Autographa californica multiple nucleopolyhedrovirus ME53 interacts with VP80, LEF5, EXON0, VP39 and GP64



Emine Özşahin¹, Éva Nagy², **Peter J. Krell**¹

¹Department of Molecular and Cellular Biology and ²Department of Pathobiology, University of Guelph, Ontario, Canada Corresponding author: pkrell@uoquelph.ca

me53 is a conserved gene among the alpha- and betabaculoviruses, family Baculoviridae. Its deletion causes a 99.99% decrease in budded virus (BV) production in vitro and a slower spread of the virus. ME53 is a component of the nucleocapsid of both BV and occlusion derived virus (ODV). ME53 has two zinc fingers, lacks a nuclear localization signal (NLS), but does have a nuclear translocation sequence (NTS). It is expressed at both early and late time post-infection. Confocal fluorescence microscopy showed that at early times post-infection ME53 is found in the cytoplasm and translocates to foci in the cell membrane. At late times post infection, ME53 translocates to the nucleus accumulating at the peripheral region of the nucleus (in the ring zone), where ODV envelopment and embedding occur. That ME53 localizes to the membrane suggests that ME53 plays a role as a bridge between the envelope and nucleocapsid for efficient nucleocapsid envelopment and budding. We used the yeast two-hybrid system (Y2H) to identify ME53 binding protein partners. For that, me53 and Y2H libraries were constructed from mRNAs from AcMNPV-infected Sf21 cells and used as bait and prey, respectively. Following library screening, VP80, P6.9, LEF5 and cellular receptor for activated protein kinase C (RACK1) were found as ME53interacting partners. These four genes were cloned as bait and prey and tested reciprocally with ME53-prey and ME53-bait in yeast, with positive results for VP80 and LEF5 but not P6.9. GP64, VP39 and Exon0 were also cloned as prey and bait and an interaction was found between them and ME53 using the Y2H system. The interaction between ME53 and VP80 was the strongest interaction in terms of selective media stringency among other interactions found in this study. These data suggest VP80 might be the protein that ME53 uses as a chaperone to translocate to the nucleus late post-infection and this strong interaction might facilitate the attachment of the nucleocapsid to foci in the cellular membrane for enhanced envelopment and budding allowing for faster release and spread of the virus.

Deletion of the p94 (ac134) gene of the Autographa californica multiple nucleopolyhedovirus induces a delay in virus DNA replication, BV production, and insect mortality.

Vir10 •190

Miguel S. Andrade¹, Daniel M. P. Ardisson-Araújo², Daniel R. Sosa-Gomez³, **Fernando L. Melo**¹, Bergmann M. Ribeiro¹

¹Laboratory of Baculovirus, Cell Biology Department, University of Brasília, Brasília, DF, Brazil; ²Laboratory of Insect Viruses, Biochemistry and Molecular Biology Department, Federal University of Santa Maria, Santa Maria, RS, Brazil; ³Embrapa Soja, Londrina, PR, Brazil. Corresponding author: flucasmelo@gmail.com

Homologs of p94 (ac134) are present in the genome of several alpha- and betabaculoviruses) and also in some bracoviruses. The disruption of the p94 gene presented no effect on the ability of AcMNPV to infect S. frugiperda and T. ni larvae. However, the transient expression of the bracovirus P94 is able to suppress the immune system and developmental processes of lepidopteran larvae. To understand the evolutionary history of p94 gene, we constructed a phylogenetic tree of several homologs found in the Genbank. We found p94 homologs in alpha- and betabaculoviruses, bracoviruses and Lepidoptera. Our phylogenetic analysis shows that baculovirus sequences grouped together forming a well-supported group with betabaculoviruses sequences as a sister group to alphabaculovirus. The bracoviruses p94 homologs clustered together along with homologs from the monarch butterfly D. plexippus, confirming that horizontal gene transfer between bracovirus and Lepidoptera is likely common. To further investigate the biological impacts of this gene, we constructed three recombinant baculoviruses: vAc-pol-p94-del (p94 deleted), vAc-pol-p94-ha-rep (p94 repaired with an HA tag) and vAc-pol (parental virus). We found that the P94 was detected at 6 h post-infection (p.i.) in the cytoplasm of infected cells with maximum accumulation at 24 h p.i.. The deletion of the p94 gene in the AcMNPV genome resulted in a delay in DNA replication, BV production and IE1 expression. OB production in vitro was also decreased. Although the p94 gene of AcMNPV is not an essential gene, its presence is necessary for efficient viral DNA replication and establishment of viral infection.

AcMNPV-miR-6 down-regulates expression of host gene alg-2 Vir11 •191•



Jin Zhao, Weiwen Qin, Jinwen Wang

School of Life Science, Sun Yat-Sen University, Guangzhou 510275, China Corresponding author: wangjinw@mail.sysu.edu.cn

An Autographa californica multiple nucleopolyhedrovirus (AcMNPV) encoded miRNA, AcMNPV-miR-1 has been reported regulating viral own genes. However, AcMNPV encoded miRNA also regulate host genes. This study focused on AcMNPV-miR-6 targeting a host gene apoptosis linked gene-2 (alg-2) and the effects of this miRNA on viral infection. AcMNPV-miR-6 was expressed at 6 h.p.i. and reached the maximum expression at 24 h.p.i. detected by quantitative reverse transcription PCR (gRT-PCR), alg-2 was verified down-regulating by AcMNPV-miR-6 through dual-luciferase reporter assay. The expression levels of alg-2 mRNA and protein were decreased dramatically in AcMNPV-infected Sf9 cells at 24h.p.i. Administering AcMNPVmiR-6 mimic, the expressions of alg-2 mRNA and protein were decreased after transfected with alg-2 recombinant expression vectors in Sf9 and 293T cells, respectively. Constitutively overexpress AcMNPV-miR-6 recombinant viruses were constructed and mimic transient transfection was used in parallel to further explore the function of this miRNA. In the context of AcMNPV-miR-6 overexpression, the infectivity and production of budded

virus were reduced, the polyhedron formation was delayed, but the viral DNA replication did not show obvious alteration. It suggests that AcMNPVmiR-6 can auto-regulate viral infectivity and prolong the life span of the host cells, thereby facilitates the viral proliferation.

In vivo transcriptome under suppression subtractive hybridization Vir12 •192• (SSH) of the Betabaculovirus TnGV in Trichoplusia ni larvae

Ma. De los Angeles Bivián-Hernández, Ma. Fabiola León Galván, Mayra Chico Andrade, Ma. Cristina Del Rincón-Castro

> Graduate Program in Biosciences, Life Science Division University of Guanajuato Campus Irapuato-Salamanca 36500 Irapuato, Guanajuato, México Corresponding author: cdelrincon@uqto.mx

Five subtractive libraries under suppression conditions were constructed and sequenced obtained from Trichoplusia ni larvae infected at different times with the Betabaculovirus TnGV. At 24, 48, 72, 96 and 120 hours postinfection (h.p.i.), 35, 20, 7, 9 and 14 clones were obtained respectively. These were sent to be sequenced. At 24 h of the 35 clones, 10 sequences were showed no similarity with other proteins, 3 sequences showed similarity with hypothetical proteins, some of these sequences showed similarity with 2/3 actin complex proteins, heat shock proteins, factor elongation translation 2 isoform 1, elongation translation factor 2 isoform 2, ubiquitin E3 ligase, eukaryotic initiation factor 4A isoform 1 (eIF4A), galactinol synthetase-2. At 48 h.p.i. within the 20 clones, sequences with chaperone similarity, eukaryotic initiation factor 4A isoform 1 (eIF4A), copper center partial sequence, galactinol synthetase-2, cellulose synthetase, trans-criptional regulator of the TetR family were found. At 72 h.p.i. of the 7 clones sequenced, we identified 4 protein sequences that gave similarity to hypothetical proteins, and protein sequences with similarity to ubiquitin E3 ligase and RNA ligases. At 96 h.p.i. of the 9 clones sequenced, we found 2 sequences with similarity to hypothetical proteins, one with similarity to eukaryotic initiation factor 4A isoform 1 (eIF4A), one to cytochrome oxidase III subunit, and another to that of cellulose synthetase. At 120 hpi, of the 14 clones obtained, one of the sequences did not show similarity to other proteins, 4 sequences gave similarity to hypothetical proteins, and sequences were found with similarity to the ubiquitin E3 ligase, to the partial center copper sequence, and the B1,3-glucanases. In some tissues in infected T.ni larvae, the expressed proteins are controlled by the Betabaculovirus TnGV, and this leads to a reduction of proteins and mRNAs of the cell. On the other hand, the host cell responds to the viral infection with the overexpression or repression of some point genes and the virus takes advantage of the machinery and proteins of the host cell for its own replication.

Functional assay of ORF105 (APSUP) from Lymantria xylina multiple nucleopolyhedrovirus (LyxyMNPV)

Vir13 •193•



Yu-Shin Nai¹, Ju-Chun Chang¹, Se Jin Lee², Jae Su Kim²

¹Depatment of Biotechnology and Animal Science, National Ilan University, Yilan, Taiwan; ²Department of Agricultural Biology, College of Agriculture and Life Sciences, Chonbuk National University, Jeonju 561-756, Republic of Korea Corresponding author: ysnai@niu.edu.tw

Baculoviral anti-apoptotic genes, p35 and iap (inhibitor of apoptosis), play important roles in the initiation stage of viral infection. Recently, a new antiapoptotic gene (apoptosis suppressor, apsup) was identified from Lymantria dispar multiple nucleopolyhedrovirus (LdMNPV); herein, the apsup homologues gene (Lyxy105) was amplified and cloned from L. xylina MNPV (LyxyMNPV) and further studied. A 1,002 bp of PCR product was amplified by using gene specific PCR and sequenced. Comparison of nucleotide and putative amino acid sequences to 19 homologues genes, the Lyxy105 showed high nucleotide identity (88%) and high amino acid identity (85 %) and similarity (88 %) to that of LdMNPV. Moreover, five conserved protein domains were identified. Functional analysis of Lyxy105 was performed using an overexpression method in Sf9 cells. From our result, full-length LYXY105 could inhibit the Drosophila RPR protein (DRPR)-induced apoptosis. More data should be provided (i.e., gene expression profile and anti-apoptotic activity of ActD-induced apoptosis, etc.) to further evaluate the full picture of the anti-apoptotic mechanism of LYXY105.

Construction of hyper baculovirus expression vector by the optimization of enhancer factors

STU Vir14 •194•

JiHoon Lee, WonSeok Gwak, Jiln Ma, DongJun Kim, HwiGeon Yun, JonaMin Oh, SooDona Woo

Department of Agricultural Biology, College of Agriculture, Life & Environment Science, Chungbuk National University, Cheongju 28644, Korea Corresponding author: sdwoo@cbnu.ac.kr

The baculovirus expression vector system (BEVS) is an effective and widely used system for the production of recombinant proteins in insect cells or larvae. However, the expression efficiency of recombinant proteins using the polyhedrin promoter could not acquire the protein yields observed for native polyhedrin. To overcome this limit, various enhancer factors have been developed by many researchers and enhanced successfully the production efficiency of recombinant. In this study, we tried to develop hyper expression vector by the optimal combination of previously reported various enhancer factors. The selected enhancer factors for optimal expression consists homologous region 5 (hr5), VP39 promoter and burst sequences. Enhanced green fluorescent protein (EGFP) was fusion expressed as recombinant protein for visual and numerical observations. Seven recombinant viruses were generated using these enhancer factors to compare expression efficiency. Each recombinant virus showed different expression levels each other, and the most of expression level was observed with higher than those of the previous vectors. The present study suggests a new option for higher expression of useful foreign recombinant protein using the BEVS.

The effects of transfection of bacmid DNA together with the helper plasmid on determination of defective virus multiplication capacity

Vir15 •195•

Zhimin Tang¹, Qinying Lai¹, Meijing Yuan¹, **Wenbi Wu**¹, Kai Yang¹

¹ State Key Laboratory of Biocontrol, Sun Yat-sen University, Guangzhou 510275. China Corresponding author: wuwenbi3@mail.sysu.edu.cn

The Bac-to-Bac Baculovirus Expression System has been widely used in the studies of baculovirus gene functions because of the rapid and efficient way it provides to generate recombinant baculoviruses. The Escherichia coli strain DH10Bac from the system contains a baculovirus shuttle vector (the bacmid bMON14272) and a plasmid that encodes the transposase to provide the Tn7 transposition in trans (the helper plasmid pMON7124). After generating recombinant bacmid by transposition, the presence of the helper plasmid will not affect the transfection of the bacmid DNA. However, we noticed that when transfecting DNA in a quantified way, the existence of the helper plasmid may affect the determination of a defective-virus multiplication capacity. In the present study, we used an AcMNPV orf34 (ac34)-knockout bacmid DNA as a backbone to generate different recombinant bacmids. After the transposition, the helper plasmid was kept in the DH10Bac cells or removed from the cells by electroporation of the bacmid DNA into the helper plasmid-free DH10B cells. To clarify the effects of the helper plasmid when determine the multiplication capacity of the viruses, we transfect the DNA mixtures (purified from DH10Bac cells with helper plasmid existed) or only the bacmid DNAs (purified from the DH10Bac cells free of helper plasmid) into Sf9 cells in a quantified way. After transfecting the quantified amount of DNA into Sf9 cells, we photographed the cells and measured the virus titers at different time points post transfection. The results were analyzed and discussed.

Induced plant defenses delay within-instar developmental resistance of fall armyworms, Spodoptera frugiperda, to the baculovirus SfMNPV Vir16 •196

Ikkei Shikano¹, Elizabeth M. McCarthy², James Slavicek³, **Kelli Hoover**¹

¹Department of Entomology and Center for Chemical Ecology, Pennsylvania State University, University Park, PA, USA: ²Department of Chemistry, Indiana University, Bloomington, IN, USA 3USDA Forest Service, Delaware, OH, USA Corresponding author: kxh25@psu.edu

Resistance of larval lepidopterans to baculoviruses increases with each larval instar and can also increase over time within an instar. Most studies measure

developmental resistance at incremental time points using insects that were feeding on a single defined food source, usually artificial diet. However, insects in the field are exposed to food sources that vary widely in nutritional and allelochemical composition. A few studies have shown that food quality (different host plant species or plant vs. artificial diet) can influence levels of developmental resistance. One of the most common variations in plant quality experienced by caterpillars in the field is differences in constitutive and/or induced plant defenses due to caterpillar feeding. We investigated the effects of induced plant defenses on developmental resistance in the fall armyworms, Spodoptera frugiperda. We fed newly molted fourth instars foliage from non-induced or induced soybean (Glycine max) plants for 2, 4, 6, 8 or 12 h prior to baculovirus (SfMNPV) challenge. Plants were induced with the phytohormone, jasmonic acid, which stimulates anti-herbivore defensive responses. We found that the mortality of SfMNPV-challenged larvae decreased significantly more over time (i.e., resistance increased over time) when larvae fed on non-induced than on JA-induced foliage before viruschallenge. This suppression of developmental resistance over time on JAinduced foliage was associated with slower larval weight gain and there was not a reduction in consumption of foliage. Results imply that plant-mediated larval growth inhibition can increase susceptibility to baculovirus. Moreover, our findings indicate that larval weight gain, not time post molt, was responsible for within-instar developmental resistance of fall armyworm to SfMNPV.

The hemolymph of Mythimna separata larvae infected STU Vir17 •197• with an entomopoxvirus kills parasitoid-derived cell



Rie Ohta, Maki N. Inoue, Yasuhisa Kunimi, Madoka Nakai

Tokyo University of Agriculture and Technology, Japan Corresponding author: madoka@cc.tuat.ac.jp

When the gregarious endoparasitoid Cotesia kariyai parasitizes Mythimna separata larvae infected with Mythimna separata entomopoxvirus (MySEV), its embryos and larvae die inside the host. C. kariyai larvae die even in the virus-free hemolymph of M. separata larvae previously infected with MySEV, which we call VFP (virion-free plasma). Previous studies showed that a 28kDa protein purified from the VFP was lethal to C. kariyai. This protein was named parasitoid killer toxin (PKT) and MySEV encodes pkt homologues. The VFP shows killing activity to some Microgastrinid parasitoids including C. kariavai, but not to other braconid parasitoids including Meteorus pulchricornis. However, it remains unclear whether the VFP affects not only at the individual level but also at the cellular level. In this context, understanding how the VFP acts on the parasitoid cell is important in terms of elucidating PKT mode of action. In this study, we examined the effect of the VFP on cultured cells established from C. karivai (Ck1) and M. pulchricornis (Mp4). Ck1 showed unusual morphology, such as loss of pseudopod, blebbing and floating when exposed to the VFP. Moreover, the VFP suppressed Ck1 cell prolification completely and increased mortality of Ck1 significantly 24 hours after exposure. On the other hand, such deleterious effects have not been observed in Mp4. These results suggest that the VFP, containing PKT, specifically kills C. kariyai at the cellular level.

Effect of gut bacteria on Sindbis virus replication in the mosquito Aedes aegypti





Sarah Pendell. Rollie J. Clem

Division of Biology, Kansas State University, Manhattan, KS, USA Corresponding author: rclem@ksu.edu

The Aedes aegypti mosquito is an important vector for several arboviruses that are a major public health concern around the world, including dengue virus, Zika virus, and chikungunya virus. A. aegypti is found in tropical and subtropical regions, with travelers enabling the spread of viruses to other regions that may not be endemic. We use Sindbis virus (SINV; Togaviridae) as a model arbovirus to help further understand viral replication within this species of mosquito. Past experiments by our group have consistently revealed a high degree of variability in SINV titer between individual mosquitoes when the mosquitoes are infected by the oral (bloodfeeding) route, with titers ranging from 10^2 to 10^6 infectious units per mosquito. In

contrast, mosquitoes that are infected by intrathoracic injection exhibit consistently high virus titers in the range of 10⁴ to 10⁶. The reasons for this difference between infection routes are unclear, but may be related to the fact that intrathoracic injection bypasses the midgut, while oral infection requires initial replication in the midgut before the virus can disseminate to other tissues. In this study, we wanted to determine whether resident gut microbes affect SINV replication via oral infection, and whether the gut microbiome could contribute to the extreme variability observed in virus titers by reducing virus replication in some individual mosquitoes. To test this, we treated mosquitoes with antibiotics prior to oral infection, in order to reduce the numbers of gut bacteria. We found that antibiotic treatment was successful in reducing the numbers of cultivatable gut bacteria. However, when antibiotic-treated mosquitoes were infected with SINV by the oral route, virus titers were similar to control mosquitoes that had not been treated with antibiotics. These results do not support the hypothesis that the presence of gut bacteria has a major influence on SINV replication in orally infected A. aegypti mosquitoes, in contrast to previous reports that gut bacteria reduce arbovirus replication.

Evaluation of Anticarsia MNPV and Pseudoplusia SNPV co-infection in insect cell culture

Vir20

Claudia Efigenia Pereira Silva^{1,2}, William Sihler¹, Ana Cristina M.M. Gomes¹,Marlinda Lobo de Souza¹, **Márcio Martinello Sanches**¹

¹Embrapa Recursos Genéticos e Biotecnologia,Brasília, Brasil; ²Universidade Paulista,Brasília, Brasil Corresponding author: marcio.sanches@embrapa.br

It is well known that one particular insect cell line may be susceptible to different baculovirus. Therefore co-infection of a host cell with two distinct viruses can result in co-oclusion phenotype. This feature may be important to reduce costs of baculovirus in vitro production. The baculoviruses Pseudoplusia includes single nucleopolyhedrovirus (PsinSNPV) and Anticarsia gemmatalis multiple nucleopolyhedrovirus (AgMNPV) that infect two important soybean pests, Chrysodeixis includens and Anticarsia gemmatalis respectively, were tested regarding their infectivity to different cell lines. The cell lines IPLB-SF-21AE, Sf9 and BTI-Tn-5B1-4 were previously evaluated as productive to AgMNPV infection. In this work the susceptibility of these cell lines to single infection of PsinSNPV and mixed infection of PsinSNPV and AgMNPV was tested. The cells were seeded at a density of 1X10⁶ per 60mm² dish. The viruses were obtained through hemolymph from infected larvae at 4 d.p.i. and allowed to adsorb to cells during 1 hour (P0). Infected cells were kept in TNMFH complete medium at 27°C. Morphological analysis was initially monitored by light microscopy during five days. Then, the supernatants were collected for new infections (P1) using the same procedure. At 5 d.p.i. the supernatants were collected for the second passage (P2). The ultrastructure of the occlusion bodies was analyzed by Transmission Electron Microscopy (T.E.M.) for PO passage. The amount of DNA obtained from Budded Viruses (BVs) of the three passages was monitored by real-time PCR (qPCR) using Sybr green system with primers designed to gp64 of AgMNPV and photolyase of PsinSNPV. The experiment was repeated twice. The qPCR methodology presented the limit of detection 10⁻⁵ ng DNA for both viruses. The results demonstrated successful coinfection in these cells, but the amount of PsinSNPV tends to decrease in serial passages whereas the amount of AgMNPV tends to increase in the first passage and to stabilize in the second passage. Also, in PsinSNPV single infection experiments, the amount of BVs DNA has a tendency to reduce in further passages. Cell culture visualization by optical microscopy, in conjunction with the qPCR assays, indicates that infection with PsinSNPV is restricted to individual cells, resulting in low polyhedra production. Support:FAP-DF

Production of porcine epidemic diarrhea virus (PEDV) VLP using the baculovirus expression vector system in Bombyx mori cells

STU Vir21

WonSeok Gwak, JiHoon Lee, Jiln Ma, Dong Jun Kim, HwiGeon Yun, SooDong Woo

¹Department of Agricultural Biology, College of Agriculture, Life & Environment Science, Chungbuk National University, Cheongju 28644, Korea Corresponding author: sdwoo@cbnu.ac.kr

Porcine epidemic diarrhea virus (PEDV) causes viral diarrhea disease in pigs resulting in large economic loss to the swine industry in Asia. However, there is no effective vaccine to prevent PEDV. Viral particles of PEDV consist of a four structural proteins are Spike(S) protein, envelope(E) protein, glycosylated membrane(M) protein, and unglycosylated RNA-binding nucleocapsid(N) protein. The PEVD virion is assembled and released into the lumen of the endoplasmic reticulum (ER)-Golgi intermediate compartment (ERGIC), followed by secretion by exocytosis. The M and E proteins plays important in virus assembly and budding. The S protein mediated responsible for receptor binding and membrane fusion during viral infection and therefore the main target of neutralizing antibodies. Virus-like particles (VLPs) are consisted of one or more viral structural proteins, and their morphologies closely resemble those of the native virus. VI Ps have no virulence and can elicit robust immune responses as compared with inactivated or live-attenuated virus vaccines. Thus, in this study, we tried two methods for VLP construction, one is traditional method and the other is chimeric VLP method using the influenza matrix protein. The PEDV four structural proteins (S, M, N and E) which were co-expressed in Bombyx mori 5 (Bm5) cells formed successfully VLPs. The chimeric VLP was also constructed with a matrix 1 protein from influenza virus and a modified spike protein that was generated by mutation or deletion of KxHxx motif in the cytoplasmic tail. Both methods could produce successfully PEDV VLPs with spike protein on their surfaces.

BACTERIA DIVISION POSTERS

Sex ratio distortion in tea pest *Homona magnanima*: STU Bac01 •199• A complicated association of host and endosymbiotic microbes

Hiroshi Arai, Takumi Takamatsu, Tatsuya Hirano, Madoka Nakai, Yasuhisa Kunimi, Maki N. Inoue

Tokyo University of Agriculture and Technology, Tokyo, Japan

Corresponding author: makimaki@cc.tuat.ac.jp

Endosymbiotic microbes are frequently detected from a variety of insects, and some of them interact with their host in some mutualistic or parasitic manners. For example, some microbes are known to manipulate host reproduction via male-killing or feminization. Homona magnanima (Tortricidae, Lepidoptera) is a serious tea pest, which widely distributed in East Asia. From field surveys across Japan, Wolbachia, Spiroplasma and RNA virus were detected from H. magnanima, and the latter two microbes caused sex-ratio distortion in their host. Although Wolbachia-infected Japanese host exhibited normal sex-ratio, we recently collected a Wolbachia infecting H. magnanima from Taiwan, which showed female biased sex-ratio. Here, we characterized both Japanese and Taiwanese Wolbachia strains to investigate factors correlated to the difference of phenotype in terms of sex-ratio distortion. First of all, we conducted phylogenetic analysis of Wolbachia by using Wolbachia MLST genes. As a result, Japanese host harbored three distinct Wolbachia strains (wHm-a, -b and -c) and Taiwanese host harbored a Wolbachia strain (wHm-t). Although wHm-c and wHm-t showed 100% similarity at MLST level, only wHm-t caused sex-ratio distortion to the host. We next surveyed the density of Wolbachia (wHm-a, -b, -c and -t) in their host to examine the correlation between density and the phenotype. The density of wHm-t was significantly higher than that of any other Wolbachia strains. In particular, over 1000-fold difference in density was detected between wHm-t and wHm-c. We also observed the difference in density between two wHm-t infecting host lines and named wHm-t with high-density as 'WTH, line and wHm-t with low-density as 'WTL, line. In terms of the phenotypes, WTH line exhibited higher total mortality than W^{TL} line. Almost

all males of \boldsymbol{W}^{TH} line were killed during embryonic stage like Spiroplasmainfected host, whereas males of W^{TL} line were killed during both embryonic and larval stage and exhibited same symptoms of RNA virus; such as white body color and abortion of larva. Given these results, the phenotype of sexratio distortion caused by Wolbachia is likely to depend on Wolbachia strain and its density, and wHm-t can exhibit various phenotypes in terms of malekilling in H. maananima.

Bacteria Collection of Invertebrates, one of the four Brazilian collections of the Global Net of Biological **Resources: Challenges and expectations**

Bac02 •200•

Lílian Botelho Praça, Ester Yoshie Yosino da Silva, Rose Gomes Monnerat

Embrapa Recursos Genéticos e Biotecnologia, Parque Estação Biológica, Brasília - DF. Corresponding author: lilian.praca@embrapa.br

In 2005, a project supported by Ministry of Science and Technology in Brazil was started in order to prepare Brazilian collections to act as National Biological Resource Centers. 16 collections were evaluated regarding compliances to the ISO 17025 standard requirements and four collections were selected aiming a Quality System (QS) implementation in order to prove their competence in doing technical and specialized services and in offering certified biological materials. The Embrapa Brazilian Invertebrate Bacteria Collection was one of the four collections selected by the project. Since then, the collection team and the Quality Management Center of Embrapa conducted the following activities to implement the ISO 17025: diagnosis of the actual situation of the collection regarding the Standard requirements; staff training; preparation of a Quality Plan to be used through the mapping process, preparation of QS documentation (Quality Manual, master list, 37 technical and 31 management procedures; 11 equipment instructions); implantation of the 5S program and disposal management; validation methods, quality control of the validation methods, physical infrastructure adequacy; equipment maintenance, calibration and qualification and external evaluations. During four years, the collection project has been externally evaluated three times in 2005, 2007 and 2009. The collection received respectively 34, 21 and 4 non-conformities. These results show the great progress of the collection activities within the QS. As a result of this effort, the collection, besides attending the ISO 17025 requirements, implemented the Standard for Biological Resource Center of the Organization for Economic Cooperation and Development (OECD). Thus, the collection can be part of the Network of Biological Resource Centers when Inmetro establishes a specific program for assessing conformity of Brazilian collections. To attend the OECD standard (NIT DICLA 061), all the information of the collection was inserted in a data bank and all strains were preserved under two methods (over 2600 strains on filter paper and lyophilized). These actions provided the collection to obtain three tests accredited by Cgcre/Inmetro in 2015 and have this accreditation maintained in 2017: identification, viability and purity test of Bacillus thuringiensis and Lysinibacillus sphaericus.

Bacillus thuringiensis israelensis reference material production to be used in assessment tests accreditation of bioinsecticides toxicity for controling Aedes aegypti

Bac03 •201•

Lílian Botelho Praça¹; Zonaite Gomes Almeida¹; Ester Y. Y. da Silva¹;

Carla Ferreira Caixeta¹; Carlos Marcelo Soares², Rose Gomes Monnerat¹ ¹Embrapa Recursos Genéticos e Biotecnologia, Parque Estação Biológica, Brasília – DF, Brasil; ²Instituto Mato-Grossense de Algodão, Cuiabá - MT, Brasil. Corresponding author: lilian.praca@embrapa.br

The difficulty in finding suppliers of certified reference materials is a limitation that laboratories with accredited tests face today. Embrapa was accredited for the toxicity determination of B. thuringiensis to control mosquitoes and due to this it was necessary to look for viable and non-costly alternatives to produce its own reference material. In 2016 Embrapa started to produce its own material. The B. thuringiensis israelensis strains was multiplied in a fermenter and lyophilized. The purity and viability of the material was then evaluated through seeding in Embrapa medium and the features analyzed through repeatability, reproducibility and sensitivity parameters. The material stability was also evaluated through LC 50

bioassays of 2nd instar Aedes aegypti larvae. The LC 50 results were calculated by Poloplus Probit program and the data was released on an average control chart previously built to evaluate the stability of the material over time. Other evaluated feature was the material stability by means of LC 50 bioassays using 2nd instar Aedes aegypti larvae. The purity and viability results presented a 100% satisfactory outcome and The LC 50 presented values between 0.203 and 4.37 ng/mL. These results were within the control limits, demonstrating the stability of the reference material data and the method suitability. So it is possible to assure our clients that Embrapa provides services with technical competence, ensuring the quality of its

Superior in vivo growth of Bacillus thuringiensis strains suggests a specialisation towards insect pathogenicity and single original B. thuringiensis clade

STU Bac04 •202

C J Manktelow, G Simmons, B Raymond

College of Life and Environmental Sciences, University of Exeter, Penryn Campus Corresponding author: cjm244@exeter.ac.uk

Bacillus thuringiensis (Bt) is a gram-positive bacterium, used as a biopesticide and source of Crystal (cry) toxin genes for transgenic crops. Bt strains have been identified by the presence of cry toxins, but genomic data has shown that Bt is not monophyletic, instead being distributed throughout the Bacillus cereus family. However, cry genes are often plasmid-borne and horizontal transfer rates are high in this family. It is therefore hypothesised that cry toxins evolved in a single 'Bt' clade, before being lost/transferred giving the current Bt distribution in the family. Additionally, it has been previously suggested that Bt is not an obligate insect pathogen. Taken together, Bt strains in the supposed 'Bt' clade are expected to show lower spore production in vitro and higher fitness in vivo suggesting that Bt originally evolved as a specialised insect pathogen. Spore production in media showed that 'Bt' clade strains produced fewer spores than 'Bc' clade ones, while in vivo competitions against a standard strain in Plutella xylostella showed that 'Bt' clade strains had higher relative fitness. 'Bt' clade strains also showed a negative relationship between spore production and in vivo fitness whereas 'Bc' ones showed a positive relationship, suggesting specialisation in the 'Bt' clade.

Genome sequence of mosquitocidal native Bacillus thuringiensis strain from Argentina



J. Nicolás Lazarte, Rocio P. Lopez, Corina M. Berón

Instituto de Investigaciones en Biodiversidad y Biotecnología (INBIOTEC - CONICET), FIBA, Mar del Plata, Argentina. Corresponding author: corina.beron@inbiotec-conicet.gob.ar

Bacterial insecticides have been used for the control of insects of public health importance. Mosquito-borne diseases are among the leading causes of mortality and morbidity in humans. For the control of these viral and parasitic diseases the continuous development of different management strategies of mosquito populations is essential. In a previous work, we characterized an Argentinian Bacillus thuringiensis strain (FCC 41) that exhibited mosquitocidal activity against Aedes aegypti, Aedes (Ochlerotatus) albifasciatus, Culex pipiens and Culex apicinus. In this work, we conducted the whole genome sequencing of B. thuringiensis native strain using next generation sequencing on an Illumina 1500 HiSeq. The paired short reads were assembled with SPADES, and a preliminary annotation was obtained using PROKKA. Furthermore, we identified Cry proteins coding sequences (CDS) using a HMMER search. With this approach we found 10 positive matches: these CDS were identified as two Cry4-like, two Cry19-like with their corresponding downstream Orf2 regions, a Cry24Ca with its Orf2, a Parasporin 3-like, and three contiguous small CDS within the same operon with homology to domains of several Cry proteins. The next step will be to clone and express these crv-like nucleotide sequences in heterologous systems to determine their specific toxicological activity and potential application against vector insects of public health importance. Besides, we will analyze Orf2-like regions in order to determine their function in the crystallization and/or stability of Cry toxins. Supported by ANPCyT (PICT No 2013-0431 y PICT No 2015-0575) and Universidad Nacional de Mar del Plata (15/E692-EXA742/15).

Toxicity of Bacillus thuringiensis Cry1Ac toxin in different Plutella xylostella (L.) (Lepidoptera: Plutellidae) populations

Bac06 •204•

Caroline Placidi De Bortoli¹, **Rafael Ferreira dos Santos**¹, Alessandra Marieli Vacari¹, Neil Crickmore², Sergio Antonio De Bortoli¹, Ricardo Antonio Polanczyk1

¹Department of Plant Protection, Sao Paulo State University, 14884-900, Jaboticabal, Sao Paulo, Brazil: ²Department of Biochemistry, School of Life Sciences, University of Sussex, BN1 9QG, Brighton, UK. Corresponding author: carubortoli@yahoo.com.br

Plutella xylostella (Linnaeus, 1758) (Lepidoptera: Plutellidae), the diamondback moth (DBM), is a major insect pest of crucifers (Brassicaceae) worldwide. The most common bioinsecticides used to control P. xvlostella are based on the entomopathogenic bacterium Bacillus thuringiensis (Bacillaceae) (Bt). Although many studies have focused on the action of Bt on various agricultural pests, such as DBM, many doubts still persist, particularly regarding the toxicity of Bt proteins. We analyzed the virulence of Cry1Ac protein in Brazilian populations and in a Hawaii population of *P. xylostella*. Bioassays of susceptibility in five Brazilian populations (PC, PA, PX, SBT, and BT) and one Hawaii population (NO_QA) of P. xylostella and B. thuringiensis Cry1Ac protein, estimating the virulence of the toxin, were performed. Also a complementation crossing between NO-QA and Bt was performed estimating the virulence of the toxin on the first and second generation. Seven concentrations were used to calculate the values of LC₅₀. Three replicates were performed, with each replicate being a petri dish containing 20 larvae, totaling 60 insects per concentration for each population. The PC, PA, PX, SBT, BT and NO_QA P. xylostella populations exhibited different levels of susceptibility to the Cry1Ac toxin. The PA, PX, and SBT populations showed LC₅₀ values of 0.022, 0.016, and 0.058 $\mu g/mL$. The LC₅₀ estimate for the BT and NO_QA populations was 1.26 and 1.51 $\mu g/mL$, while that for PC it was $0.001 \, \mu g/mL$. The LC_{50} estimated for the NO_QA population was 1510 fold higher than PC P. xylostella population. In the complementation crossing the LC₅₀ estimated for NO QA male X Bt female crossing was 2.08 µg/mL in the first generation and 0.37 $\mu g/mL$ in the second, while for NO_QA female X Bt male crossing was 1.73 $\mu g/mL$ in the first generation and 0.25 $\mu g/mL$ in the second. Support: FAPESP.

Factors involved in resistance to Bacillus thuringiensis of Brazilian Plutella xylostella populations.

Bac07 •205•



Caroline Placidi De Bortoli¹, Rafael Ferreira dos Santos¹, Siobhan Clerkin², Neil Crickmore², Sergio Antonio De Bortoli¹, Ricardo Antonio Polanczyk¹

¹Department of Plant Protection, Sao Paulo State University, 14884-900, Jaboticabal, São Paulo, Brazil; ²Department of Biochemistry, School of Life Sciences, University of Sussex, BN1 9QG, Brighton, UK, Corresponding author: carubortoli@yahoo.com.br

Plutella xylostella is a major insect pest of cruciferous crops worldwide. Although controlled with both synthetic and biological insecticides it can rapidly evolve resistance to a variety of insecticides. The most common biopesticides used to control P. xylostella are based on the entomopathogenic bacterium Bacillus thuringiensis. Although many studies have been performed on Bt, the mode of action is still not fully understood. High levels of resistance to Crv1 toxins, due to reduced toxin binding, have been genetically linked to mutations or expression alterations of receptor genes. Resistance can also result from the overexpression of genes involved in protecting the insect against Bt toxins, genes that are normally expressed in response to exposure to the toxin. A wide diversity of genes are differentially expressed in the midgut of resistant insects, this suggests that a variety of cell processes may be involved in the preservation of resistance. Recent discoveries have shown that mutations in the gene encoding an ABCC2 transporter are responsible for resistance to Bt toxins in various different insect species. One group found no association between mutations in ABCC genes and resistance, they subsequently compared levels of expression for all five ABCC genes in BtR-1 locus in susceptible and resistant strains of P. xylostella using qPCR. Their analysis revealed that ABCC2 was significantly down-regulated in the resistant population as was the expression of another putative receptor ALP. They also found that MAP4K4 was constitutively upregulated in larvae from the resistant strain compared to the susceptible strain. In our study we tested the hypothesis that the susceptibility of P.

xylostella to Bt correlates with the level of expression of components of a putative stress-response regulon. In particular we were testing whether our resistant populations would show down-regulation of ALP, APN, ABCC2 genes and up-regulation of CDKAL1 and MAP4K4 genes; our research demonstrated that there were no patterns in their expression that associated with their resistance/susceptibility. We also investigated the DNA sequence of ABCC2 cDNA and found a frameshift mutation in the resistant population that could be responsible for the resistant phenotype. Support:

Laboratory resistance Plutella xylostella (L.) (Lepidoptera:Plutellidae) to Cry1Ac toxin and HD1 strain of Bacillus thuringiensis



Caroline Placidi De Bortoli¹, Rafael Ferreira dos Santos¹, Gilmar da Silva Nunes¹, Camila Pires Cardoso¹, Alessandra Marieli Vacari¹, Neil Crickmore², **Sergio Antonio De Bortoli**¹, Ricardo Antonio Polanczyk¹

¹Department of Plant Protection, Sao Paulo State University, 14884-900, Jaboticabal, São Paulo, Brazil; ²Department of Biochemistry, School of Life Sciences, University of Sussex, BN1 9QG, Brighton, UK. Corresponding author: carubortoli@yahoo.com.br

The diamondback moth (DBM), Plutella xylostella (Linnaeus, 1758) (Lepidoptera: Plutellidae), is a key pest of crucifers. Although can be controlled with insecticides, P. xylostella can quickly develop resistance to insecticides, such as those of from Bacillus thuringiensis. We analyzed the virulence of Cry1Ac protein and HD1 strain in Brazilian populations of P. xylostella. Bioassays in two Brazilian populations (susceptible and resistant) of P. xylostella estimating the virulence of the Cry1Ac and HD1, were performed. The resistant population came from the susceptible and was selected in the laboratory with HD1 every generation. Seven concentrations were used to calculate the values of LC₅₀. Three replicates were performed, with each replicate being a petri dish containing 20 larvae, totaling 60 insects per concentration for each population. The P. xylostella populations exhibited different levels of susceptibility to the Cry1Ac toxin and HD1 strain. The HD1 strain showed LC₅₀ values of 9.8 × 10⁵ spores/mL to resistant population and 0.93 × 10³ to susceptible population. The Cry1Ac protein showed LC $_{50}$ values of 1.26 $\mu g/mL$ to resistant population and 0.016 $\mu g/mL$ to susceptible population. The LC_{50} estimated for the HD1 strain was 1054 fold higher and for Cry1Ac was 78.7 higher for resistant population than susceptible population. Support: FAPESP.

Study of the susceptibility of Spodoptera frugiperda (LEPIDOPTERA: Noctuidae) in corn cultures expressing the Cry1F toxin from Bacillus thuringiensis in Brazil

Bac09 •207•



Cristina Macedo ^{1,2}, Erica Martins⁴, Paulo Queiroz⁴, Lilian Praça², Marcelo Soares⁴, Barbara Eckstein², Isabel Gomez³, Mario Soberon³, Alejandra Bravo³, Rose Monnerat²

¹Fundação de apoio a pesquisa (FAP-DF), ²Embrapa Recursos Genéticos e Biotecnologia (CENARGEN), Brasília, Brazil; ³Universidad Nacional Autonoma de México, Morelos – Mexico; ⁴Instituto Mato-Grossense do Algodão (IMAmt) – Brazil Corresponding author: crispepi@yahoo.com

Brazil ranked second only to the United States in hectares planted to genetically modified crops in 2016. The use of the technology of genetically modified plants expressing toxins with activity obtained from Bacillus thuringiensis (Berliner) (Bt) emerged as an important tool for reducing the intensity of the application of agrochemicals and showed to be effective in controlling some of the major cotton pest. Recently corn producer in the Cerrado region reported that the control of Spodoptera frugiperda with Bt corn expressing Cry1Fa has decreased, forcing them to use chemicals to reduce the damage caused by this insect pest. Many putative Cry toxin receptors have since been reported, of which the best characterized are the aminopeptidase N (APN) receptors and alkaline-phosphatases (ALPs). The bioassay data reported here show that insects collected from Cry1Fa corn in the Cerrado region were resistant to Cry1Fa suggesting that resistance contributed to field failures of Cry1Fa corn to control S. frugiperda. This study the detection of the midgut receptors of the larvae of S. frugiperda (BBMV) was analyzed by determining the specific activity of ALP and APN by SDS-PAGE gel using the substrate for detecting the activity, and also western-blot

with ALP and APN antibodies of Manduca sexta (Lepidoptera). These data indicate that the levels of ALP receptors detection and the APN of some of the resistant populations showed low levels of ALP in all analysis, which is a low activity of this receptor when compared to the susceptible population. Demonstrating that the insect populations may be involved with ALP receptors, but not with APN, in these Brazilian populations.

Analysis of cross-resistance of resistant insect colonies STU Bac10 •208• from different species to Vip3Ca from Bacillus thuringiensis

Joaquín Gomis-Cebolla¹, Tom Walsh², Sharon Downes³, Wendy Kain⁴, Ping Wang⁴, Kathy Leonard⁵ Tom Morgan⁵, Brenda Oppert⁵ and Juan Ferre¹

¹ ERI de Biotecnología y Biomedicina (BIOTECMED), Department of Genetics, Universitat de València, Burjassot, Spain; ² CSIRO, Black Mountain Laboratories, Canberra, ACT 2601, Australia: 3 CSIRO, Myall Vale Laboratories. Kamilaroi Highway, Narrabri, NSW 2390, Australia; ⁴ Department of Entomology, Cornell University, New York State Agricultural Experiment Station, Geneva, NY 14456, USA; 5 USDA, Agricultural Research Service, Center for Grain and Animal Health Research, 1515 College Avenue, Manhattan, KS, USA Corresponding author: juan.ferre@uv.es

Bacillus thuringiensis (Bt) is a gram positive bacterium used in insect control because some strains produce insecticidal proteins. The Vip3 proteins are synthesized and secreted to the medium during the vegetative growth phase. The mode of action of Vip3 proteins parallels that of Cry proteins in that it involves activation by gut proteases, recognition and binding to midgut receptors, pore formation, and cell lysis, finally causing the death of the insect. We tested the susceptibility to the Vip3Ca protein of susceptible and Cry1-, Cry2-, and Vip3-resistant insect colonies from different insect species, to determine whether resistance to other insecticidal proteins could confer cross-resistance to Vip3Ca. As expected, the insect colonies resistant to Cry1 (from Plodia interpunctella, Helicoverpa armigera and Trichoplusia ni) or resistant to Cry2 (from H. armigera and T. ni) didn't show cross resistance to the Vip3Ca protein. However, insect colonies resistant to Vip3Aa or to Vip3Aa/Cry2Ab (from H. armigera) showed cross-resistance to the Vip3Ca protein. These results are in agreement with the published binding models where Vip3 proteins do not compete with Cry1 and Cry2 for binding sites, but different Vip3 proteins (Vip3A and Vip3Ca) compete for the same binding sites.

Modification of a Bacillus thuringiensis toxin to target soybean aphid (Aphis glycines)

Biviana Flores Escobar 1,2, Benjamin Deist 2,

Corresponding author: b.floresescobar@ufl.edu

M. Teresa Fernandez Luna ², Bryony Bonning^{1, 2} ¹ Department of Entomology and Nematology, University of Florida. Gainesville, Florida USA; ²Department of Entomology, Iowa State University, Ames, Iowa, USA

Current hemipteran pest management is heavily reliant on chemical pesticides. Management of this order of insects represents one of the greatest challenges in agricultural biotechnology. Bacillus thuringiensis (Bt)based technologies have been widely employed for decades to manage populations of lepidopteran and coleopteran pests, but few Bt toxins have activity against hemipteran species. To overcome this limitation, we have applied an innovative strategy to enhance the activity of Bt toxins against hemipteran pests. Having isolated a peptide that binds aminopeptidase N in the gut of the pea aphid (Acyrthosiphon pisum), we demonstrated that addition of GBP3.1 to the Bt Cyt2Aa toxin enhanced activity against the pea aphid and the green peach aphid (Myzus persicae) (Chougule et al., 2013). We applied this same approach to new hemipteran targets including the soybean aphid (Aphis glycines). Soybean aphid gut binding peptides were selected and the Cry4Aa toxin modified with these peptides for enhanced activity against soybean aphid. We used two approaches for isolation of soybean aphid gut binding peptides, an in vivo screen by feeding aphids on a phage display library and eluting phage bound to the gut epithelium, and in vitro selection for peptides that bound to recombinant soybean aphid gut proteins, aminopeptidase N and alkaline phosphatase. Five sites were selected for insertion of gut binding peptides into the Cry4Aa toxin. Our results show that the addition of gut binding peptide sequences to Bt toxins

can enhance their activity against pests that are otherwise challenging to control.

Modification of a Bt toxin to target Asian citrus psyllid (Diaphorina citri, Hemiptera)

Bac12 2100



M. Teresa Fernandez-Luna¹, Michael Blackburn², David Hall³, Biviana Flores-Escobar^{1,4}, **Pavan Kumar**⁴, Bryony Bonning^{1,4}

¹ Department of Entomology, Iowa State University, Ames, IA; ² USDA - ARS, Beltsville, MD; ³ USDA - ARS, Ft. Pierce, FL; ⁴ Department of Entomology and Nematology, University of Florida, Gainesville, FL Corresponding author: bbonning@ufl.edu

The citrus industry is threatened by citrus greening, a disease caused by the bacterium Candidatus liberibacter asiaticus (CLas). CLas is vectored by the Asian citrus psyllid (ACP) and management of ACP has become a primary focus to limit disease spread. We previously demonstrated enhanced efficacy of a Bt toxin on addition of a peptide that binds to the gut of the target pest (N. Chougule et al., 2013, PNAS). Toward this end, Bacillus thuringiensis (Bt) isolates from the collection housed at USDA-ARS Beltsville, Maryland, were screened to identify strains with ACP toxicity. Five strains that show basal toxicity on feeding of adult ACP at 500 $\mu\text{g/ml}$ of proteolytically activated toxin were identified, and the trypsin proteolytic profile of the toxic strains characterized. Individual Cry toxins were identified by MS/MS for one of the toxic strains. One of these Cry toxins was selected for modification with ACP gut-binding peptide to enhance toxicity against ACP. ACP gut-binding peptides were isolated by feeding adult psyllids on a loop-constrained heptapeptide (Ph.D.-C7C) library. Sequences encoding four of the selected peptides were cloned for production of mCherry-peptide fusion proteins. A peptide selected based on relative binding strength under both in vivo and in vitro conditions, was used to engineer the selected Bt toxin. Following engineering, the stability of the modified toxin was evaluated by trypsin digestion and the impact of modification on toxicity against ACP assessed.

Patterns of gene expression and histopathological effects in western corn rootworm (Diabrotica virgifera virgifera) neonates, challenged with Cry 34/35 Ab1



Premchand Gandra¹, Kenneth E. Narva¹, Andrew J. Bowling¹, Heather E. Pence¹, Haichuan Wang², Blair Siegfried²

> ¹Dow AgroSciences, Indianapolis, Indiana, United States ²University of Nebraska, Lincoln, Nebraska, United States Corresponding author: PGandra@dow.com

Western corn rootworm (WCR, Diabrotica virgifera virgifera LeConte) is a major corn pest in the United States, causing annual losses of over \$1 billion. One approach to protect against crop loss by this insect is the use of transgenic corn hybrids expressing one or more crystal (Cry) proteins derived from Bacillus thuringiensis. Cry34Ab1 and Cry35Ab1 together comprise a binary insecticidal toxin with specific activity against WCR. Cry34/35Ab1 is a pore forming toxin, but the specific effects of Cry34/35Ab1 on WCR cells and tissues have not been fully characterized. In this presentation we integrate results from transcriptomics and histopathology for first instar WCR fed Cry34/35Ab1 proteins in diet based bioassays. High resolution histopathology revealed symptoms of intoxication for Cry34/35Ab1 including swelling and sloughing of enterocytes, constriction of midgut circular muscles, stem cell activation, and obstruction of the midgut lumen. With Next Generation Sequencing technologies, high-throughput RNA sequencing (RNA-seq) was conducted to examine gene expression in WCR neonates challenged with individual and combined Cry34Ab1 and Cry35Ab1 proteins. Results of three different statistical comparisons identified 114 and 1300 differentially expressed transcripts (DETs) in the Cry34Ab1 and Cry34/35Ab1 treatments, respectively, as compared to the control. The combined results of the histopathology and differential gene expression studies lay the groundwork for testable hypotheses aimed at extending the durability of Cry34/35Ab1 as an insect resistance trait.

The intracellular region of the Bombyx mori cadherin-like protein is not necessary to mediate cytotoxicity of Cry1A toxins

STU Bac14 •212•

Haruka Endo¹, **Satomi Adegawa**^{1,2}, Ryoichi Sato¹

¹Graduate School of Bio-Applications and Systems Engineering, Tokyo University of Agriculture and Technology, Koganei, Tokyo, Japan; ²Japan Society for the Promotion of Science, Research Fellowship for Young Scientists Corresponding author: s174092v@st.go.tuat.ac.jp

The cadherin-like protein in lepidopteran insects, known as a receptor for Bacillus thuringiensis Cry1A toxins, is a single-pass membrane protein and divided into extracellular and intracellular regions. The extracellular region has been shown to be important for toxin binding and oligomerization, but the role of the intracellular region is still obscure, although the signal transduction hypothesis suggested its role in Cry1A intoxication. Recently the cadherin-like protein is also paid attention in the synergistic effect with the ABC transporter C2 (ABCC2) in Cry1A intoxication, but the mechanism is poorly understood. In the present study, we generated two mutants of the Bombyx mori cadherin-like protein (BtR175): BtR175-DEL, which completely lacks the intracellular region, and BtR175-GPI, which is fused with the glycosylphosphatidylinositol (GPI)- anchor signal of B. mori aminopeptidase N1 to express as a GPI-anchored protein. As well as wild-type BtR175, both the mutants conferred cultured cells the susceptibility for Cry1Aa, Ab, and Ac toxins. Furthermore, when both mutants were co-expressed with B. mori ABCC2, swollen cells were observed in lower toxin concentrations, compared to cells expressing BmABCC2 alone and namely both mutants preserve the synergistic effect with ABCC2. These results indicate that the role of the cadherin-like protein as a Cry toxin receptor, even in the synergism with ABCC2, depends solely on its extracellular region.

Critical amino acids for the insecticidal activity of Vip3Af from Bacillus thuringiensis

Bac15 •213•



Núria Banyuls¹, C. Sara Hernández-Rodríguez¹, Jeroen Van Rie², **Juan Ferré**¹

¹ERi de Biotecnología y Biomedicina (BIOTECMED), Departament de Genètica, Universitat de València, 46100 Burjassot, España. ²Bayer CropScience N.V., Ghent, Belgium Corresponding author: juan.ferre@uv.es

Vip3 vegetative insecticidal proteins from Bacillus thuringiensis are an important tool for crop protection against caterpillar pests in IPM strategies. Despite already being implemented in Bt-crops, while there is wide consensus on their general mode of action, the details of their mode of action are not completely elucidated and their structure remains unknown. In this work the alanine scanning technique was performed on 558 out of the total of 788 amino acids of the Vip3Af1 protein. From the 558 residue substitutions, 19 impaired protein expression and 10 compromised the insecticidal activity against Spodoptera frugiperda. Substitutions that drastically reduced insecticidal activity mainly clustered in two regions of the protein sequence (amino acids 167-272 and amino acids 689-741). Most of the substitutions that impaired the activity to S. frugiperda behave likewise to Agrotis segetum, with few exceptions. The characterisation of the sensitivity to proteases of these 14 mutant proteins displaying decreased insecticidal activity revealed 6 different band patterns as evaluated by SDS-PAGE. The study of the intrinsic fluorescence of all selected mutants revealed only slight shifts in the emission peak, likely indicating only minor changes in the tertiary structure. An in silico modelled 3D structure of Vip3Af1 is proposed for the first time.

Construction of plasmid vector for dsRNA synthesis with Bacillus thuringiensis by using sporulation-dependent promoter





Min Gu Park¹, Jong Hoon Kim¹, Jae Young Choi², Seok-Hee Lee¹, Ying Fang¹, Dong Hwan Park¹, Ra Mi Woo¹, Bo Ram Lee¹, Woo Jin Kim¹, Yeon Ho Je^{1,2}

¹Department of Agricultural Biotechnology, Seoul National University, Seoul, Korea; 2Research Institute for Agriculture and Life Sciences, Seoul National University, Seoul, South Korea Corresponding author: btrus@snu.ac.kr

RNA interference (RNAi) is a post-transcriptional gene regulation mechanism found in virtually all plants and animals including insects. RNAi has been considered as an alternative strategy to control agricultural pests whereby double-stranded RNA triggers a potent and specific inhibition of its homologous mRNA. Since double-stranded RNAs are required for various RNAi applications, there is a need for cost-effective methods for producing large quantities of high-quality dsRNA. The gram-positive bacterium Bacillus thuringiensis has insecticidal proteins produced during the stationary/sporulation phase of growth with expression of its genes being driven by sporulation dependent promoters. To develop a dsRNA massproduction method utilizing B. thuringiensis, the pHT1K-EGFP plasmid vector, which transcribes sense and anti-sense EGFP under the control of cyt1Aa sporulation-dependent promoter with STAB-SD sequence was constructed, and the transcription level was measured by Illumina RNA-seq. The sequencing result was calculated by the Kallisto program to demonstrate that the transcription level of EGFP was approximately 30 and 4,000 times higher than that of the reference genes, mbl and rpoB, respectively. Furthermore, qPCR analysis results were in good accordance with RNA-seq results. These results demonstrated the potential of B. thuringiensis as a new platform for dsRNA production.

Calcium-independent cellular PLA₂ prevents lipid peroxidation, which is detrimental to development and immunity of the Indian meal moth, Plodia interpunctella

Bac17 •215•



Yonggyun Kim

Department of Plant Medicals, Andong National University, Andong 36729, Korea Corresponding author: hosanna@anu.ac.kr

Reactive oxygen species (ROS) induces oxidation against various biomolecules including fatty acids. Calcium-independent cellular phospholipase A₂ (iPLA₂) has been known to function maintenance of fatty acids in phospholipids in mammals. An insect iPLA₂ (Pi-iPLA₂) was predicted from transciptomes of the Indian meal moth, Plodia interpunctella. It encodes 835 amino acid sequence, which possesses 5 ankyrin repeats in N terminal domain and papatin lipase domain in C terminal region, respectively. It was expressed in all developmental stages from egg to adult. In larval stage, it was expressed in all tested tissues of hemocytes, fat body, gut and epidermis. RNA interference (RNAi) against Pi-iPLA₂ was performed with specific double-stranded RNA (dsRNA) with more than 70% efficiency in reduction of its expression. Under this RNAi condition, P. interpuctella exhibited developmental retardation with significant accumulation of lipid peroxidation measured by the amount of melondialdehyde. The RNAi of Pi-PLA₂ also impaired cellular immune responses of P. interpunctella, in which hemocyte nodule formation was significantly decreased. The hazard effect of lipid peroxidation was simulated with treatment of chlorine dioxide (ClO₂), which is an insecticidal agent by generating ROS in insect cell and P. interpunctella. CIO2 treatment increased lipid peroxidation in a dosedependent manner in Sf9 cell line. However, the addition of an antioxidant (vitamin E), reduced the formation of lipid peroxidation. CIO₂ treatment also increased lipid peroxidation in the larval fat body of P. interpunctella. Furthermore, the larvae treated with RNAi against Pi-PLA₂ were significantly susceptible to ClO₂ treatment. These results suggest that Pi-iPLA₂ plays a crucial role in remodeling fatty acid composition of phospholipids. It also suggests that chlorine dioxide treatment causes the lipid peroxidation via ROS, which explains its insecticidal activity.

Selection of Bacillus thuringiensis strains against Fusarium oxysporum f.sp. vasinfectum

Bac18 •216•



Elias Ferreira Sabiá Júnior^{1,2}, Sandro Coelho Linhares Montalvão^{1,2} Marcelo Tavares de Castro 1,2, Carlos Marcelo Soares3, Luiz Eduardo Bassay Blum¹, Rose Gomes Monnerat²

¹Instituto de Biologia, Universidade de Brasília; ²Embrapa Recursos Genéticos e Biotecnologia, Brasília, Brasil; ³Instituto Mato-grossense do Algodão, Mato Grosso, Brasil. Corresponding author: elias.fsabia@amail.com

Fusarium wilt caused by Fusarium oxysporum f.sp. vasinfectum on cotton is considered a major disease of this crop. The pathogen is difficult to control, having high persistence in the soil due to its capacity to parasitize other crops, being saprophyte and able to produce chlamydospores which enable it to survive in the soil for long time without presence of the main host. The control of this pathogen is based on the use of varieties with some degree of resistance, use of certified seed and the use of fungicides for seed treatment. Once detected in the field, plant protection is based on the use of chemicals. In this context, it is justified the development of alternatives for the management of this pathogen in the field. Biological control stands out as a viable alternative for the control of this pathogen. Bacillus are known bioregulators and natural antagonists of various pathogens. Thus, the objective of this study was to select toxic strains of Bacillus thuringiensis (Bt) to F. oxysporum and adjust an in vitro method for implementation and evaluation of the tests for selection. It was possible to determine the best culture medium for direct confrontation test, proposing a grading scale to facilitate the selection of the best strain with the paired test, identified among the strains selected in the confrontation assays, those that can produce volatile metabolites and non-volatile metabolites toxic to F. oxysporum and test these selected strains to the control of the disease during seed germination in greenhouse assays. With this last assay, it was noted a potential protective effect of some Bt strains during cotton seed germination.

Dynamic distribution and colonization of inoculated Bac19 •217 Bacillus thuringiensis on the tissues of Arabidopsis thaliana

Songqing Wu ^{1,2}, Bairong Lin², Yan Peng², Carballar-Lejarazú Rebeca³, Ivan Gelbič⁴, Lei Xu², Xiong Guan², Lingling Zhang²

¹College of Forestry, Fujian Agriculture and Forestry University, 350002
Fuzhou, People's Republic of China; ²Key Laboratory of Biopesticide
and Chemical Biology, Ministry of Education, Fujian Agriculture
and Forestry University, 350002 Fuzhou, Fujian, People's Republic of
China; ³Department of Molecular Biology and Biochemistry, University of
California Irvine, Irvine, CA, 92697, USA; ⁴Biology Centre of the Czech
Academy of Sciences, Institute of Entomology, Branišovská 31, 37005
České Budějovice, Czech Republic

Corresponding author: lingling00264@163.com

The dynamic distribution and colonization of *Bacillus thuringiensis* (Bt) in *Arabidopsis thaliana* (L.) tissues was studied using the transformed Bt HD73^{Cry—}GFP strain which expresses the green fluorescent protein (GFP) as a marker. After HD73^{Cry—}GFP inoculation, the distribution of the labeled Bt was recorded both internally and externally over the entire plant, including the phyllospere and rhizoshere using fluorescence and a confocal microscopy. The Bt HD73^{Cry—}GFP strain was able to stably colonize both internally and externally the roots, stems and leaves of *A. thaliana* under gnotobiotic and non-UV conditions. This work provides fundamental ecological information on the dynamic distribution, both internally and externally, of Bt HD73^{Cry—}GFP strain on *A. thaliana* tissues, our work provides basic information for a better understanding of Bt-plant interactions.

Promotion of plant growth by Bacillus thuringiensis in Brazil Bac20 •218•

Flávia S. S. Santana¹, Carlos M. Soares², Rose G. Monnerat³

¹Faculdade de Agronomia e Medicina Veterinária, Universidade de Brasília, Brasília, Brasil; ²Instituto Mato-grossense do Algodão, Mato Grosso, Brasil; ³Prédio de Controle Biológico, Embrapa Recursos Genéticos e Biotecnologia, Brasília, Brasil *Corresponding author: flaviasantana_agr@hotmail.com*

The use of *Bacillus thuringiensis* (Bt) in pest control is already well known. This bacterium also has other important characteristics, such as the ability to promote plant growth, which should be further investigated. For this was used a Bt strain toxic to lepidopteran insects. The effect of the seeds being immersed in bacterial suspension (with and without chemical treatment) of 10 cotton genotypes was evaluated. The seeds were immersed in bacterial suspension with 10⁸ CFU (spores) per mL and then seeded in trays for seedling production. The plant emergence velocity was evaluated and on the 15th days of the sowing, four genotypes (IMA 6035, IMA 8405 and IMA 5675 (not chemically treated) and IMA 2106 (chemically treated)) were harvested due to their most vigorous appearance. The height, root length, fresh and dry matter of the whole plant were recorded. The results indicate that seeds being immersed in bacterial suspension, with and without chemical

treatment, can affect the emergence speed of the plants and that the IMA 6035 genotype with the seeds imposed under these conditions of bacterial suspension immersion presented the higher results for height (8%) and for plant dry matter (13%) compared with the treatment without the bacteria. In a second moment, the effect of the methods of inoculation of Bt was evaluated. For this assay, the IMA 1318 genotype and three inoculation methods were used: (i) Seeds being immersed in bacterial suspension with 10⁸ CFU per mL (BS), (ii) BS (method (i)) + weekly applications of 1 mL of the bacterial suspension around the roots of the plants (with 10' CFU per mL), (iii) BS + addition of the bacterial pellet to the soil (with 10⁷ CFU per mg). The evaluation occurred at the 35th days of the planting. The results showed that there was an increase in root dry matter yield provided by the inoculation methods for the analyzed genotype. A higher number of cells were observed inside the roots in the treatment (ii). For the IMA 1318 genotype, the root dry matter production increased regardless of the inoculation method of the bacteria.

The Caenorhabditis elegans CUB-like-domain containing protein F35E12.10 functions as a receptor for Bacillus thuringiensis Cry6Aa toxin

Bac21 •219•



Jianwei Shi, Donghai Peng, Fengjuan Zhang, Lifang Ruan, Ming Sun

State Key Laboratory of Agricultural Microbiology, College of Life Science and Technology, Huazhong Agricultural University, Wuhan 430070, China Corresponong author: m98sun@mail.hzau.edu.cn.

Plant-parasitic nematodes caused huge economic losses to agriculture per year, and nematicidal activity was exhibited in two major families of Bacillus thuringiensis crystal proteins, Cry5 and Cry6. Previous work showed that binding receptors were critical factors involved in processes of crystal protein intoxication. Now, the receptors of crystal protein Cry5Ba were identified as glycolipids in Caenorhabditis elegans. However, the receptors of Cry6 remain unknown. In this study, the CUB-like-domain containing protein F35E12.10, from C. elegans proteins released by phosphatidylinositol-specific phospholipase C (PI-PLC), was identified as a binding protein for Cry6Aa by affinity chromatography. F35E12.10 contained a predicted glycosylphosphatidylinositol (GPI) anchor site and was reported to locate in lipid rafts of intestinal cell surface. Western ligand blot and ELISA verified the binding interaction between Cry6Aa and F35E12.10 with high affinity and specificity. Bioassays showed that gene f35e12.10 was implicated in Cry6Aa intoxication. Furthermore, f35e12.10 mediated Cry6Aa uptake in C. elegans gut cells, suggesting that f35e12.10 was required for interaction of Cry6Aa with intestinal cells. Finally, we observed that the mutation of gene f35e12.10 did not decrease susceptibility to Cry5Ba in C. elegans. These results demonstrated that F35E12.10 is a functional receptor for Cry6Aa protein. To our knowledge, this is the first report about Cry6 protein binding receptors in C. elegans.

Physiological and histopathological effects of new proteins toxic to southern green stink bug (*Nezara viridula*)

Bac22 •220•

Theodore W. Kahn, Mandy Bush, Sara Lenzen, Jessica Monserrate. Alberto Bressan

Bayer, Crop Science Division, Morrisville, NC, USA

Corresponding author: ted.kahn@bayer.com

A number of Hemipteran insect species, including stink bugs, are pests of major crops such as cotton and soybean. Stink bugs feed by extra-oral digestion. They pierce plant tissues, inject digestive enzymes, and suck nutrients from the tissues. New traits are required that will be capable of controlling Hemipterans when expressed in crop plants. Screening of diverse microbial strains by the Crop Science Division at Bayer has led to the identification of proteins that are toxic to the southern green stink bug. To begin to elucidate the mode of action of the proteins, insects were fed purified proteins, and their digestive systems were dissected and examined microscopically. Morphological and histological changes in different compartments of the digestive systems were observed, giving an indication of the location where the toxic effect is exerted.

Mining microbial metagenomes for novel insecticidal proteins

Poster - Bac23

The purification and mechanism analysis of Holotrichia oblita toxic Vip1/Vip2 binary protein Poster – Bac25

I.S. Shilova¹, A.J. Johnson¹, L. Chan¹, M. David¹, I.W. Davis², J.A. Haas², S. Jain¹, S. Iwai¹, P. Loriaux¹, P. Ramachandran¹, E. Rutherford¹, K.M. Wegener², T. Weinmaier¹, R.J. Williams², Y. Wu¹, T.Z. DeSantis¹, K.A. Bennett¹

¹Second Genome, Inc, 341 Allerton Ave, South San Francisco, CA 94080, USA ²Monsanto, 700 Chesterfield Parkway West, Chesterfield, MO 63017, USA Corresponding author: irina@secondgenome.com

Microbes are genetically highly diverse and have evolved a variety of strategies in order to compete for nutrients, including pathogenesis of insects. Insecticidal microbes and their toxin genes have been explored for agricultural uses for decades. Traditional methods to discover insecticidal bacteria and toxin genes are based on isolating bacteria on selective media, focusing primarily on the spore-forming bacteria Bacillus thuringiensis. Thus, the diversity and novelty of isolated bacteria is limited by what can form colonies on the selective media. An alternative approach to identifying toxin genes is based on nucleotide homology searches of all available sequenced genomes. However, most of the bacteria from soil microbial communities have not been cultured, and their genomes have not been sequenced. Our approach to identify novel insect toxins is based on mining soil metagenomes, where the genetic potential of the whole microbial community includes uncultured species. In addition, we use multiple enrichment strategies on natural microbial communities to increase the abundance of insecticidal bacteria. The metagenomes from the enriched communities are then assessed using the Second Genome discovery platform which includes statistical models trained on known insect toxins. The output from this analysis is a set of potentially novel insecticidal proteins that can be tested in vivo.

The analysis of *B.thuringiensis* tolerance to insect hemolymph and its effect on infection efficiency

Poster - Bac24

Ying Huang, Kui Wang, Changlong Shu, Jie Zhang *

State Key Laboratory for Biology of Plant Diseases and Insect , Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing, 100193, P.R. China

Corresponding author: jzhang@ippcaas.cn

B.thuringiensis (Bt) is an impotent insect pathogenic bacteria and it infects insects by breaking the intestine and multiplication in the living host's hemolymph. Because its high and specific larvicidal activity, Bt is currently used as an insecticide. When Bt spore and crystal mixture was ingested by susceptible insects, the protoxins solubilized and activated proteolytically in the host midgut. The activated protein specific bind to midgut receptors, forming pores in the epithelial midgut and break down the epithelial cells. Then the Bt spores or germinated cells were go through the hole into host hemolymph. The Bt spores or cells multiplication in the hemolymph caused septicaemia and kill the insects. Although the Bt are armed with immune inhibitor A (InhA) metalloprotease, the insects had evolved strong immune system to combat with the bacteria infection. The ability of Bt break through immune system and multiply in hemolymph was important for Bt toxicity. However, there are still lack of data on compare Bt multiply ability in insect's hemolymph directly. In this report, we assayed the hemolymph (from Protaetia brevitarsis, removed cells) effects on Bt cells. The strain HBF-18 shown sensitive to hemolymph (influence rate: 0.06) and the strain G03 shown resistance to hemolymph (influence rate: 1.52) were selected for further analysis. The inject infection assay were performed on third instar larvae of Protaetia brevitarsis. The data shows, G03 have higher lethality than HBF-18 in lower injection dose while HBF-18 have stronger toxicity than G03 in higher injection dose. A time course of hemolymph effects on HBF-18 and G03 were subsequence performed. The data shows, most of the HBF-18 strain cells were died in earlier period (first two hour) but it quickly multiplies in later period (after six hour). Different from HBF-18, G03 shows resistance to hemolymph in earlier period, and have lower multiplies speed later period. In conclusion, we illustrated, although few strain sensitive to insect hemolymph, most strain were resistance to insect hemolymph and the distribution of hemolymph infect rate on Bt was approximately normal distribution. People can screen for high resistance Bt strain for pesticides development.

Jian Jiang, Changlong Shu, **Jie Zhang**

State Key Laboratory of Biology for Plant Diseases and Insect Pests, Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing 100193, P. R. China Correspondence: izhanq@ippcaas.cn

White grubs (larvae of Scarabaeidaes family) are serious agricultural pests worldwide. In China, Holotrichia oblita is one of main white grub pests and causing significant reductions of yield and great economic losses. White $% \left(1\right) =\left(1\right) \left(grubs are difficult to control, principally because of its soil living habits. In previously work, Bacillus thuringiensis (Bt) strain HBF-18 (CGMCC2070), containing two Bt genes (vip1Ad1 and vip2Ag1), has been confirmed toxic to H. oblita larvae. The Vip1Ad1 and Vip2Ag1 protein is a binary toxin toxic to H. oblita larvae, and its toxicity is significant higher than those of Cry 8 proteins. The study of the mechanism of action of this Vip1Ad1/Vip2Ag1 binary toxin is important for their utilization in the control of H. oblita. In this study, we investigated the methods of extraction and purification of Vip1Ad1 and Vip2Ag1. The results showed that Vip1Ad1 and Vip2Ag1 are secretory proteins, and its inefficiency to extract these two proteins using ammonium sulfate precipitation method. But we have obtained purified Vip1Ad1 and Vip2Ag1 from fermentation broth after treated with Cross-Flow and then purified by ion-exchange and anion-exchange, respectively. Furthermore, we investigated the activation process of these two proteins. The results indicated that Vip1Ad1 (approximately 90 kDa) digested with Trypsin resulted in a protease-resistant core fragment of approximately 70 kDa (Vip1Ad1-T), but Vip2Ag1 could not be digested by both Trypsin and Chymotrypsin. What's more, we analyzed the binding characteristics of Vip1Ad1/Vip2Ag1 with brush border membrane vesicles (BBMV) in H. oblita larvae. The results showed that the protoxin of Vip1Ad1 and Vip2Ag1 could not binding with each other in solution; And, the Vip1Ad1, either protoxin or trypsinized, bound to BBMV from H. oblita, whereas the Vip2Ag1 alone did not; The binding of Vip2Ag1 was determined only when the BBMV were reacted with trypsinized Vip1Ad1, but not when they were reacted with Vip1Ad1 protoxin; In particular, the Vip2Ag1 bound to Vip1Ad1 oligomers-BBMV complex and the oligomers could generated both in solution and on the cell surface after Vip1Ad1 monomers bound with BBMV, but in either scenario, the Vip1Ad1 oligomers were generated only after proteolysis of Vip1Ad1 protoxin; And, the results of saturation binding assays demonstrated that Vip1Ad1 bound specifically to binding sites on the BBMV from H. oblita larvae. In addition, we identified Vip1Ad1-binding proteins in the midgut of *H. oblita* larvae. The results demonstrated that the Vip1Ad1 binding protein is not cholesterol and not GPI-anchored protein. Our work promotes the understanding of mechanism of Vip1/Vip2 binary toxin against H. oblita larvae.

A novel Type Six Secretion System in the entomopathogenic bacterium *Xenorhabdus* bovienii may impact insect virulence and bacterial competition

STU Bac26

Christine Bradshaw¹, Rebecca McQuade¹, S. Patricia Stock^{1, 2, 3}

¹School of Animal Comparative Biomedical Sciences, University of Arizona, ²Center for Insect Science, ³Department of Entomology, University of Arizona Corresponding author: christineb@email.arizona.edu

Xenorhabdus bovienii is a Gram-negative bacterium that exhibits a dual lifestyle, behaving as a mutualist to at least Steinernema nematode species and as a pathogen to insects. X. bovienii displays a type VI secretion system (T6SS) in its genome that has yet to be studied. The T6SS is a one-step mechanism widely used by Gram-negative bacteria to transport effector proteins into target cells. T6SSs have been shown to play a key role in pathogenesis of host cells and in the competition with other bacteria. In this study, we used a homologous recombination approach to generate a X. bovienii mutant lacking hcp, a crucial structural gene of the T6SS, then conducted bioassays to assess the role of this secretion system. Preliminary results show a decrease in melanization and mortality in insects infected with the T6SS mutant compared to those infected with wild-type X. bovienii, suggesting a decrease in virulence in the mutant strain. Additionally, In vitro bacterial competition assays resulted in the wild-type strain outcompeting

the mutant strain. We speculate the T6SS may play a role in bacterial competition and insect virulence, which may contribute to the success of the mutualistic relationship between X. bovienii and Steinernema nematodes.

Evaluation of Galleria mellonella (Lepidoptera: Pyralidae) as a complementary model organism to study enteropathogenic E. coli

STU Bac27

Isabel Forlastro¹, Rebecca McQuade², V. K. Viswanathan¹, S. Patricia Stock^{1,2,3}

¹Department of Microbiology, School of Animal and Comparative Biomedical Sciences, ²Center for Insect Science, ³Department of Entomology, The University of Arizona

Corresponding author: iforlastro@email.arizona.edu

Enteropathogenic Escherichia coli (EPEC) is a common diarrheal pathogen that affects approximately 0.8 million children per year in developing countries. EPEC uses a complex bacterial structure called the Type III Secretion System (T3SS) to inject bacterial effector proteins directly into intestinal epithelial cells, manipulating the cells' behavior and causing disease. Rabbits are a commonly used model to study EPEC. However, rabbits have several limitations as hosts including expense, specialized handling, ethical concerns and federal regulations regarding the use of laboratory vertebrates. Insects are potential complementary models for studying bacterial pathogenesis in vivo. Benefits of using insect hosts include low-cost, ability for mass infections, and analogous immune strategies to vertebrates. We hypothesize that larvae of the greater wax moth Galleria mellonella (GM) can serve as an effective model in studying the EPEC T3SS. In order to determine if EPEC can infect GM we injected EPEC into the body cavity (hemocoel) of the insects. We observed that live wild-type EPEC kills GM and that heat-killed EPEC does not kill, indicating that EPEC actively infects GM. To explore the mechanism of killing, we also considered EPEC mutant strains that lack different components of the T3SS. These mutants were defective in killing GM, indicating that EPEC kills these insects in a T3SSdependent manner. We are currently adapting this model to test potential T3SS inhibitors and to study individual T3SS effectors. We are also attempting to model oral infections in insects to more closely represent human infection.

MICROSPORIDIA DIVISION POSTERS

The role of the decapping enzyme EOL-1 in C. elegans intracellular infection response

Msp01 •221•

Jessica N. Sowa¹, Emily R. Troemel¹

Division of Biological Sciences, University of California at San Diego, La Jolla. Corresponding author: jsowa@ucsd.edu

Microsporidia comprise a phylum of obligate intracellular parasites most closely related to fungi. Although they infect a wide range of animal species and can cause lethal infections in immunocompromised patients, relatively little is known about the mechanisms of host defense against microsporidia. Nematocida parisii is a species of microsporidia that is a natural pathogen of C. elegans. N. parisii infects the C. elegans intestine, causing fusion of intestinal cells and ultimately lethality (Troemel et al, PLoS Biology 2008; Balla et al, Nature Microbiology 2016). Our lab has profiled the C. elegans transcriptional response to N. parisii (Bakowski et al, PLoS Pathogens 2014), with the goal of identifying features of the host response to microsporidia infection. One gene found to be induced by N. parisii infection was eol-1, a de-capping and exonuclease enzyme that has been reported to be involved in regulating pathogen-induced aversive olfactory learning (Shen et al, J. Neuroscience 2014). Our RNAseq data shows that eol-1 is strongly upregulated during N. parisii infection, beginning early at 8 hours post infection. In uninfected worms, we find that EOL-1 is expressed primarily in the spermatheca and URX neurons based on analysis of EOL-1 transcriptional and translational fusions. In addition, we found that EOL-1 expression is also induced in response to infection by the Orsay virus, another natural intracellular pathogen of C. elegans (Félix et al, PLoS Biol. 2011). Interestingly we find that in worms exposed to N. parisii or Orsay virus, eol-1 expression is induced in the intestine, which is the tissue targeted by both pathogens. Our

preliminary results from analysis of eol-1 mutants suggest that eol-1 plays a role in host response to intracellular infections.

Secretion of the peritrophic matrix in the honey bee, Apis mellifera, midgut is impaired by Nosema ceranae infection

Msp02 •222•

Thomas C. Webster, Martin Matisoff, Katherine Kamminga, Cecil Butler

College of Agriculture, Food Science and Sustainable Systems, Kentucky State University, Frankfort KY, USA Corresponding author: thomas.webster@kvsu.edu

The honey bee (Apis mellifera) peritrophic matrix (PM) degrades after the bee's midgut becomes infected with the microsporidian Nosema ceranae. Consequently the PM becomes less functional in its roles as a substrate for digestive enzymes, as a barrier against ingested pathogens, and as a structure to move pollen grains through the midgut. We were able to observe PM structure and integrity by removing the midgut of a bee and clearing it in 10% KOH. The high chitin content of the PM provides it with a strong affinity for the fluorescent stain calcofluor white. Pollen grains also stain well with calcofluor white. By this method we could easily observe the structure of the PM, and its retention of pollen grains. We found that N. ceranae spores do not damage the PM directly, by releasing their polar tubes. Instead, PM secretion by midgut epithelial cells is impaired when these cells are infected.

The abundance and distribution of Nosema ceranae in honeybees in Mo'orea and Tahiti, French Polynesia. Msp03 •223•

Julie V. Hopper

University of California, Davis, One Shields Ave., Davis, California 95616 USA Corresponding author: jvhopper@ucdavis.edu

The global distribution and extent of infection by Nosema ceranae and N. apis in the European honeybee, Apis mellifera, is of increasing interest due to the widespread concern over the current decline of honeybees. Although Nosemosis has been found in A. mellifera across the globe, this disease has not yet been investigated in French Polynesia, where honey production, pollinator services, and the honeybee trade contribute substantially to the local economy. From October to November 2013, I collected 20-40 bees (A. mellifera) per hive from 21 hives across 14 locations around Mo'orea and Tahiti, French Polynesia. The midgut was excised from each bee and rinsed in DI water prior to examination under phase contrast microscopy at 400 magnification. All tools were sterilized with 5% bleach before dissection of each bee. Infection levels were categorized by spore intensity and remaining midgut tissues of each bee were stored in 0.01M EDTA at -20°C for DNA extraction. DNA was extracted from at least two infected bees from each hive using a modified Chelex method. I used a previously published duplex-PCR protocol based on species–specific primers for N. ceranae and N. apis to amplify the RNA polymerase II large subunit (RPB1). Visualization of the fragments on a 1.2% electrophoresis gel determined the presence of N. ceranae in all of the infected bees that were examined. Prevalence of infection with N. ceranae within hives ranged from 3% to 77% in Mo'orea and from 10% to 33% in Tahiti. Results were reported to the Department of Agriculture in Mo'orea, as well as to the local apiculturists in Mo'orea and

Anti-Nosema activity of entomopathogenic fungi cultural filtrates in honeybee nosemosis STU Msp04 •224•

Jiln Ma, DongJun Kim, InHui Kim, WonSeok Gwak, HwiGeon Yun, JongMin Oh, JiHoon Lee, SooDong Woo

Department of Agricultural Biology, College of Agriculture, Life & Environment Science, Chungbuk National University, Cheongju 28644, Korea Corresponding author: sdwoo@cbnu.ac.kr

The population of managed honeybees has been dramatically declining the recent past in the worldwide. The one of most common disease of honeybees is nosemosis which is caused by Nosema ceranae. Nosema ceranae causes significant detriment to honey production and results in economic losses critically. In our knowledge, Fumagillin is the only antibiotic approved for control of nosemosis in honeybees. However, this antibiotic may have unintended effects on the honey bee host, ultimately contributing to increased prevalence and pathogenicity of N. ceranae. To select fungal isolates with anti-Nosema activity against N. ceranae, in this study, entomopathogenic fungi cultural filtrates were prepared from 342 fungal isolates, and anti-Nosema activities were evaluated by in vitro polar tube germination assay method. As a result, six isolates, Paecilomyces marquandii 364, Metarhizium anisopliae 296, Beauveria brongniartii 183, B. bassiana 35 and 59, Pochonia sp. 60 were finally selected. These fungi cultural filtrates had bactericidal activity against N. ceranae. Safety and inhibition of nosemosis were further evaluated in honeybees. Paecilomyces marquandii 364 and Pochonia sp. 60 incrreased the survival rate of honeybees and decreased the production of N. cerange spores. Fungi cultural filtrates of selected isolates have a good effect on inhibiting the nosemosis of honey

Serendipitous Finding: Wild populations of Spruce Budworm are infected with both Nosema fumiferanae and a Cystosporogenes species Msp05 •225•



George Kyei-Poku , Debbie Gauthier

Canadian Forestry Service, Great Lakes Forestry Centre. 1219 Queen Street East, Sault Ste. Marie, Ontario, Canada P6A 2E5 Corresponding author: aeorae.kvei-poku@canada.ca

The most ubiquitous natural enemy found in close association with spruce budworm, Choristoneura fumiferana populations is the microsporidian Nosema fumiferanae. However during a recent survey, a Cystosporogenes species was isolated for the first time from wild spruce budworm populations. Detecting and distinguishing these natural enemies in 2nd stage *C. fumiferana* larvae by light microscopy to determine the health of the population for onward forest protection decisions is not feasible on an operational basis. We therefore developed a much more sensitive molecular platform, a multiplex real-time loop-mediated isothermal amplification (m-LAMP) assay to synchronously amplify, detect and distinguish N. fumiferanae and the Cystosporogenes spp. DNA in a single reaction using crude DNA without conventional extraction. The optimum amplification occurred within 30 min at 64 °C, and the amplified products were distinguished based on the different melting temperatures (Tm values) of the two specifically m-LAMP amplified products. Alternatively, the amplified products were detected by fluorescence, lateral flow dipstick and also confirmed by gel electrophoresis. The detection limit of (LOD) of the m-LAMP assay was determined to be 100 pg/µl and similar to that of multiplex PCR. The specificity of the m-LAMP assay was evaluated using 16 known microsporidia strains in the genera, Nosema, Vairimorpha, Cystosporogenes and Endoreticulatus. The m-LAMP showed 100% inclusivity and exclusivity. Additionally, the developed m-LAMP assay was validated using DNA extracts from spores recovered from pooled C. fumiferana larvae. In all cases, Nosema fumiferanae and the Cystosporogenes spp. were detected in pooled spores isolated from three different geographical populations of C. fumiferana. The developed m-LAMP assay is simple, rapid, sensitive, specific and suitable for direct detection of N. fumiferanae and a Cystosporogenes spp. in field populations of C. fumiferana without complex DNA extraction. Hence it has the capability to be used for on-site disease diagnosis and field surveys. Most importantly in order to avoid predetermination of a microsporidian species and resultant inaccurate data (confounded effects) generated in a bioassay when using spores isolated from pooled insects: it will be prudent to implement a similar assay to delineate microsporidia species present in pooled spores prior to bioassays.

Application of RNAi to the control of microsporidian parasite Nosema ceranae in honeybee (Apis mellifera)

Msp06

WonSeok Gwak¹, InHui Kim¹, JiIn Ma¹, JongMin Oh¹, Dong Jun Kim, HwiGeon Yun¹, JiHoon Lee¹ SooDong Woo¹

¹Department of Agricultural Biology, College of Agriculture, Life & Environment Science, Chungbuk National University, Cheongju 28644, Korea Corresponding author: sdwoo@cbnu.ac.kr

Honeybees are one of the most important insects in the world as a pollinator of numerous agricultural crops. Recently, however, honeybees have faced many diseases which threaten bee colony including a serious population decline phenomenon called CCD (colony collapse disruption). Nosema

ceranae is a pathogen causing nosemosis, which is one of the most common diseases in honeybees. Microsporidian N. ceranae is an obligated intracellular parasites and depends on its host for energy and metabolic needs. Here we selected the several genes from mitosome of N. ceranae to develop RNAi for the control of Nosema species. Especially, TOM40 (translocase out membrane), FNR (Ferredoxin NADPH reductase) 1 and 2, Nar1 genes were chosen and their dsRNA were synthesized because their amino acid sequences show less homologous with those of the honeybees (Apis mellifera). After infection of N. ceranae, the honeybees were treated with dsRNA either by using single or combining two or more. The infection rate and specific gene silencing of Nosema were than analyzed. The targeted FNR1 and FNR2 genes were affected highly by dsRNAs, resulting in low productivity of Nosema spores. In addition, dsRNA FNR1 and FNR2 enhanced honeybee survival rate. To increase the effect of gene silencing, the effect of dsRNA complex particles with chitosan was investigated. Treatment of chitosan to dsRNA enhanced the effects for reducing spore productivity and bee survival rate more than the naked dsRNA treatment. To verify the function of FNR genes, additionally, the recombinant FNR proteins were overexpressed in insect cells using baculovirus expression system. The predicted FNR activity was confirmed through Cytochrome C oxidase activity assay, and the overexpression of the FNR increased the cell viability.

FUNGI DIVISION POSTERS

Studying the interaction among fungi involved in the multiple biological control of leaf-cutter ants



Daniela Goffré, Ema Cavallo, Patricia J. Folgarait

Laboratorio de Hormigas, Departamento de Ciencia y Tecnología. Universidad Nacional de Quilmes, Bernal, Argentina. Corresponding author: patricia.folgarait@gmail.com

The use of multiple biological control agents simultaneously requires that the different agents do not affect negatively each other. With the aim to test this hypothesis with microorganisms used against leaf-cutter ants, we set up laboratory assays to evaluate the in vitro interactions among an entomopathogen, a mycopathogen and the fungi cultivar of leaf cutter ants. We first tested the two biocontrol agents against each other; i.e. the entomopathogen Beauveria bassiana with the mycopathogen Trichoderma lentiforme. Second, we tested the effect of B. bassiana on one experiment and of *T. lentiforme* on another, against the ants' cultivar, *Leucoagaricus* sp. We measured their growth rate through time on PDA in co-ocurrence as well as alone (controls). We found no effect of the entomopathogen over Leucoagaricus sp. as well as of the Leucoagaricus sp. over the mycopathogen. We also found a negative effect of the mycopathogen over Leucoagaricus sp. which encourages the use of T. lentiforme as a potential biological control agent, as well as over the entomopathogen, in comparison to the controls. The second result warns regarding the simultaneous use of both controllers. The use of conidia in the application of these two agents could prevent the negative effect of the mycopathogen over the entomopathogen due to the fact that the germination should occur over different substrates in nature.

The importance of studying preservation methods of microorganisms used in the biological control of ants Fun03 •228•

Daniela Goffré, Lucas A. Martínez, Nicolás F. Lucero, **Patricia J. Folgarait**

Laboratorio de Hormigas, Departamento de Ciencia y Tecnología. Universidad Nacional de Quilmes, Bernal, Argentina. $Corresponding\ author:\ patricia. folgarait@gmail.com$

Biological control methods rely on the use of live organisms against target pests. There are different types of biological control strategies; however with social insects such as ants, which formed supernumerary colonies with a reproductive queen highly protected, it is necessary to apply the biological control agents several times over long periods in the field. To fulfill this need appropriate methods for conserving the biological control agents are needed. In our case we studied common used laboratory methods to conserve fungi such as keeping cultivars in Petri dishes in the fridge at 4°C as well as storage of conidia in ultrafreezers at -80°C. Our biological control agents involved the

entomopatho-gens Beauveria bassiana and Purpureocillium lilacinum and the micopathogen Trichoderma lentiforme (ex T. harzianum complex). We grew these fungi on PDA media in order to obtain their conidia and prepare suspensions of known concentrations which were used, either for inoculating ants in the case of entomopathogens, or for doing Petri dishes challenges with the mycopathogen against Leucoagaricus sp., the leaf-cutter ants' cultivar. We found that both methods were effective in maintaining their virulence for all the fungi except for P. lilacinum which conidia have lost its virulence using both preservation methods. Due to this result, we further evaluate the virulence when P. lilacinum was stored using a layer of mineral oil on top of PDA slant culture tubes kept in the fridge. Again, we did not detect virulence when expected although we observed an increased mortality at the end of the 15 days trial, and we recovered few ants with this fungus. Therefore, we have shown in this study that B. bassiana and T. lentiforme can be stored in ultrafreezers for at least 2 years and in the fridge for 6 months without losing their pathogenicity whereas for P. lilacinum further storage methods need to be tested to maintain its virulence, and we suggest that several passages through the host might be necessary in order to keep its virulence.

Intensive studies on the biodiversity of fungal entomopathogens of mosquitoes and other dipteran vectors of major human and animal diseases in central Brazil

Richard A. Humber¹, Christian Luz², Cristian Montalva^{2,3}, Manuel E. Rueda-Páramo^{2,4}

¹Emerging Pests and Pathogens Research, Robert W. Holley Center for Agriculture & Health, Ithaca, New York, USA: ²Instituto de Patologia Tropical e Saúde Pública, Universidade Federal de Goiás, Goiânia, GO, Brazil: ³Facultad de Ciencias, Instituto de Bioquímica y Microbiología, Casilla 167, Universidad Austral de Chile, Valdivia, Chile; 4CEPAVE-CONICET-UNLP, La Plata, Buenos Aires, Argentina Corresponding authors: richard.humber@ars.usda.gov; wchrisluz@hotmail.com

This poster summarizes three years of work (2014–2017) in the central Brazilian states of Goiás and Tocantins to explore, to identify, to isolate and preserve (whenever possible), and to characterize the biodiversity of fungal entomo-pathogens (EPFs) affecting dipteran vectors of serious human and animal diseases. Not unexpectedly, the major emphasis was on pathogens affecting mosquitoes, but fungi affecting other arthropods were also collected from all sites visited. Major surprises among the results of these studies were that Leptolegnia chapmanii was-while little collected in the US and once in Argentina before now-occurred widely and commonly in central Brazil but without obviously producing its sexual stages on those Brazilian sites. Collections of Pandora bullata from calliphorid flies in a mountainous area of northern Goiás were extensive but this fungus, too, was found to be extremely reluctant to produce its resting spores that formed readily in previously known North American collections of this species. This poster will present the key highlights of this project including new species of fungi and oomycetes, and larvae (maggots) of Musca domestica were confirmed to be suspectible in laboratory studies to, and a new hypothesis about possible biogeographical effects on the types of spores produced by fungal entomopathogens, and a listing of the isolates of EPFs obtained during this study. This project was funded as CAPES Science Without Borders project PVE 71/2013, 149988.

STU Fun05 •230• Activity of Culicinomyces spp. (Hypocreales: Cordycipitaceae) against Aedes aegypti eggs, larvae and adults

Juscelino Rodrigues¹, Richard A. Humber², Éverton K. K. Fernandes¹, Christian Luz¹

¹Laboratório de Patologia de Invertebrados, Instituto de Patologia Tropical e Saúde Pública, Universidade Federal de Goiás, Goiânia, GO, Brasil; ³USDA-ARS Robert W Holley Center for Agriculture & Health, Ithaca, NY, USA. Corresponding author: juscelinorf@hotmail.com;

Culicinomyces spp., that infect and kill aquatic nematoceran larvae after ingestion of conidia, need more attention in the effort to develop a mycoinsecti-cide against mosquito vectors of arboviruses, especially Aedes aegypti. In this study, larvicidal activity against A. aegypti of the majority of available C. clavi-sporus (ARSEF 372, 582, 644, 706, 964, 1260, 2478, 2479, 2480) and C. bisporalis isolates was tested. Ovicidal and adulticidal activities of the most promising isolates, ARSEF 644, 964 and 2479, were also investigated in A. aegypti. Third instar larvae treated with ARSEF 372, 644, 964 or 2479 at 10⁶ conidia/ml died most rapidly (LT₅₀ 1–1.3 days and LT₉₀ 2.6-2.8 days). Values of LC₅₀ and LC₉₀ of these isolates varied between $2.1x10^5 - 3.6x10^5$ conidia/ml and $1.9x10^6 - 2.5x10^6$ conidia/ml, respectively. However, none of the selected isolates had any activity against eggs when treated directly with conidia (5x10⁶ conidia/ml). Adults, which were fed with 10% sucrose solution added with 10⁶ conidia/ml, were slightly susceptible to oral infection (25-46% mortality 10 days after feeding and 13% in the control without conidia). Results, made clear that highly virulent *C. clavi-sporus* isolates with potential for the control of A. aegypti larvae are available, and these isolates should be considered for the development of biological control of this important vector.

Adaptation of a simple method to detect entomopathogenic fungi from mosquito larvae

Fun06 02310

Caroline Bergamini¹, **Cristian Montalva**^{1,2}, Juscelino Rodrigues¹, Richard A. Humber³, Christian Luz¹

Laboratório de Patologia de Invertebrados, Instituto de Patologia Tropical e Saúde Pública, Universidade Federal de Goiás, Goiânia, GO, Brasil: ²Facultad de Ciencias, Instituto de Bioquímica y Microbiología, Casilla 567, Universidad Austral de Chile, Valdivia, Chile; ³USDA-ARS Robert W Holley Center for Agriculture and Health, Ithaca, NY, USA. $Corresponding\ authors:\ montal va.cristian @gmail.com;\ wchrisluz @hotmail.com$

Aquatic immature mosquito stages, whether alive or dead, are delicate and not easy to handle in the field for monitoring fungal infections. The effectiveness of a new, simple technique to detect entomopathogenic fungi directly from larvae was checked for four fungal species with known larvicidal activity in A. aegypti. Larvae were exposed for 24 h to conidia of Beauveria bassiana, Culicinomyces clavi-sporus, Metarhizium anisopliae or Tolypocladium cylindrosporum and then set in Petri dishes on a soft water agar medium (WA) with chloramphenicol, thiabendazole, crystal violet and pH 5.5 and incubated at 25°C. Mortality of larvae, and the presence of mycelium or conidia of the tested fungus, and of any saprobic fungi or bacteria on the larvae were recorded daily. All tested fungi except C. clavisporus developed mycelium and conidia on a considerable portion of dead larvae regardless of the initial conidial inoculum within 10 days; C. clavisporus required more time, and 15 days are recommended as a reasonable minimal period of incubation. Larvae exposed to C. clavisporus, whether infected or not, were able to survive for some time in the water film on the WA, and fungal growth on the larvae only started after their death. In the poster, the convenience of the method will be discussed.

Results of preliminary investigations on entomopathogenic Fun07 •232• fungi affecting important weevil and aphid forest pests in southern Chile

Cristian Montalva¹, Alex Gonzalez¹, Eladio Rojas², Eduardo Valenzuela¹, Richard A. Humber

¹Facultad de Ciencias, Instituto de Bioquímica y Microbiología. Casilla 567, Universidad Austral de Chile, Valdivia, Chile; ²Servicio Agrícola y Ganadero, Laboratorio Regional SAG Osorno, Ruta a Puerto Octay U-55-V, Calle de Servicio, Osorno, Chile; ³USDA-ARS Emerging Pests and Pathogens Research, R.W. Holley Center for Agriculture & Health, Ithaca, New York, USA Corresponding author: montalva.cristian@gmail.com

This poster summarizes seven months of collecting activities between November 2016 and May 2017 in an ongoing study on the biodiversity of pathogenic fungi affecting important weevil and aphid forest pests in the southern Chilean region of Los Lagos. In this work we identify, isolate and preserve (whenever possible) the fungal entomopathogens. Among the results of this study were that fungal entomopathogens species belonging to the genus Lecanicillium and Neozygites occurred widely and commonly in southern Chile on aphids associated to native, urban and monoculture plantations trees. Strains of Isaria were isolated from collections of Gonipterus platensis captured on the foliage of Eucalyptus globulus close to the city of Osorno in Los Lagos region. Fungi affecting other arthropods were also collected and isolated from all sites visited from the vegetation and soil

surfaces. These vegetation surveys were undertaken to help to expand the comparatively poor understanding of the biota of Chilean fungal entomopatho-gens. This poster will present the key highlights of this project including new reports of fungal entomopathogens, and a listing of the isolates obtained during this study. This project was funded by FONDECYT (Fondo Nacional de Desarrollo Científico y Tecnológico, Chile) project 11160555.

Pathogenicity of an isolate of Beauveria bassiana at Hypsipyla grandella collected in Brasilia, Brazil

Fun08 •233•



Marcelo T. de Castro^{1,2}, Sandro C. L. Montalvão¹, Daniela A. de Souza², Rose G. Monnerat²

¹Universidade de Brasília, Brasília, Brasil; ²Prédio de Controle Biológico, Embrapa Recursos Genéticos e Biotecnologia, Brasília, Brasil Corresponding author: marceloengflorestal@gmail.com

One of the main obstacles to the establishment of mahogany plantations is the occurrence of Hypsipyla grandella Zeller, which attacks the terminal bud in new seedlings and trees, forming galleries. This study aimed to isolate, identify and verify the pathogenic ability of fungi associated with H. grandella in Brasilia, Brazil. For this, 200 fruit attacked by insects were collected and analyzed. A caterpillar with fungal infection symptom was found. Symptomatic caterpillar was placed in a humid chamber for increased formation of spores and then pure colonies were obtained from serial dilutions and successive subcultures of the fungus. Its molecular, morphological and pathogenic was made. The isolate was identified as Beauveria bassiana (Balsamo) Vuillemin, an entomopathogen widely used in biological control programs. Pure colonies were white, and the optical microscope showed unicellular conidia, conidiogenous cells with denticles conspicuous, featuring the species. As for the pathogenicity test, the isolated was effective, with 70% mortality of newly hatched caterpillar. This is the first report of an entomopathogenic fungus found on H. grandella in Distrito Federal, Brazil. The isolate was incorporated into the Coleção de Fungos de Invertebrados of Embrapa Recursos Genéticos e Biotecnologia.

Occurrence and ecological aspects of entomopathogenic fungi in maize agroecosystems of Central Mexico





 $\textbf{\textit{Miguel Bernardo Najera-Rincon}}^{1}, \textit{Abraham Barajas-Mendoza}^{2}, \textit{Netzahualcoyotl}$ Barron Valle³, Guadalupe Zitlalpopoca-Hernandez⁴, John Larsen⁴

¹Campo Experimental Uruapan, Instituto Nacional de Investigaciones Forestales, Agrícolas y Pecuarias (INIFAP), Uruapam, Michoacan, Mexico; ²Facultad de Biología, UMSNH, Morelia, Michoacan, Mexico; ³Universidad Tecnologica de Morelia, Morelia, Michoacán, Mexico; ⁴Instituto de Investigaciones en Ecosistemas y Sustentabilidad, Universidad Nacional Autónoma de México (UNAM), Morelia, Michoacán, Mexico

Corresponding author: minaj47@hotmail.com

Mexico is characterized for a wide variety of maize agroecosystems. Plant health is a key element in crop production. Soil dwelling pests in maize are commonly managed with chemical insecticides. However, the rhizosphere of crops is naturally inhabited by a broad range of beneficial microbes as entomopathogenic fungi (EPF). The conservation and use of this biodiversity is a promising strategy for agroecological soil pest management. In this sense, we examined the occurrence of native populations of B. bassiana, M. anisopliae and Paecilomyces spp in three contrasting maize agroecosystems: a) high input (improved maize varieties, intensive pesticide use); b) low input (native landraces, no pesticide use); c) intermediate (native landraces, moderate pesticide use), along three phenological crop stages (V6, flowering and senescence). The results showed 343 native isolates of EPF. The low input system registered the highest number (181), the intermediate system 145 and the high input system 17. B. bassiana was found in all agroecosystems while M. anisopliae only in the low input system. The ANOVA analysis and LSD test showed significant differences between systems and B. bassiana occurrence in relation to phenological stages. In a comple-mentary mesocosm experiment, we evaluated the interactions among B. bassiana, native populations of arbuscular mycorrhizal fungi (AMF) and the root feeding insect Phyllophaga vetula. Our results show that the combination of native AMF populations and the EPF B. bassiana induce

tolerance in maize from root herbivory by P. vetula in terms of plant growth performance and nutrition.

Biologically-based control of Varroa destructor

STU Fun10 •235•



Scott Dwyer, Dave Chandler

Warwick Crop Centre, University of Warwick UK Corresponding author: s.dwyer@warwick.ac.uk

The ectoparasitic mite, Varroa destructor and the pathogenic viruses that it transmits, are a significant cause of European honeybee, Apis mellifera (EHB), overwintering colony losses. Varroa has developed resistance to many of the selective synthetic pesticides that were previously used to control it, while the small number of alternative control measures are only partially effective. Therefore, there is a need for new control methods, including microbial control, underpinned by fundamental knowledge to ensure their sustainable use. Previous research has shown that Varroa is susceptible to a wide range of entomopathogenic fungi (EPF), which have potential therefore to be used as biocontrol agents. We have identified strains of fungi that infect varroa but do not kill EHB. However, there are knowledge gaps about how EPF interact with varroa and bees and this is an impediment to commercialisation and use. This project focuses on characterizing the virulence of different EPF against Varroa and bees using a series of laboratory bioassays. A key part of the work is to improve understanding of the effects of temperature upon the activity of candidate EPF strains, as the warm temperatures that are maintained in bee colonies can inhibit fungal activity. In addition, work is being done to investigate the effects of fungal treatment on transmission and activation of bee viruses and to determine whether fungal treatment has priming effects on EHB immune pathways associated with antiviral defenses.

Use of electrical penetration graphs (EPG) and quantitative PCR to evaluate the relationship between feeding behaviour and Pandora neoaphidis infection levels in green peach aphid, Myzus persicae



Chun Chen¹, Yaqiang Hu¹, Sudan Ye², Huajun Hu¹, Xiaoping Yu¹

¹China Jiliang University, Zhejiang Provincial Key Laboratory of Biometrology and Inspection & Quarantine, Hangzhou, 310018, China ²Zhejiang Economic & Trade Polytechnic, Hangzhou 310018, China Corresponding author: aspring@cjlu.edu.cn

A real-time qPCR method was developed, validated, and used to quantity the fungal pathogen, P. neoaphidis, within aphids at different times during infection; colonization rate fitted the Gompertz model well (R²=0.9356). Feeding behaviour of *P. neoaphidis*-infected and uninfected *M. persicae* were investigated, for the first time, using DC-electrical penetration graphs (DC-EPG) that characterized the waveforms made during different aphid stylet probing periods corresponding to epidermis penetration, salivation and ingestion. In the 6 hours following the 12-hour incubation period (to achieve infection), there were significant differences in the number of events of Np (non-probing) and C (stylet pathway) between infected and uninfected aphids. However, the difference between total duration of Np and C were not significantly different between infected and uninfected aphids. There were no significant differences in the number of events or total duration of E1 (phloem salivation) or E2 (phloem ingestion) between infected and uninfected aphids. There were significant differences in mean number of events and total duration of the pd waveform (intracellular punctures) in infected and uninfected aphids. In the 16 hours prior to death, the same differences in behaviour were observed but they were even more obvious. Furthermore, the total duration time of E2 was significantly greater in uninfected aphids than infected aphids, a change that had not been observed in the first 6 h observation period. In conclusion, qPCR quantification demonstrated 'molecular' colonization levels throughout infection, and EPG data analysis during the two periods (during early infection and then during late infection just prior to death) demonstrated the actual physical effects of fungal infection on feeding behaviour of M. persicae. These studies increase our understanding of the interaction between P. neoaphidis and its host aphid.

The antifungal peptide diapausin-1 from Manduca sexta binds to β -1, 3-glucan in fungal cell walls

Fun12 •237•

Qasim Al Souhail¹, Daisuke Takahash ², Michael R. Kanost¹

¹Biochemistry and Molecular Biophysics Department. Kansas State University, USA; ²Chiba University, Japan Corresponding author: Kanost@ksu.edu

Manduca sexta diapausin-1 is a 45 amino acid residue peptide expressed in the fat body after immune challenge, and it is also expressed in the fat body and midgut of naïve larvae at the prepupal stage. Diapausin-1 has activity against yeast and some tested ascomycete filamentous fungi. FITCconjugated diapausin-1 was detected at the surface of Saccaromyces. cerevisiae cells, suggesting that the peptide may bind to fungal cell walls. To investigate the binding target(s) of this peptide, pull-down assays were performed, which demonstrated that diapausin-1 binds to zymosan, an isolated cell wall preparation from the yeast S. cerevisiae. Diapausin-1 binding to zymosan is consistent with its interaction with a component of the yeast cell wall, which may be a clue toward its mechanism of action in blocking fungal growth and survival. Diapausin-1 did not bind to bacterial cell wall peptidoglycan, consistent with its lack of activity against bacteria. Yeast cell walls have many components, including β-1, 3-glucan, mannans, and chitin, but the major structural polysaccharide is β-1, 3-glucan. We found that diapausin-1 binds to curdlan, an insoluble β-1, 3-glucan but not to starch, an α -1, 4-glucan used as a negative control. Surface plasmon resonance analysis of binding between diapausin-1 and β-1, 3-glucan yielded a dissociation constant of ~100 nM. Based on our current and previous results we hypothesize that the binding of diapausin-1 to β -1, 3-glucan in cell walls may directly affect the growth S. cerevisiae and filamentous fungi by interfering with cell wall growth or division. This hypothesis needs to be investigated further using additional methods

Effects of endophytic entomopathogenic fungi on soybean aphid, Aphis glycines.

Fun13 •238•

Eric H. Clifton¹, Stefan T. Jaronski², Brad S. Coates^{1,3}, Erin W. Hodgson¹, Aaron J. Gassmann¹

¹ Department of Entomology, Iowa State University, Ames, Iowa, USA; ² U. S. Department of Agriculture, Northern Plains Agricultural Research Lab. Sidney, Montana, USA: 3 U.S. Department of Agriculture, Corn Insects & Crop Genetics Research Unit, Ames, Iowa, USA Corresponding author: stefan.jaronski@ars.usda.gov

Terrestrial plants can harbor endophytic fungi that may induce systemic changes in plant defenses that in turn affect interactions with herbivorous insects. We evaluated whether the application of entomopathogenic fungi Beauveria bassiana GHA and Metarhizium brunneum F52 to soybean seeds could establish them as endophytes and impact soybean aphid (Aphis glycines Matsumura). In studies conducted in growth chambers, were able to recover both fungi as endophytes in soybean stems but B. bassiana only from the leaves. Furthermore, we found that M. brunneum increased populations of A. glycines, whereas B. bassiana had no measurable effect. The present study confirms that some entomopathogenic fungi could be made endophytic in soybean, however these fungi may have a negative effect on the plants by increasing susceptibility of soybean to the aphid.

STU Fun14 •239• Insect growth regulatory and insecticidal activities of secondary metabolites from entomopathogenic fungi

Ra Mi Woo¹, Jong Hoon Kim¹, Jae Young Choi¹, Seok-Hee Lee¹, Ying Fang¹, Dong Hwan Park¹, Min Gu Park¹, Bo Ram Lee¹, Woo Jin Kim¹, Jae Su Kim³, Yeon Ho Je^{1,2}

¹Department of Agricultural Biotechnology, Seoul National University, Seoul, Korea; ²Research Institute for Agriculture and Life Sciences, Seoul National University, Seoul, Korea; ³Department of Agricultural Biology, Chonbuk National University, Jeonju, Korea Corresponding author: btrus@snu.ac.kr

It is increasingly recognized that the biodiversity in agroecosystems deliver significant ecosystem services to agricultural production such as biological control of pests. Entomopathogenic fungi have been widely studied for their potential as the effective biological control agents. They produce variety of secondary metabolites with insecticidal activities, and it is reasonable to

assume that entomopathogenic fungi might produce secondary metabolites modulating juvenile hormone for their survival against defense mechanisms of host insect. In this study, Beauveria spp. and Lecanicillium spp. strains cultured on unpolished rice medium were extracted with acetone. Both extracts showed juvenile hormone antagonist (JHAN) activities in the yeasttwo hybrid β-galactosidase assay and high insecticidal activity against Aedes albopictus and Plutella xylostella. In addition, to compare bioactivities of secondary metabolites from solid and liquid culture, the Lecanicilium spp. strain cultured on unpolished rice medium or PDB (Potato Dextrose Broth) medium were serially extracted with acetone and ethyl acetate respectively. Both extracts showed JHAN activity and high insecticidal activity against A. albopictus. These results suggested that secondary metabolites of entomopathogenic fungi could be useful for development of novel IGR insecticides.

Solid culture of Isaria javanica and I. fumosorosea on grain substrates for enhanced conidial thermotolerance



So Eun Park. Se Jin Lee. Sihveon Kim. Jong Cheol Kim, Mi Rong Lee, Jae Su Kim*

Department of Agricultural Biology, College of Agricultural & Life Sciences, Chonbuk National University, Jeoniu 561-756, Korea Corresponding author: jskim10@jbnu.ac.kr

Entomopathogenic fungi have high potential in controlling insect pests in agriculture and forest for pest management, however optimizing conidial production system with high productivity and stability still needs additional efforts for successful field application and industrialization. Although many virulent entomopathogenic fungal isolates received many interested in lab environment, very a few numbers of isolates reached agricultural fields as commercial products. Here in this work, we investigated the feasibility of grains substrates for conidial production, particularly focusing on conidial productivity and thermotolerance. Isaria javanica and I. fumosorosea conidia (provided by National Academy of Agricultural Science, Korea) were solidcultured on ten cereal grains for 14 days in Petri dish conditions, and of the tested grains, millet, perilla seed and barley-based cultures showed high conidial production, 15.7 \pm 8.4, 17.3 \pm 5.2 and 13.5 \pm 1.9 \times 10 conida/g in *l*. javanica and 15.9 \pm 0.6, 15.0 \pm 0.5 and 15.0 \pm 0.7 \times 10 conida/g in *l*. fumosorosea, respectively. The two isolates had enhanced thermotolerance at 45°C when cultured on millet, rice, sorghum, and perilla seed. This work suggests possible inexpensive grain substrates for conidial production with enhanced stability for high temperature.

Effect of culture media on storage stability of Beauveria pseudobassiana microsclerotia formulated as dry granules

Fun16 •241•

Laura Villamizar¹, Tracey Nelson², Sean Marshall¹, Sandra Jones¹, Marie Foxwell¹, and Trevor A. Jackson¹

¹AgResearch, Lincoln Research Centre, Private Bag 4749, Christchurch 8140, New Zealand; ²Lincoln University, PO Box 85084, Lincoln 7647, New Zealand Corresponding author: laura.villamizar@agresearch.co.nz

Conidia are the traditional fungal propagules used as the active ingredient in commercial biopesticides. These aerial structures are usually produced in solid fermentation using large volumes of substrate, and requiring complex downstream processes to recover the propagules. Recently, resistant structures -microsclerotia (MS) - that can be easily produced, harvested and applied to the soil for pests and diseases control have been described for Trichoderma harzianum and some species of Metarhizium spp. Microsclerotia are small, compact, melanized structures of hardened fungal mycelia which allow extended persistence in the environment. Microsclerotia are produced by a New Zealand strain of Beguveria pseudobassiana which is under investigation for development as a potential bioproduct to control soil-dwelling scarab pests. The aim of this research was to study the storage stability of MS produced in liquid fermentation using two C:N ratios, formulated with diatomaceous earth (DE) and air dried to <3% moisture, to obtain granules with a final concentration of 10⁵ MS/g. For comparison, conidia were produced on cooked rice and formulated with DE under the same conditions. Capacity to germinate of MS granules varied with C:N ratio and storage temperature. MS produced using a higher carbon source concentration retained 100% germination after six months of storage

at 4°C; when stored at 20°C, the germination rate was reduced to 68%. In contrast, MS produced in low carbon medium showed reduced germination (0 to 18%) after storage for more than 2 months at either temperature and had lost viability completely after six months. For surviving MS, conidia production seven days-post rehydration was similar for granules stored at either temperature and produced from both C:N ratios (>10° conidia/g). Conidia germination from formulated granules was also stable under refrigeration, but no germination was detected after 4 months at 20°C which could be related to the low moisture content in the formulation. The results demonstrate that MS produced with high carbon media are capable of surviving under low moisture, ambient temperature storage and able to produce around 1000 conidia/MS after germination; these attributes suggest that MS could be used as a new sustained release strategy to control soildwelling insects.

Beauveria bassiana produces microsclerotia-like propagules with active peroxisome biogenesis

Fun17 •242 •

Carla Huarte-Bonnet¹; Flávia R. S. Paixão¹, Éverton K. K. Fernandes²; Nicolás Pedrini¹

¹Instituto de Investigaciones Bioquímicas de La Plata (CCT La Plata CONICET-UNLP), Facultad de Ciencias Médicas, La Plata, Argentina; ²Instituto de Patologia Tropical e Saúde Pública, Universidade Federal de Goiás, Goiânia, Brasil Corresponding author: carlahb@hotmail.es

Entomopathogenic fungi are able to produce several types of propagules such as aerial conidia in solid media, or blastospores, submerged conidia and micro-sclerotia in liquid cultures. Microsclerotia (MS) are usually melanized compact hyphal aggregates that are tolerant to dessication. As far as we know, there are no reports about the production of melanized microsclerotia by Beauveria bassiana isolates. The aim of this work was to describe microsclerotial growth in B. bassiana and to start with studies about the molecular and physiological mechanisms implicated in the production of these propagules. B. bassiana strain GHA was cultured in complete liquid medium for 4 days at 26°C in the dark with vigorous agitation (250 rpm). Microsclerotia-like structures were separated by centrifugation and processed for microscopy observation and real time qPCR analysis. Under these culture conditions, B. bassiana was able to produce compact brownish aggregates that were able to germinate and produce viable conidia after desiccation. Optical microscope images showed similarity in form, structure and size with microsclerotia reported from other entomopathogens, and staining with 3,3-diaminobenzidine (DAB) revealed high peroxidase activity. Genes encoding for peroxisome biogenesis factors, named peroxins, showed high expression levels (up to 17-fold induction) for Bbpex5, Bbpex7, Bbpex14/17 and Bbpex19 genes in 4 days cultures compared with conidia used as starting inocula. Additional studies are being carried out to elucidate the relationship between microsclerotia formation and peroxisomal biogenesis, as same as stress tolerance, cell surface alterations and virulence against insect hosts.

Quantification of fungal growth and destruxin A during infection of Galleria mellonella L. (Lepidoptera: Pyralidae) larvae by Metarhizium brunneum Petch (Ascomycota; Hypocreales)



Inmaculada Garrido-Jurado¹, Álex Ríos-Moreno¹, María del Carmen Raya-Ortega², Enrique Quesada-Moraga¹

¹Department of Agricultural and Forestry Sciences, ETSIAM, University of Cordoba, C4Building, Campus of Rabanales, 14071 Cordoba, Spain; ²Department of Agronomy, ETSIAM, University of Cordoba, C4Building, Campus of Rabanales, 14071 Cordoba, Spain.

Corresponding author: g72gajui@uco.es

The entomopathogenic ascomycete Metarhizium sp. produce several secondary metabolites, but principally destruxin A, where lack of studies concerning production of this compound is most likely the biggest obstacle for the registration of new fungal strains. Although several studies focus on the production of destruxin A in culture media, only few studies examine destruxin A in vivo during host infection. For that in the current work, the dynamic of fungal growth and secretion of destruxin A was monitored in vivo

during the infection process of Galleria mellonella used as an insect model by M. brunneum EAMa 01/58-Su or BIPESCO5 strains. Total mortality of sixth G. mellonella larvae that were immersed in a suspension of 1.0 x 10⁸ conidia mL⁻¹ ¹ of *M. brunneum* EAMa 01/58-Su or BIPESCO5 strains reached 85.5 % and 78.8 %, respectively, and the percentage of cadavers with fungal outgrowth was low at 12.2 % and 4.4 %, respectively. The average survival time of treated larvae was 5.5 days for both fungal strains. Destruxin A secretion was parallel to the fungal growth of EAMa 01/58-Su, but not coupled with that for BIPESCO5. EAMa 01/58-Su and BIPESCO5 strains secreted destruxin A from days 2 to 6 and from days 2 to 5 post treatment, respectively. For EAMa 01/58-Su and BIPESCO5, the maximum titer of destruxin A in the host was on day 4 at 0.369 and 0.06 µg/larva, respectively, and throughout the pathogenic process, the total production was 0.6 and 0.09 µg/larva, respectively. These results suggest that destruxin A could be virulence factor for EAMa 01/58-Su strain, whereas for BIPESCO5, the virulence could require the involvement of other factors as well as destruxin A during the infection process.

Infection of the tick Rhipicephalus microplus treated with conidia or blastospores of Metarhizium spp. and Beauveria bassiana

Fun19 •244•

Cíntia C. Bernardo¹, Lucas P. Barreto¹, Cárita de S. R. de Silva¹, Christian Luz¹, Walquíria Arruda², **Éverton K.K. Fernandes**¹

¹Instituto de Patologia Tropical e Saúde Pública, Universidade Federal de Goiás, Goiânia, Goiás, Brazil; ²Instituto de Ciências Biológicas, Universidade Federal de Goiás, Goiânia, Goiás, Brazil. Corresponding author: evertonkort@ufg.br

The current study compared the virulence of conidia and blastospores of Metarhizium robertsii (IP 146), Metarhizium anisopliae s.l. (IP 363) and Beauveria bassiana (IP 361 and CG 307) against larva and engorged female of Rhipicephalus microplus; in addition, this study evaluated the development of both fungal propagules on the tick cuticle. R. microplus larvae were treated with Tween 80[®] 0.01% solution (control) or fungal suspensions at 10⁶, 10⁷ or 10⁸ propagules mL⁻¹. In the groups treated with 10⁶ propagules mL⁻¹, only the larvae treated with conidia of IP 361 presented mortality superior than that recorded in the control group at day 10 after treatment, 87% and 5% respectively. Engorged females were immersed in fungal suspension (10⁷ propagules mL⁻¹) for 1 min, and their biological parameters monitored daily until the end of oviposition. Only females treated with blastospores suspensions of IP 146 or IP 361 presented lower nutrient and egg production indices in comparison to the control groups; the percent control was higher when females were treated with blastospores suspensions of IP 146 (98%) or IP 361 (92.9%) in comparison to the groups treated with conidial suspensions, 71% and 63.3%, respectively. Engorged females treated with IP 146 or IP 361, conidia or blastospores, were processed for scanning electron microscopy. Germination of blastospores of IP 146 and IP 361 were observed on the tick cuticle after 4 h incubation. No structures suggesting cuticle penetration were observed in blastospores of IP 146; however, evidences of blastospores penetration in natural openings were seen 96 h after treatment. Appressoria in blastospores of B. bassiana IP 361 were observed after 4 h of incubation, whereas germ tubes from conidia of both isolates were observed only 48 h after treatment. In conclusion, conidia were more virulent to R. microplus larvae than blastospores, whereas blastospores were more virulent against engorged females and developed faster than conidia on tick cuticle. Blastospores might be promising for use in biocontrol of ticks, since they were virulent against R. microplus and presented rapid development on its

Secretome of the biocontrol fungus Beauveria bassiana Fun20 •245• related to infection of the cattle tick Rhipicephalus microplus

Lucélia Santi¹, Caio Jr B. Coutinho-Rodrigues², Markus Berger¹ Wendell M. S. Perinotto³. John R. Yates⁴. Jorge A. Guimarães¹. Vania R.E.P. Bittencourt², Walter O. Beys-da-Silva

¹Hospital de Clínicas de Porto Alegre (HCPA), Centro de Pesquisa Experimental, Porto Alegre – RS. Brazil: ²Universidade Federal Rural do Rio de Janeiro (UFRRJ), Departamento de Parasitologia Veterinária, Seropédica – RJ, Brazil; Universidade Federal do Recôncavo da Bahia, Laboratório de

Parasitologia e Doencas Parasitárias, Cruz das Almas - BA, Brazil: ⁴The Scripps Research Institute, Department of Chemical Physiology, La Jolla - CA, USA

Corresponding author: luca.santi@yahoo.com.br

The biological control is considered a natural and ecological alternative tool to overcome the problems caused by the use of chemical methods in pest control. In addition, the biocontrol presents several advantages when compared to its chemical analogues, regarding to environmental impact, cost, handling, specificity and resistance development. Among microorganisms used in biocontrol, fungi stand out, including the filamentous fungus and arthropod pathogen Beauveria bassiana. This fungus is one of the most studied and applied worldwide. In this study, using proteomics and bioinformatics approaches, a total of 256 proteins were identified in Rhipicephalus microplus tick cuticle-induced secretome. Proteins involved in different important processes for the biocontrol activity, including degradation of the cuticle (proteases and lipolytic enzymes), defense/antioxidants (catalases, oxidoreductases) and effectors (glycoside hydrolases) were detected. The identification of possible molecular mechanisms involved in fungal infection on specific pests provides a better understanding of biocontrol activity and consequently helps to develop strategies to optimize this process. Moreover, these potential molecular markers, proteins, in a model that induces the infection system enables the oriented search for more efficient strains with greater virulence and specificity in the biological control of R. microplus tick.

Secretome of the biocontrol fungus Metarhizium anisopliae Fun21 •246• related to infection of the cattle tick Rhipicephalus microplus

Walter O. Beys-da-Silva¹, Markus Berger¹, John R. Yates², Jorge A. Guimarães¹, Lucélia Santi¹.

¹Hospital de Clínicas de Porto Alegre (HCPA), Porto Alegre, RS, Brazil; ²The Scripps Research Institute, Department of Chemical Physiology, San Diego, CA, USA Corresponding author: walterbeys@yahoo.com.br

The biological control is considered a natural and ecologic alternative to control pests avoiding the issues related to the use of chemical compounds. The biocontrol presents several benefits as the lower environmental impact, cost, handling, specificity, and the non-development of resistant pest strains. However, normally biopesticides need more time to kill and control pests when compared to chemical control. Among microorganisms used in biological control the fungus Metarhizium anisopliae is one of the most studied and applied to control several pests, including the cattle tick Rhipicephalus microplus. The understanding of the host infection process may contribute to optimize the biocontrol through the reduction of the time to kill the target pest. In this work, applying shotgun proteomics and an in vitro culture strategy to induce the infection system we identified more than 400 proteins in M. anisopliae secretome. Among these proteins, 133 were exclusively identified in the infection condition and 56 were found differentially regulated when compared to control. Several of these proteins are related to host cuticle digestion and penetration, fungal defense and modulators of host immune system. Besides, many of these proteins are being reported for the first time linked to M. anisopliae infection and 12 are shared in infection of insects and probably not connected to host specificity. The secreted protein profile identified here represents a contribution to the understanding of host-pathogen interaction and may help to optimize biocontrol strategies and to select fungal strains more specific and efficient to control the cattle tick R. microplus.

Isolation and characterization of Bionectria ochroleuca as an entomopathogenic fungi from edible Saturniid in Kenya

Fun22 •248•

Kimemia, Jane Wanjiru¹; Ombura, Levi Odhiambo¹, Elizabeth Kusia^{1,2}, Fathiya Khamis¹, Christian Borgemeister², Sunday Ekesi¹, **Sevgan Subramanian**¹

> ¹International Centre of Insect Physiology and Ecology, Nairobi, Kenya, ² Center for Development Research, University of Bonn, Germany, Kenya Corresponding author: ssubramania@icipe.org

Traditionally, endophytic fungi are known to reside in living plant tissues and are asymptomatic to their host. However, the potential of this special niche of fungi to have other roles especially as an entomopathogen is increasingly gaining relevance. This study reports on Bionectria cf. ochroleuca as a

potential candidate for microbial control of lepidopteran pests. This strain was isolated from Bunaea alcinoe (Stoll, 1780) cadaver and it had the characteristic pale yellow to white ascomata and two-celled hyaline ascospores associated with Bionectriaceous fungi. Conidia were obtained from two-week-old plates cultured on potato dextrose agar. The bio efficacy of the fungus was established on second instar Galleria mellonella larvae subjected to fungal concentrations of 1 x 10² to 1 x 10⁹ conidia mL⁻¹. The treatment of fungus was undertaken both as contamination of artificial diet and onto the sterile soil. The treated insects were observed for mortality at 24h interval for 21 days. At a dose of 1 x 10⁹ conidia mL⁻¹, highest mortality of only 56 % was recorded in the artificial diet bioassay as compared to 83.3 % in the sterile soil bioassay. Mycosis was observed in larvae subjected to sterile soil bioassay with mycosis rates of 38, 18 and 10% at concentrations of 1 x 10⁵, 1 x 10³ and 1 x 10⁴ conidia mL⁻¹. This fungus was characterized using morphological features and through molecular analysis of its internally transcribed spacer (ITS) region. To our knowledge this is the first report of entomopathogenic fungi belonging to Bionectria isolated from edible saturniid caterpillars. Further studies on the efficacy of this entomopathogen to Saturniids and other lepidopteran pests needs to be undertaken.

Colonization of some horticultural crops with indigenous Beauveria bassiana as endophytic prophylactic protective agent

Fun23

Mohammed. I. Elbashir¹, Ebtisam M. Bashir², Alnazir. I. Mohamed²

Department of Bio pesticides and Bio fertilizers- Environment and Natural Resources and Desertification Research Institute, P.O. Box 6096, Khartoum, Sudan

> ²Assistant Professor, Department of Plant Protection and Environmental Studies, Faculty of Agriculture, Al Zajem Al Azhari University, Khartoum North, Sudan; 2 Department of Plant Protection and Environmental Studies, Faculty of Agriculture, Al Zajem Al Azhari University, Khartoum North, Sudan, Corresponding author: fataloope@yahoo.com

Beauveria bassiana has been recovered as an endophytic colonist from several plant species and has been shown to protect plants against plant pathogens and insect pests worldwide. This research was conducted at the laboratories of National Center for Research and fields of Zadna International Company for Agricultural Production and Investment. The purpose was to to establish Beauveria bassiana inside some horticultural crops viz; okra, eggplant, mango, orange and grapevine so as to see the possibility of colonization of the plant parts. For the first time worldwide one of Sudanese indigenous isolate of Beauveria bassiana successfully colonized mango crop, orange and okra crop. Whereas this the first study in Sudan to prove colonization of grapevine by B.bassiana. Ten to two weeks after inoculation; the fungus which had been sprayed on the plants was recovered in the leaves, stems and roots of all treated crops by 80-100% except roots in case of mango, grape and orange plants.

Establishment protocols for Beauveria bassiana in Arachis hypogea, potential for controling plant pests and diseases

Fun24

M.I. Elbashir¹, Abdelbaset Hassan², Edur Balla Zahran²

Department of Bio pesticides and Bio fertilizers, Environment and Natural Resources and Desertification Research Institute, P.O. Box 6096. Khartoum, Sudan; Alzaiem-Alazhari University, Faculty of Agriculture Corresponding author email: fataloope@yahoo.com

Beauveria bassiana is one of entomopathogenic fungi with the ability to colonize plants endophytically. These fungi have recently proven their efficacy as endophyte in protecting plants from plants pests and diseases. This study aimed to test four protocols to establish B. bassiana endophytically in the groundnut (Arachis hypogea), expecting subsequent evaluations of of this fungus as biocontrol agent against pests and diseases on groundnut. Plants were grown from surface-sterilized seeds for two weeks before receiving B. bassiana suspension at 1x10⁹ conidia/ml applied either as a foliar spray or a soil drench or soaked seeds or coated to seeds. dividually surface sterilized, cut into five sections, and incubated in selective media up to the presence of the fungus on the surface of the plant part. The media is checked every 2-3 days to observe fungal outgrowth on plant sections and record the existence of B. bassiana to estimate the extent of its endophytic colonization. Analyses of inoculation success comparing the occurrence of B. bassiana within a given plant part (i.e., leaves, stems or roots) across treatments and controls. The percentage colonization of the fungus within one month for root stem and roots respectively was 93, 86, 26% for seed coating 100, 73, 33 for seed soaking. When drenched the percentage was 100, 86 and 73. When leaves were sprayed the results were 73, 100, and 100.

Baiting system-mediated collection and characterization of entomopathogenic fungi for library construction; JEF entomopathogenic fungal library

STU Fun25

Jong Cheol Kim, Sihyeon Kim, Se Jin Lee, Mi Rong Lee, Seok Ju Lee, So Eun Park, Jae Su Kim

Department of Agricultural Biology, College of Agricultural & Life Sciences, Chonbuk National University, Jeonju 561-756, Korea Corresponding author: jskim10@jbnu.ac.kr

Entomopathogenic fungi are facultative microorganisms and high potential in pest management. Some genera have been used as biological control agents worldwide. Construction of entomopathogenic fungal library could be a strong platform for the development of highly effective biopesticides. Herein this work, we have isolated entomopathogenic fungi using two different methods; selective agar medium-based isolation and Tenebrio molitor (mealworm) larvae-mediated baiting system. The T. molitor-mediated baiting tool showed much higher isolation efficiency than the selective agar medium including antibiotics and fungicide. The entomopathogenic fungal library, which was named as Jeonbuk (=Chonbuk) National University Entomopathogenic Fungi (JEF), was constructed and it had 350 isolates which were identified as 12 genera and 29 species. The fungal collection was grouped to two different categories based on the Internal Transcribed Spacer (ITS) sequence in a phylogenetic analysis. The morphology and physiological characteristics of the 17 species-representative isolates were characterized. Some of the species-representative isolates showed high virulence against *T.* molitor, Riptortus pedestris and Plutella xylostella in laboratory conditions. Overall the production of pathogenesis-related enzymes such as chitinase, Pr1 protease and lipase in LB medium was higher than in the SDB and PDB media. In the thermotolerance assay, Purpureocillium lilacinum isolate had higher thermotolerance than other isolates. This work reports the efficient construction of entomopathogenic fungal library using *T. molitor* and this library could be used as an attractive resource for research and development of fungal insecticides in pest management

NEMATODE DIVISION POSTERS

Mechanism of action of Bacillus methylotrophicus to control Meloidogyne incognita on cotton

Nem01 •249•

Sandro C. L. Montalvão^{1,2}, Marcelo T. de Castro², Rose G. Monnerat², Luiz E. B. Blum¹

¹Departamento de Fitopatologia, Instituto de Biologia, Universidade de Brasília. Brasília. Brasil: ²Prédio de Controle Biológico, Embrapa Recursos Genéticos e Biotecnologia, Brasília, Brasil Corresponding author: sandro.coelho@yahoo.com.br

Cotton crop (Gossypium hirsutum) is affected by various diseases of economic importance, among them, the root galls caused by Meloidogyne incognita races 3 and 4 has such significance. The objective of this study was to study the interaction of Bacillus methylotrophicus with M. incognita. The first assay testing different biological products to control M. incognita allowed us to select the product Onix (B. methylotrophicus) to conduct the subsequent tests to understand the mechanism of action of the bacteria to control nematodes. For this evaluation it was adopted a method that enabled to visualize the pathogen inside the root, allowing monitoring nematode penetration in the roots and its development until beginning of reproduction. With the method of staining with acid fuchsin it was possible to verify that penetration o J2 decreased in early days after inoculation in plants treated with bacteria, but this difference has been lost over the course of days. Through adoption of a method of toluidine blue were noticeable

abnormalities on the giant cells with vacuoles and thinner cell wall, and females presenting large vacuoles inside. The reactions observed in the histopathology assay may indicate the induction of plant resistance mediated by the B. methylotrophicus. This suggests further studies to understand this interaction.

Mechanism of action and evaluation of efficiency of products with Bacillus to control Meloidogyne incognita races 3 and 4 on cotton under greenhouse

Nem02 •250•

Sandro C. L. Montalvão^{1,2}. Marcelo T. de Castro². Rose G. Monnerat², Luiz E. B. Blum¹

¹Departamento de Fitopatologia, Instituto de Biologia, Universidade de Brasília. Brasília. Brasil: ²Prédio de Controle Biológico, Embrapa Recursos Genéticos e Biotecnologia, Brasília, Brasil Corresponding author: sandro.coelho@yahoo.com.br

The root-knot nematode (Meloidogyne incognita races 3 and 4) of cotton, is an important pathogen of cotton crop that can cause damage higher than 40%. The objective of this study was to test the efficiency of commercial products with Bacillus to control M. incognita races 3 and 4 of cotton under greenhouse. In addition to biological products, a treatment with Carbofuran and an untreated control were included. After the application of the products, the plants were evaluated for 200 days. During the assay were evaluated plant height, fresh and dry shoot weight, fresh and dry root weight and the nematode reproduction factor. There was a positive response during initial growth of plants treated with biological agents in comparison with the control plants treated with and without chemical product. The products with B. subtilis (Rizos), B. amyloliquefasciens (NemaControl) and B. subtilis + B. lincheniformis (Nemix) showed better results for fresh shoot weigh when compared to the control plants with nematode. With respect to dry shoot weight, the treatments with active ingredients of B. methylotrophicus (Onix) and B. subtilis it was higher than in control, especially when treated with B. methylotrophicus. As for fresh root weight, only the treatment with B. methylotropicus provided weight statistically higher compared to the control. As far dry root weight is concerned, in plants without nematode, the treatment with B. methylotrophicus was better than the others treatments again. Same trend was observed for nematode reproduction factor. Therefore, there is evidence that the use of biological products can be effective to control M. incognita, becoming a tool to be used in the management to this important pathogen of cotton crop.

Susceptibility and viability of larval Diabrotica virgifera virgifera as a host for entomopathogenic nematodes.

Nem03 •251•

Ryan Geisert, Bruce Hibbard, Kent Shelby, Tom Coudron

Agricultural Research Service, U.S. Department of Agriculture, Columbia, MO Corresponding author: rwg5h8@mail.missouri.edu

Seven populations of entomopathogenic nematodes were obtained and reared in mass for laboratory experiments. Populations included species of Steinernema feltiae, S. riobrave, S. carpocapsae, S. diaprepesi, Heterorhabditis megidis, H. bacteriophora, and a Missouri wild type population of H. bacteriophora. Larval third instar Diabrotica virgifera virgifera were exposed to either 60 or 120 nematodes for six days. Afterwards the larvae were dissected in order to determine the presence of nematode infection. Results indicated a significantly higher proportion of larvae were infected with species in the Heterorhabditis family and S. diaprepesi species than the other Steinernema families for both exposure rates. An additional experiment involved exposing third instar larvae to 60 nematodes of either S. feltiae, H. megadis, H. bacteriophora, or the wild type H. bacteriophora and allowing the nematodes to complete their lifecycles and emerge. Viability of the emerging nematodes was observed for up to nine weeks after leaving the host. After nine weeks nematode viability was found to be: H. megadis (87%), H. bacteriophora (95%), wild H. bacteriophora (89%), S. feltiae (86%). Results have shown that while the rate of infection may vary, all nematode species tested were capable of infecting larval D. virgifera. Additionally the viability of those nematodes able to emerge from D. virgifera hosts was not negatively impacted.

Efficacy of Steinernema carpocapsae formulated in sodium alginate capsules against Phyllophaga vetula in semi-field conditions

STU Nem04 •252•

Studying entomopathogenic nematodes in an undergraduate lab setting: the consequences of soil sterilization on EPN infection rates.

Nem06 •254•

Jaime Ruiz-Vega¹, C. Cortés-Martínez¹ C. García-Gutiérrez², F. Diego Nava¹

¹Instituto Politécnico Nacional, CIIDIR U. Oaxaca. Sta. Cruz Xoxocotlán, Oax., Mexico; ²CIIDIR U. Sinaloa, Mexico Corresponding author: iveaa@ipn.mx

To evaluate the capacity of the nematode Steinernema carpocapsae formulated in alginate capsules to control third instar larvae of Phyllophaga vetula, capsules of alginate containing each 700 IJ's were elaborated. Prior to its elaboration by the ionic gelation process, the sodium alginate solution received 10% maize sap as a phago-stimulant, to which sodium benzoate had been added at concentrations of 1.5 and 3.0 g / L. To obtain the maize sap, 5 $\,$ stems of maize seedlings were ground in a rotary juice extractor, applying the juice obtained to the sodium alginate solution. The Criollo Bolita white maize variety was planted the field in plastic bags of 40 x 40 cm filled with moist soil. Although in general it rained well, irrigation was applied when it was required. After two weeks two plants per bag were left and five larvae of P. vetula were applied per bag, discarding those that did not buried into the soil after 3 minutes. During plant growth, plant heights were recorded by duplicate every two weeks from the soil to each one of the top three leaves. At maize flowering the plants were uprooted to evaluate the presence of alive larvae and to determine plant and root dry weights. The data were processed in Excel and later analyzed with the SAS package to obtain the ANOVA and the Tukey test (α <0.05). All the treatments applied were different to the control (water suspension), but no differences were found in percent control (PC) between concentrations of sodium benzoate. Neither the final height of the maize nor the dry weight of the forage were statistically different; however, the root-forage ratio (PRF) was higher in treatments with capsules. The treatment where IJ's were applied in aqueous suspension tended to produce the largest PC, as well as the highest PRF. The encapsulated nematodes had the same control efficacy as the nematodes applied in suspension, without giving significant differences between the levels of phago-stimulant. However, the root-foliage weight ratio was higher in treatments with alginate capsules.

Detecting and quantifying EPN in Virginia, USA, vineyards using qPCR Nem05 •253•

Glen N. Stevens¹, Greta Mosley², J. Chris Bergh³

¹School of Natural Sciences and Mathematics Ferrum College, Ferrum, VA, USA, ²Department of Biological Sciences, Virginia Tech, Blacksburg, VA, USA, ³Department of Entomology, Virginia Tech, Blacksburg, VA, USA Corresponding author: gstevens3@ferrum.edu

For over a century, the grape root borer (GRB) has been known as an insidious threat to vineyard production in much of the eastern USA. While GRB has been detected in most vineyards sampled in Virginia, the degree of infestation varies widely among them. In addition, the soil factors that appear to promote GRB infestations are considered to be relatively unsuitable for entomopathogenic nematodes (EPN). Our research objectives were to identify and quantify populations of the entomopathogenic nematodes Heterorhabditis bacteriophora and Steinernema carpocapsae to determine whether variability in EPN may help explain GRB infestation patterns. These two EPN species are common and widely distributed, are considered important biological control agents of numerous crop pests, and are known to be virulent to grape root borer (GRB) larvae. Research was conducted over the 2016 growing season at 14 vineyards across Virginia. Using a qPCR approach to identify EPN presence, we found both EPN at varying densities across the 14 vineyards. Total seasonal grape root borer captures in pheromone traps also varied substantially among the 14 vineyards. This talk presents a model that integrates assessments of soil texture, EPN presence, and GRB abundance. In addition, we focus on next steps, including the suitability of different vineyards conditions for inundative EPN applications to treat existing GRB infestations.

Glen N. Stevens, Glen'Asia Cox

School of Natural Sciences and Mathematics, Ferrum College, Ferrum, VA, USA

Corresponding author: gstevens3@ferrum.edu

Entomopathogenic nematodes (EPN) are intriguing organisms to use in a teaching context, as they integrate concepts of predation, parasitism, mutualism and symbioses. While the biology of EPN is complex, their life cycle is simple to understand, and their small size, short life cycles, and amenability to lab culture makes them suitable for original, student-derived research projects that are integrated within semester-long courses. This presentation details how we integrated this approach into a semester-long research project within a sophomore-level Ecology lab at Ferrum College. In collaboration with the faculty member, the student is asked to come up with a research question, write a proposal justifying the question and laying out the experimental protocol, conducting the research, and presenting the data; all of this is done in the context of a single semester-long project. The results we present here focus on the consequences of soil sterilization on EPN virulence, but the process is amenable to many different questions. Current plans describe a similar approach for freshman-level Biology courses at Ferrum College beginning in the fall of 2017; if such an active learning format can be scaled up to multiple sections it is possible that 100 or more students per semester may be exposed to EPN biology and the process of research. Beyond its value as a teaching tool that makes biological research real, this approach offers an unusual opportunity to crowd-source questions regarding EPN biology. We welcome suggestions for questions relating to EPN behavior or ecology that might be effectively answered in this context.

Occurrence of Steinernema arenarium in agricultural soil Nem07 •255• of School Farm of the State University of Londrina, Paraná, Brazil

Bruna A. Guide¹, Thiago A. Fernandes¹, Matheus C. Marcomini¹, Dhiego G. Ferreira², Dayanne da S. Alves², Viviane S. Alves², Pedro. M. O. J. Neves¹

> ¹State University of Londrina, Paraná, Brazil; ² State University of the North of Paraná, Cornélio Procópio, Brazil. Corresponding author: pedroneves@uel.br

The entomopathogenic nematodes (EPNs) (Steinernematidae and Heterorhabditidae) are cosmopolitan and can be found in soils of the most diverse ecosystems. Thus, the objective of this work was to isolate and identify EPNs from crop soils of farm school of the State University of Londrina, Paraná, Brazil. Eighteen soil samples were collected in cultivated areas with soybean and corn. In the laboratory, each sample was divided into three sub-samples (approximately 100g of soil) and each was placed in a 500mL plastic vials. Then, each vial received five last instar Galleria mellonella (Lepidoptera: Pyralidae) caterpillars and were covered and stored, without light, in a climatic chamber at 25 ± 1°C, for 72 hours. Dead caterpillars with symptoms of EPNs infection were transferred to Petri dish with a dry filter paper and stored under the same conditions for five days. After this period, the caterpillars were transferred to a White trap to collect the infective juveniles. The infective juveniles (IJs) were again inoculated in G. mellonella larvae for multiplication. Molecular identification was done from the amplification and sequencing of the rDNA ITS region using the primer pair SSU18A (AAAGATTAAGCCATGCATG) and SSU26R (CATTCTTGGCAAATGCTTTCG). Sequences were obtained from automated sequencer (ABI-PRISM 3500 XL) and later edited and aligned with the help of the MEGA v5.0 program. The BLAST application (NCBI) was used to compare the sequences obtained with the GenBank database, seeking information (genetic similarity) to identify the species. Of the 18 soil samples collected, one was positive for NEPs, and preliminary genetic analyzes identified the isolate as Steinernema arenarium. To the best of our knowledge, this is the

first citation of the natural occurrence of this species in Brazil

Investigation of how the entomopathogenic nematode Heterorhabditis bacteriophora suppresses the host immune system

STU Nem08 •256•

Paul Medina, Dihong Lu, Dennis Chang, Adler Dillman Department of Nematology, University of California, Riverside Corresponding author: adlerd@ucr.edu

Entomopathogenic nematodes (EPNs) are insect parasites often used in pest management. The infective juvenile (IJ), which is the only free-living stage and analogous to the dauer stage in C. elegans, is associated with pathogenic bacteria, which they carry in their gut. Developmentally arrested IJs invade insect hosts and initiate the parasitic part of their life style by releasing pathogenic bacteria from their gut. It is widely believed that the bacteria kills the insect hosts and then proliferates and supports nematode growth and reproduction. However, it has been shown that EPNs in the genus Steinernema contribute to pathogenesis through their secreted products, to manipulate host immunity and promote bacterial growth. We are testing this hypothesis in Heterorhabditis bacteriophora, an EPN that is widely used in both research and pest control, but that has not been previously shown to contribute to pathogenesis. We have adapted a method to obtain large amounts of bacteria-free IJs by culturing nematodes on a non-colonizing bacterial strain. We have developed a method to activate the IJs in vitro and are currently harvesting secreted protein products from activated IJs. Here we discuss the method of activation and discuss the quantity and identity of secreted protein products. The goals are to determine whether H. bacteriophora nematodes secrete active products to modulate host immune system and to characterize these components, focusing on the protein products.

DISEASES OF BENEFICIAL INVERTEBRATES DIVISION POSTERS

First confirmed report of a bacterial brood pathogen in stingless bees

DBI01 •257•



Jenny L. Shanks^{1,2}, Anthony M. Haigh¹, Markus Riegler³, Robert N. Spooner-Hart^{1,3}

¹School of Science and Health, Western Sydney University, Locked Bag 1797, Penrith, NSW 2751, Australia; ²Plant Health Australia, Phipps Close, Deakin, ACT 2600, Australia; 3 Hawkesbury Institute for the Environment, Western Sydney University, Locked Bag 1797, Penrith, NSW 2751, Australia Corresponding author: m.riealer@westernsvdnev.edu.au

Susceptibility to brood pathogens in eusocial stingless bees (Meliponini), alternative pollinators to honey bees in subtropical and tropical regions of the world, is unknown. Brood losses in managed colonies of the Australian stingless bee Tetragonula carbonaria were studied over 20 months. Symptoms and bee behaviour were noted and recorded. We isolated a disease-causing bacterium, Lysinibacillus sphaericus (Firmicutes, Bacillaceae), from worker and gueen larvae, brood cell provisions and honey stores, by using microbiological and molecular techniques. This is the first confirmed record of a brood pathogen in stingless bees. Subsequently, a L. sphericus strain with an identical 16S rRNA gene sequence has been isolated from a symptomatic Austroplebeia australis colony. Our findings warrant further research into the prevalence of L. sphaericus infections and possibly other brood pathogens across Meliponini.

Transcriptome sequencing and in silico cDNA library construction of Varroa destructor, a parasitic mite of honeybee



Woo Jin Kim¹, Seok-Hee Lee¹, Ying Fang¹, Jong Hoon Kim¹, Dong Hwan Park¹, Ra Mi Woo¹, Min Gu Park¹, Bo Ram Lee¹, Jae Young Choi², Yeon Ho Je^{1,2}

¹Department of Agricultural Biotechnology, Seoul National University, Seoul, Korea: ²Research Institute for Agriculture and Life Science, Seoul National University, Seoul, Korea Corresponding author: btrus@snu.ac.kr

Varroa destructor is a small ectoparasitic mite belonging to Acari which attacks Honeybee, Apis mellifera. V. destructor feeds on honeybee

hemolymph, and often stays inside the capped brood cell to lay eggs on the larvae and pupae. This mite also harbors small RNA viruses such as deformed wing virus, and transmits these viruses in the infested beehive. To survey the genes of V. destructor, its total RNA was subjected to RNA-seq to construct in silico cDNA library. By using the Illumina HiSeq 2000 sequencing platform, 101 base paired-end sequencing generated 2 x 107,748,792 raw reads to obtain 2 x 8.3 Gbase of quality filtered nucleotide sequences. An in silico cDNA library was constructed by Trinity de novo assembler, and streamlined by using TransDecoder ORF finding, and CD-HIT clustering program. The final version of cDNA library contains 28,023 contigs with protein coding capability. This library was subjected to arthropoda reference sequences from EMBL database by BLASTX, and it returned 15,075 contigs which aligned with known sequences (e-value < 1e5). The contig sequences also reflect the metagenomic viral flora in the mites which harbors bee pathogens. These newly assembled gene sequences will provide valuable information for molecular biology of V. destructor and future pest control approaches such as using RNAi technology.

MICROBIAL CONTROL DIVISION POSTERS

MCn01 •259• Serratia marcescens as a bacterial pathogen of sunn pest, Eurygaster integriceps (Hemiptera: Scutelleridae)

> **Mehdi Dastranj^{1,2}**, Marjan Heidarian Dehkordi¹, Sajjad Sarikhan², Ghasem Hosseini Salekdeh²

¹Department of Plant Protection, University of Tehran, Karaj, Iran; ²Systems Biology Department, Agricultural biotechnology Research Institute of Iran, Agricultural Research, Education and Extension Organization (AREEO), Karaj, Iran Corresponding author: m.dastranj@ut.ac.ir

Sunn pest, Eurygaster integriceps, is the most important pest of wheat and barley in the Middle East which threat food security. In order to find pathogenic bacteria with the potential to control sunn pest, a red-pigmented bacterial strain was isolated from diseased sunn pest collected from barley fields in Varamin, Iran. Bacterial isolate DNA was extracted using gram negative bacteria genomic DNA extraction kit (IBRC), then PCR was conducted with F27 and R1492 primers and finally PCR product was cloned in pTZR/T57 (Fermentas). Bacterial isolate was identified as Serratia marcescens by 16S rRNA gene sequencing and biochemical tests. The bioassay was conducted on hibernating adults using topical application. Bioassay results showed that the concentration of 10⁸ CFU caused 60% mortality 3 days postexposure. Contact bioassay also led not only to change in sunn pest color to the pink-red, but also the insect cuticle seems to be degraded. Although S. marcescens cells could not use as a biological control due to their pathogenicity toward vertebrates, the produced toxic compounds could utilize in integrated sunn pest management.

Developing a novel virus biopesticide to target major lepidopteran pests

MCn02 •260•



Freya Scoates¹, Annabel Rice², Ken Wilson², David Grzywacz³, Igor Curcic¹, Aoife Dillon¹

¹Exosect Ltd, Leylands Business Park, Colden Common, Hampshire, SO21 1TH, UK; ²Lancaster Environment Centre, Lancaster University, Lancaster, LA1 4YQ, UK 3Natural Resources Institute, University of Greenwich, Central Avenue, Chatham Maritime, Kent ME4 4TB, UK Corresponding author: freya.scoates@exosect.com

The biopesticide market is estimated to be worth \$2.3 billion¹, and expected to increase to \$6.6 billion by 2020². Baculoviruses are a category of biopesticides routinely used against lepidopteran pests, including Diamondback Moth (Plutella xylostella), Egyptian Cotton Leafworm (Spodoptera littoralis) and Codling Moth (Cydia pomonella). Biopesticides have many advantages over chemical pesti-cides: registration costs tend to be lower, a narrower host range reduces the scope for non-target effects, and they allow growers to meet consumer demands for reduced chemical inputs in food production. However, baculo-viruses are highly sensitive to UV degradation, with significant reductions in efficacy observed within 4-24 hrs

of application in full temperate sunlight³. As such, applications need to be precisely timed and repeated applications are often required to achieve control of multiple generations. A consortium of business and academic partners received funding from the UK's Agritech Catalyst fund, administered by Innovate UK, the UK's innovation agency, to investigate whether Exosect's proprietary technology Entostat® can be used to address this constraint. Entostat® is based on electrostatic micronized waxes, which delivers increased loading of active ingredients to the target surface. In addition to potentially improving delivery of virus onto leaf surfaces, it is hypothesized that by enveloping the virus in the wax, Entostat® can be used to extend the field life of the virus if the wax acts as a UV filter. Initial formulation work has confirmed that the envelopment process does not impair the efficacy of the virus. In fact, test insects (2nd-instar Spodoptera littoralis) have been shown to be significantly more susceptible to enveloped virus than unformulated material (LC50 of 1.8 x 10^4 OB/mL versus 3.5 x 10^4 OB/mL) in bioassays. The plans for testing UV stability will be discussed. REFS: Global Industry Analysts, Inc., 2015; ² Markets and Markets, 2015; ³ Ignoffo, Rice and McIntosh, 1989.

Effect of optical brighteners on ChinNPV infectivity on MCn03 •261• Chrysodeixis includens caterpillars in bioassays and greenhouse

Alessandra Benatto^{1,2}, Maria Cristina Neves de Oliveira², Suely Ruiz Giolo³, Vanessa F. Sehaber³, Gizele Rejane Baldo⁴, Jhibran Ferral Piña⁵, **Daniel Ricardo Sosa-Gómez**^{1,2}

¹ Pós-Graduação em Entomologia, Universidade Federal do Paraná, Curitiba, PR, Brazil; ² Embrapa Soja (CNPSo), Londrina, PR. Brazil: ³ Departamento de Estatística, Universidade Federal do Paraná, Curitiba, PR, Brazil; ⁴ Universidade Estadual de Ponta Grossa, Ponta Grossa, PR, Brazil; ⁵ Instituto Nacional de Investigaciones Forestales Agricolas y Pecuarias (INIFAP), Quintana Roo, México

Corresponding author: daniel.sosa-gomez@embrapa.com

Virus-based insecticides have been used as biological pest control in several countries/regions around the world such as Australia, Brazil, China, India, and sub-Saharan Africa, mainly due to their specificity and safety to non-target organisms. However, their acceptance as bioinsecticides is limited because of their relatively slow action; in some cases, high doses are required, which implies higher productions costs. To circumvent these bottlenecks and improve Chrysodeixis includens nucleopolyhedrovirus (ChinNPV) performance, synergistic agents were studied. This work aimed to evaluate effective ChinNPV/brightener combinations to control soybean looper, Chrysodeixis includens. Therefore, three concentrations of optical brighteners (OP) [Fluorescent Brightener 28 (FB28); 4,4'-diamino-2,2'-stilbenedisulfonic acid and Calcofluor White Stain, ranging from 0.025% to 1.5%] in combination with ChinNPV 5.0 × 10³ OB.mL⁻¹ were incorporated into the artificial diet, without formalin and antibiotics, when the diet temperature reached 56°C. Early 3rd instar loopers feeding on the diet *ad libitum* were kept in trays in environmental chambers (26 \pm 2°C, 70 \pm 10%, 14-h photophase). In a greenhouse test, soybean plants were spread with: ChinNPV 1.5×10^{12} OB.ha⁻¹; ChinNPV 3.0×10^{12} OB.ha⁻¹; ChinNPV 1.5×10^{12} OB.ha⁻¹ + FB28 (0.1%); and ChinNPV 3.0×10^{12} OB.ha⁻¹ + FB28 (0.1%). Leaflets were collected immediately and 96 h after applications and offered to soybean loopers. Mortality was assessed daily until the 12th day. Data were submitted to the Competitive Risk Survival Analysis, following the Fine & Gray model, to compare curves of cumulative incidence of mortality. In lab bioassays, the addition of FB28 (0.1%) greatly increased the activity of ChinNPV, decreasing mean lethal time from 8.45 to 5.93 days and increasing mortality from 42% $\,$ to 99%. In greenhouse, no significant differences were observed among the curves of cumulative incidence of mortality among the highest virus doses with or without OP. However, at the dose of ChinNPV 1.5×10^{12} OB.ha⁻¹ a significant lower cumulative probability of death was observed. The most efficient brightener was FB28, which did not have any negative effects against C. includens larvae when used alone.

Effect of Bacillus thuringiensis products on larval survival and consumption of Plutella xylostella

Nathália A. dos Santos¹, Dagmara G. Ramalho¹, Valéria L. de Laurentis², Vanessa F. P. de Carvalho², Alessandra M. Vacari², Sergio A. De Bortoli²

> ¹Biology Department, Entomology Post Graduated Course FECLRP-USP. Ribeirão Preto, SP, Brazil. ²College of Agrarian and Veterinary

Sciences-FCAV/Unesp. Via de Acesso Prof. Paulo Donato Castellane s/nº. CEP 14884-900, Jaboticabal, SP, Brazil

Corresponding author: nattysantos19@hotmail.com

Bacillus thuringiensis-based bio-insecticides stand out as biological agents against insect pests, mainly because they have specificity. These products can efficiently control *Plutella xylostella*, the main pest of Brassicaceae. The objective of the present work is to analyze the sublethal effects of B. thuringiensis (Agree®, Dipel® and Xentari®) based products, analyzing the leaf consumption by larvae as well as larval survival. The products concentrations that caused 25% mortality (CL25) in insects of two populations, susceptible-PS and resistant-PR, was tested. Autoclaved deionized water plus 0.5µl / mL of Triton-X100® was used as the control. In the bioassay, 10 P. xylostella second instar larvaes were used placed on leaf kale disks previously immersed in the different treatments. Each disc constituted a replication, being evaluated ten replications per treatment. For the larvae of the susceptible population, the highest leaf area consumed (13.8 cm²) was observed in the control, compared with Dipel® (9.2 cm2) and Xentari® (9.8 cm2), but not differing from Agree® (12, 0 cm 2). For larvae of the resistant population there was no difference between the bioinseticides in relation to the foliar consumption. It should be pointed out that, in general, the insects of the resistant population always presented greater consumption of leaf area. About the larvae survival rate in the susceptible population, all treatments were different from the control, with survival percentages being, respectively, 26.0%, 38.0%, 26.0% and 54.0% for Agree®, Dipel®, Xentari® and control. However, in the larvae survival in the resistant population, Agree® (28.7%) differed from Xentari ® (52%), being similar to Dipel® (41.3%) and to the control (44.7%). The results indicate that leaf consumption and larval survival are biological characteristics showing sublethal effects of the bioinsecticides in the two populations of *P. xylostella*.

Characterization of newly isolated Iranian Bacillus thuringiensis strains based on gene content, proteomic analysis and insecticidal activity

MCn05 •265



Ayda Khorramnejad^{1,2}, Reza Talaei-Hassanloui² Vahid Hosseininaveh², Yolanda Bel¹, **Baltasar Escriche**¹

¹Departamento de Genética/ ERI BioTecMed. Universitat de València. C/ Dr. Moliner, 50 46100-Buriassot, Comunidad Valenciana, España: ²Departament of Plant Protection. College of Agriculture and Natural Resources, University of Tehran, Karaj, 31587-77871, Iran. Corresponding author: baltasar.escriche@uv.es

Bacillus thuringiensis (Bt) is one of the most useful entomopathogens used nowadays for biological pest control. The identification of new Bt strains is a topical issue in order to discover novel Bt toxins to broaden the range of insecticidal activities and to manage better possible insect resistance problems. Seven Bt strains isolated from infected larvae and soils from Iran (IE-1, AzLp, IE-2, IP-2, IEp, RM and KhF strains), have been characterized. The spectrum of lepidopteran toxic activity of these Bt strains, was assessed against Plodia interpunctella, Spodoptera exigua, Mamestra brassicae, Grapholita molesta, Ostrinia nubilalis and Helicoverpa armigera neonate larvae. The studied Bt strains showed partial and total similarity in their protoxin and trypsinized protein profiles respectively, but they differed in their insecticidal activity spectrum. G. molesta and O. nubilalis were the most susceptible insects, and AzLp, IE-2 and IP-2 exhibited high levels of toxicity against S. exigua and M. brassicae compared to those of the reference strain, HD-1. LC-MS/MS analysis of solubilized crystal proteins was used to reveal the major proteins composing the parasporal crystal. In general, Cry1A family proteins were the most represented and abundant insecticidal crystal proteins in these strains. These results were compared with the ones obtained after PCR screening of Cry genes. As a final step in the strain characterization, the production of β -exotoxin was also checked and no type I β-exotoxin was detected in the studied strains. Results suggested that different complementary approaches are necessary for a robust characterization of new Bt strains, in order to understand their possible use as bioinsecticides. Due to the characteristics of AzLp, IE-2 and IP-2 strains, they could be considered as good candidates for lepidopteran pest control.

MCn04 •262•

Field efficacy of Bacillus thuringiensis galleriae strain SDS-502 for the management of alfalfa weevil and the impact on Bathyplectes spp. parasitization rate

MCn06 •264•

microbiome and its variation among habitats

MCn08 •266

Govinda Shrestha¹, Gadi V.P. Reddy¹, Stefan T. Jaronski²

¹Department of Research Centers, Western Triangle Agricultural Research Center, Montana State University, Conrad MT 59425 USA; ² USDA-ARS-PARL, 1500 N. Central Ave. Sidney MT 59270 USA Corresponding author: govinda.shrestha@montana.edu

The alfalfa weevil, Hypera postica (Gyllenhal) (Coleoptera: Curculionidae), is considered as one of the most serious pests of alfalfa in US and worldwide. The damaging populations are frequently found in Northern Plains alfalfa. Chemicals are registered for control of alfalfa weevils in US. However, there is increasing pressure on conventional alfalfa growers to minimize the chemicals use for alfalfa weevil management because of negative impacts on non-target organisms including pollinators and natural enemies of alfalfa weevils. In addition, organic alfalfa production has increased over the years, especially in hay exported to Asia. Recently a new beetle-active Bacillus thuringiensis (Bt*) has been commercialized for use against a number of beetles, including the alfalfa weevil. Most commercial coleopteran Bts have a beetle-specific CRY3 family protein. This new Bt, B. thuringiensis galleriae strain SDS-502, has a novel toxic protein, CRY8Da. This study was, therefore, aimed to 1) evaluate the efficacy of this Bt strain against the H. postica larvae under Montana field conditions and 2) to determine of Bt impact on parasitization rate of Bathyplectes spp. against H. postica larvae. The field study was conducted at Montana two locations (Conrad and Valier) in 2016. The result indicated that this Bt strain provided 11 to 60 percent reduction of larval populations depending on the labeled lower and high rates. Interestingly, this Bt product has no negative impact on Bathyplectes spp. populations.

Entomopathogenic Bacillus thuringiensis as biological control agent for complex disease (tomato Fusarium wilt and root knot nematode)





Jiaheling Qi 1.2, Naoki Takahashi 2, Daigo Aiuchi 2, Shin-ichiro Asano³; Masanori Koike²

¹ The United Graduate School of Agricultural Sciences, Iwate University, Iwate Japan; ² Department of Agro-environmental Science, Obihiro University of Agriculture & Veterinary Medicine, Obihiro, Japan; ³ Department of Applied Bioscience, Graduate School of Agriculture, Hokkaido University, Sapporo, Japan Corresponding author: koike@obihiro.ac.jp

In tomato production, the complex disease caused by Fusarium wilt (pathogen: Fusarium oxysporum f.sp. lycopersici, FOL) and root knot nematode (Meloidogyne incognita, RKN) is a severe problem, and it also causes huge economic loss. We already showed the plant growth promoting effect, biofilm formation ability, nematicidal effect of Bacillus thuringiensis strains. This study was estimated to use B. thuringiensis as an alternative agent to control the development of complex disease. Three strain; B. thuringiensis japonensis (BT17), B. thuringiensis kurstaki (BT18), B. thuringiensis CR371-H (BT20), Tomato (Solanum lycopersicum) cultivar Kyoryoku-Beiju(Takii Seed Co.) ,FOL and RKN were used for this experiment. Surface sterilized tomato seeds were grown at $25\pm1^{\circ}$ C in a greenhouse. After 4-week cultivation, tomato seedlings were treated with vegetative spore at 10 ml/pot (3×10^8 cfu/ml). After 7 days, 5ml J2 suspensions were inoculated around plant roots (500 J2s/pot). After 7 days, the bacterianematode treated tomato seedlings were placed in the FOL-infested soil. 4 weeks after pathogen inoculation, symptom expressions of the complex disease were evaluated. 11 experiment groups were set: 3 BT strains (BT17) BT18、BT20) + FOL; 3 BT strains (BT17、BT18、BT20) + RKN; 3 BT strains (BT17、BT18、BT20)+ FOL + RKN; RKN control; FOL control; FOL+ RKN control; distilled water untreated control. As a result, B. thuringiensis strains could significantly suppress the symptoms of complex disease, decrease gall formation on the root, wilting score and internal symptom.

A molecular survey of the wireworm

Todd Kabaluk¹, Erica Li-Leger²

¹Agriculture and Agri-Food Canada, Agassiz, BC: ²Department of Zoology, University of British Columbia, Vancouver, BC Corresponding author: Todd.Kabaluk@aar.ac.ca

Wireworms are a serious subterranean larval pest of agricultural crops for which entomopathogenic fungi are being developed and commercialized as pest control agents. How bacteria interact with fungal pathogens needs elucidation as interactions between microbes and their hosts can have major impacts on host behavior and fitness. DNA sequencing and computational biology have allowed for comprehensive genetic surveys of an organism's microbiome and can be done with relative ease, revealing taxa that might not be apparent from traditional isolation techniques such as plating. Identifying the bacteria associated with wireworms is the first step toward elucidating organismal relationships which may be invaluable for creating more effective pest control strategies. The bacterial microbiome from five separate wireworm (Agriotes obscurus and A. lineatus) populations (sites with different habitats) was sequenced, revealing 609 operational taxonomic units (OTU) among the total of individuals. The most common OTUs at each site exhibited a diverse range of bacterial groups, with the proportions of 40 genera being significantly different among the five collection sites. Principle coordinate analysis revealed clustering of two sites, indicating a difference in the composition of taxa and suggesting that habitat can influence a microbiome's members. The implications this may have on wireworm microbial control is under investigation, with symbiosis / disease resistance associated with certain taxa being an area of interest.

Feeding diel rhythm and insulin receptor expression of Maruca vitrata

MCn09 2670

Yonggyun Kim

Department of Plant Medicals, Andong National University, Andong 36729, Korea Corresponding author: hosanna@anu.ac.kr

The legume pod borer, Maruca vitrata, infests and causes serious economic damage on the Adzuki bean, Vigna angularis, but does not on other leguminous beans in Korea. However, a laboratory analysis of the larval growth of M. vitrata indicated that other beans are likely to be optimal hosts. For the purpose of the molecular interaction between host and growth in M. vitrata, this study aims to investigate an insulin signal of M. vitrata, Injection of a bovine insulin significantly changed growth rate along with feeding amount of 4th instar larvae. A transcriptome analysis using RNA-Seq provides genes predicted to be associated with insulin signal. Larval feeding rhythm was monitored every 3 h and indicated that most feeding occurred at early photophase. Starvation induced expression levels of 7 insulin signaling genes by up-regulating insulin receptor (Mv-InR) but down-regulating other 6 genes. Mv-InR encodes 1,213 amino acids and contains both functional sites of ATPbinding and tyrosine kinase. Expression of Mv-InR was confirmed in all developmental stages from egg to adults. RT-qPCR analysis showed that Mv-InR expression varied with feeding rhythm. The larvae fed with different leguminous diets showed variation in expression levels of Mv-InR. These results suggest that M. vitrata may infest other types of leguminous crops in

MCn10 •268• Molecular detection, characterization and biological evaluation of Red Palm Weevil (Rhinchophorus ferrugineus) associated fungal entomopathogen (Metarhizium anisopliae) from Pakistan

> Jam Nazeer Ahmad^{1,2}, Mujahid Manzoor^{1,2}, Samina Tanwir Ahmad^{1,2}, Muhammad Jafir¹, Samar Abbas Naqvi³, Iqrar Ahmad Khan

¹Intergrated Genomics Cellular Developmental and Biotechnology Lab (IGCDB), Department of Entomology, University of Agriculture Faisalabad, Pakistan; ²Plant Stress Physiology and Molecular Biology Lab, Department of Botany, University of Agriculture Faisalabad, Pakistan; ³Department of Horticulture, University of Agriculture Faisalabad, Pakistan Corresponding author: jamnazire@yahoo.com

Red Palm weevil (Rhinchophorus ferruginous) is an important quarantine and destructive pest of date palm worldwide. It feeds inside the plant and early

detection is very difficult. Different strategies have been adopted including application of insecticides but the use of microbial pesticides has obtained a high level of significance. In the present work, laboratory trials were conducted to identify entomopathogens associated with red palm weevil and to investigate their effect against various life stages of this pest. Molecular markers and specific primers can be used that provide a means for the molecular identification, diversity and link to virulent isolates to screen different entomopathogens. Healthy and infected Red palm weevil adults and larvae were collected from different regions of Pakistan during a survey conducted 2016-2017. Some samples of fungal infected Red palm weevil were collected and brought to laboratory in 100% ethanol for their molecular identification. Fungal DNA was extracted from infected samples and amplified in PCR machine by using PCR-based RAPD markers. RFLP analysis with restriction enzymes and sequencing was also performed. Our results of molecular study showed that Metarhizium anisopliae was found on dead samples of Red palm weevil. We reared Red palm weevil in our laboratory on artificial diets. Bioassays were conducted by using adults and 2nd, 5th, 10th instars larvae of Rhinchophorus ferrugineus in order to know their pathogenicity. The data regarding various doses of Metarhizium and correspondent mortality (LD50/LC50) was recorded after every 24h till pupation, adult emergence and egg eclosion. The analysis was done by using Probit Analysis for all the bioassays carried out under laboratory condition (25±2°C and 75% r.h). We observed high mortality of Red palm weevil in laboratory trial with our native isolate of Metarhizium anisopliae. To our knowledge, this is the first report of molecular identification, characterization and biologic evaluation of native fungal entomopathogen

Production of secondary metabolites from Azadirachta indica and their efficacy against Sf9 cell cultures

(Metarhizium anisopliae) against Red Palm weevil from Pakistan.

MCn11 •269•

Anant V. Patel¹, Peter Spieth¹, Hanna Bednarz², Ina Kleeberg³, Karsten Niehaus²

Bielefeld University of Applied Sciences, Faculty of Engineering Sciences and Mathematics, Bielefeld, Germany; ²Bielefeld University, Bielefeld, Germany; ³Trifolio-M GmbH, Lahnau, Germany Corresponding author: anant.patel@fh-bielefeld.de

Secondary metabolites from neem (Azadirachta indica A. Juss) show a broad spectrum efficacy against insect pests including insecticidal, anti-feedant or insect repellent activities. To date, less is known about the mode of action of these active compounds and their production is still based on complex extraction procedures. Several studies have shown that plant cell cultures of A. indica can produce active secondary metabolites like azadirachtin A. In light of increasing insight into plant cell cultures from A. indica it can be hypothesized that the use of elicitor agents combined with an adjustment of the culture conditions are beneficial for the production of active neem compounds. Furthermore there is evidence that a rearrangement of the natural compound mixture can increase the efficacy against insect pests. The overall aim of this project is to develop a competitive process to produce high concentrations of bioinsecticidal compounds with either neem endophytes or neem plant cell cultures. Besides we want to shed some light into the mode of action of some neem compounds accompanying azadirachtin A. Here, we present data on production of active compounds with plant cell cultures as well as first results of novel compound mixtures tested against Sf9 cell cultures. First, we induced plant cell cultures from various plant tissues with a medium that allows a callus proliferation from more than 90 % of the explants. Furthermore we analyzed the mode of action of different neem ingredients against insect cell lines. We report production of azadirachtin A obtained with a novel high-throughput microbioreactor system (BioLector® Pro from m2p-labs GmbH) that enables 48 parallel batch cultivations in a volume under 1 ml. In addition to that we will describe how different plant hormones can be used as elicitor agents in neem cell cultures. Finally, we will present data on fluorescence and confocal laser scanning microscopy as well as FACS analysis of treated Sf9 cell cultures that will lay the basis to determine how active compounds influence the development of insect cells.

The project PICTA-KILL - Novel strategies for biological psyllid pest control

MCn12 •270•



Anant V. Patel¹, Linda Muskat¹, Pascal Humbert¹, Jürgen Gross², Louisa Görg², Cornelia Dippel³, Elisa Beitzen-Heineke⁴, Wilhelm Beitzen-Heineke⁴, Michael Przyklenk⁴

¹ Bielefeld University of Applied Sciences, Faculty of Engineering Sciences and Mathematics, Bielefeld, Germany; ²Julius Kühn Institute (JKI), Dossenheim, Germany; ³IS Insect Services GmbH, Berlin, Germany; ⁴BIOCARE GmbH, Einbeck, Germany Corresponding author: anant.patel@fh-bielefeld.de

Psyllid pests are distributed all over the world and cause damage in crop plants. Novel defense strategies against these insect pests are of international interest. The use of entomopathogenic fungi as biocontrol agents for reducing psyllid pest populations represents a plant protection method of low risk for nature and humans as well. Being the vector of Candidatus Phytoplasma mali, the infectious agent of apple proliferation, the psyllid Cacopsylla picta is responsible for an annual economic loss of a threedigit-million range in Europe. Because there are no direct measures to combat apple proliferation, the vector itself has to be controlled in order to protect the plants. In Germany, there are no authorized plant protection products available, neither for organic farming nor for conventional farming. New insights into the scent preferences of *C. picta* offer innovative options for its control. Hence, the aim of this project is to develop formulations, which can be applied for Attract-and-Kill-strategies against C. picta. Furthermore, it will be examined if a combination with repellent agents supports the effect (Push-Pull-Kill- strategy). In laboratory and field trials new repellent, attractive and arresting substances, entomopathogenic microorganisms and other insecticides effective against C. picta as well as formulation materials and methods for these active ingredients will be screened. For the entomopathogenic microorganisms, cost efficient mass production processes will be developed. Appropriate formulations will be developed and tested in laboratory and field trials. This joint projects cooperates with the Eilenberg group of Copenhagen University on entomopathogenic fungi. First results of the recently started project on strain selection, cultivation, identification of semiochemicals and formulation will be presented.

Development of a biological tick control agent based on an innovative attract-and-kill strategy (BIOZEC)

MCn13 •271•

Anant V. Patel¹, Sissy-Christin Lorenz¹, Pascal Humbert¹, Marion Wassermann², Ute Mackenstedt², Michael Przyklenk³, Elisa Beitzen-Heineke³, Wilhelm Beitzen-Heineke³, Kerstin Büchel⁴. Hans Dautel⁴

¹ Bielefeld University of Applied Sciences, Faculty of Engineering Sciences and Mathematics, Bielefeld, Germany; ²University of Hohenheim, Department for Zoology, Dept. Parasitology, Hohenheim, Germany; ³BIOCARE GmbH, Einbeck, Germany; ⁴IS Insect Services GmbH, Berlin, Germany Corresponding author: anant.patel@fh-bielefeld.de

Ticks are vectors for a multitude of pathogens, causing e.g., Lyme disease and tick-borne encephalitis in many parts of the Northern hemisphere. In Germany 8-10 million people suffer from tick bites every year whereby most bites are caused by Ixodes ricinus. At present, there is no individual control measure against ticks available. The overall aim of this project is the development of a novel biological control agent against ticks based on an innovative attract-and-kill strategy. The basis of this control agent is the attractive effect of carbon dioxide (CO2) combined with tick-specific attractants and substances causing aggregation in ticks (e.g., aggregation pheromones), slowly released into the vicinity by a specially tailored biopolymer bead. The attractive bead is combined with a kill component, an entomopathogenic fungus, isolated from indigenous dead ticks in Germany. The fungus infects the tick as it comes in direct contact with the bead and finally leads to death by growing through the cuticle into the tick's hemolymph. As the ticks are attracted by the beads, a high-dose application can be avoided. Furthermore, all components are biodegradable so that the environmental pollution is minimized. First experiments dealt with the encapsulation of baker's yeast in Ca-alginate beads to achieve a sustained release of CO₂ above ground. These beads significantly attracted *I. ricinus*

nymphs. Current investigations are focused on the selection and cultivation of virulent strains of entomopathogenic fungi from the genus Metarhizium, biomass encapsulation as well as drying and rehydration of beads. Moreover, a tick specific arrestant was successfully coated on the bead's surface which proved to have an aggregating effect for I. ricinus. This work will pave the way for novel tick control strategies based on an attract-and-kill approach.

The project ATTRACAP: Optimization of an attract-and-kill strategy for wireworm control in potato

MCn14 •272•



Anant V. Patel¹, Katharina Hermann¹, Pascal Humbert¹, Michael Przyklenk³, Elisa Beitzen-Heineke³, Wilhelm Beitzen-Heineke³, Stefan Vidal²,

> ¹Bielefeld University of Applied Sciences, Faculty of Engineering Sciences and Mathematics, Bielefeld, Germany; ²Agricultural Entomology, Department for Crop Sciences, Georg-August University Goettingen. Goettingen, Germany; 3BIOCARE GmbH, Einbeck, Germany Corresponding author: anant.patel@fh-bielefeld.de

Wireworms, the polyphagous soil-dwelling larvae of click beetles (Coleoptera: Elateridae), are a major insect pest of worldwide relevance causing tremendous yield losses in several crop production systems, like potatoes, wheat and other grain crops, oil crops as well as vegetables. Within the last years wireworm damage in potato production substantially increased, both in conventional and organic production systems, resulting in existencethreatening yield losses for farmers. Currently, effective plant protection strategies are not available due to the progressive phasing-out of effective synthetic chemicals. Based on previous projects, an innovative and effective control strategy was developed, using solely biological components. A capsule releases carbon dioxide (CO₂) upon contact with the soil humidity, which attracts the wireworms towards these capsules, where they come into contact with an isolate of the entomopathogenic fungus Metarhizium brunneum, thus substantially enhancing the frequency of fungal infections of wireworms. The project aims at improving the efficacy levels of the in previous projects developed product candidate ATTRACAP®. The innovative formulation technology and resulting beads will be fine-tuned and tested and validated under varying field conditions. Apart from the academic partners, members of the project consortium comprise the company producing the product, and consultants which stay in contact with the farmers and will help with set-up of field experiments. As an outcome of the project an optimized product ATTRACAP® will be available, helping both conventional and organic farmers to maintain a sustainable potato production. This poster will present the first results of the recently started project including field trials 2017. Our formulation will pave the way towards novel Attract-and-Kill strategies in other pest problems.

Various roles of mite pathogenic fungi against Tetranychus urticae (two-spotted spider mite)

MCn15 •273•



JiHoon Lee¹, HwiGeon Yun¹, Dong Jun Kim¹, JiIn Ma¹, WonSeok Gwak¹, SooDong Woo¹

¹Department of Agricultural Biology, College of Agriculture, Life & Environment Science, Chungbuk National University, Cheongju 28644, Korea Corresponding author: sdwoo@cbnu.ac.kr

The two-spotted spider mite, Tetranychus urticae, and gray mold disease caused by Botrytis cinerea, are important pest and plant disease, respectively, against fruit, vegetable and other plants in the world. To control these, many people have been relied on chemical methods for a long time. However, these are not always effective and their continuous use has resulted in resistance of pest and disease. Thus, many researchers have studied to overcome these problems. Recently, entomopathogenic fungi have been shown to have multiple roles as endophytes, phytopathogen antagonism, rhizosphere and plant growth promoter. Therefore, we performed to explore the various roles of mite pathogenic fungi. The antifungal activities of the selected fungi with high virulence to two-spotted spider mite were tested to explore the potential for the dual control of B. cinerea as well as T. urticae. Antifungal activities against the fungus B. cinerea were evaluated by dual culture assays using both aerial conidia and cultural filtrates of mite pathogenic fungi. As a result, two fungal isolates were selected considering both the virulence to mite and antifungal activity. To maximize the use of spores and cultural filtrates, the virulence to mite was further evaluated

using the cultural filtrate, blastospore and aerial conidia. Consequently, the 2 fungal isolate selected in this study were confirmed to have diverse potential and would be used effectively for dual control agents against the twospotted spider mite and plant diseases.

STU MCn16 •274• Diverse activities of entomopathogenic fungi with the virulence against Myzus persicae (green peach aphid)

HwiGeon Yun liHoon Lee Dong lun Kim Jiln Ma, WonSeok Gwak, SooDong Woo

Department of Agricultural Biology, College of Agriculture, Life & Environment Science, Chungbuk National University, Cheongju 28644, Korea Corresponding author: sdwoo@cbnu.ac.kr

The green peach aphid, Myzus persicae, and Botrytis cinerea, causing gray mold, are major pest and phytopathogens, respectively, affecting vegetables and fruit crops in greenhouses and fields all over the world. To control this aphid and disease, farmers have depended on the use of chemical insecticides or fungicides, and their concentrated use over many years has led to the development of resistance. Recently, the potential of entomopathogenic fungi with multiple roles has been verified as endophytes in plants, phytopathogen antagonism, rhizosphere and plant growth promoter. Therefore, in this study, we tested these various roles of entomopathogenic fungi as an effective alternative to control simultaneously pest and plant disease. The antifungal activities of the selected fungi with high virulence to green peach aphid were investigated to explore the potential for the dual control both M. persicae and B. cinerea. Antifungal activities were evaluated to plant pathogenic fungi B. cinerea by dual culture assay on PDA media and 96 well plate using fungi cultural filtrates. Two fungal isolates were selected finally based on insecticidal and antifungal activities. For the usefulness of dual control, the virulence to aphid was further tested using the cultural filtrate, blastospore and aerial conidia of selected isolates. These fungal isolates selected in this study would be used effectively as the dual control agents of green peach aphid and gray mold.

A search for novel Metarhizium isolates for control of Rhipicephalus microplus ticks

MCn17 •275•



Patrícia S. Golo¹, Thais A. Correia¹, Caroline F. Pereira¹, Danilo M. Akiau¹, Donald W. Roberts², Vânia R.E.P. Bittencourt¹

> ¹Departamento de Parasitologia Animal, Instituto de Veterinária Universidade Federal Rural do Rio de Janeiro, Seropédica, RJ, Brazil. ²Department of Biology, Utah State University, Logan, UT, USA Corresponding author: patriciagolo@gmail.com

Inappropriate use of chemical acaricides for control Rhipicephalus microplus ticks (cattle ticks) cause both economic losses and human health risks due to meat, milk and environmental contamination. Control of R. microplus ticks with fungi is a promising choice as it involves low risk to human health and relatively low cost. Unfortunately, bioassays in the laboratory routinely demonstrate that entomo-pathogenic fungi (EPF) are considerably less virulent to ticks than to insects. Hence, to seek Metarhizium isolates with likely histories of infecting ticks, we conducted isolation of Metarhizium from sites where a cattle tick population occurred. Metarhizium from soil tend to be well adapted to this environment, sometimes as plant endophytes, and may be highly persistent in the field. Biological assays with fungi to control ticks are frequently performed with isolates previously deposited in fungal collections; thus, in most published studies, tick population and the fungal isolates to which they are exposed have no geographical or time congruence, which may explain a routinely low tick-control rates reported. Accordingly, the present study aimed to I) isolate Metarhizium fungi from soil samples collected close to the tick population to be tested (named FS); II) exam fungal isolates' heat tolerance to select highly tolerant strains, and III) assay these isolated fungi against FS tick larvae. The samples were examined as to their physical and chemical characteristics, and fungi were then isolated from this soil using CTC artificial selective medium (Fernandes et al. 2010). Heat tolerance tests were performed by exposing conidia to 42°C for four hours and relative germination was calculated after 24 h incubation; a well understood M. robertsii isolate (ARSEF 2575) was used as standard in thermotolerance tests. Nine isolates (named LCMS01 to LCMS09) were obtained from 100 soil samples. Isolate LCMS04 showed exceptionally high

thermotoler-ance (over 50% germination). Bioassay showed LCMS02, LCMS04 and LCM06 had the highest larval death 5 days after fungal exposure, reaching 91.9%, 85.3% and 95% average tick larval mortality. Accordingly, LCMS04 was selected for further studies as a promising new agent for control of cattle ticks in the field.

Enhancement of Beauveria bassiana virulence and efficacy to control Diatraea saccharalis in a sugarcane crop for panela production

Mcn18 •276•

Juliana Gómez-Valderrama¹. Lorena García Riaño¹. Diana Marcela Monroy¹, Gustavo Adolfo Araque¹, Carlos Espinel¹, Laura Villamizar^{1,2}

¹Corporación Colombiana de Investigación Agropecuaria, Centro de Investigación Tibaitatá. Mosquera, Colombia; ²AgResearch Ltd. Lincoln Research Centre. Christchurch 8140, New Zealand Corresponding author: jagomez@corpoica.org.co

The crop of sugarcane for panela production is of great importance in Latin America, Central America and India. Panela yield depends on sucrose content in sugarcane, characteristic seriously affected by the presence of the stem borers complex of the genus Diatraea sp., difficultly controlled by chemical insecticides. A Colombian isolate of Beauveria bassiana (Bv062) demonstrated high potential to develop a bioinsecticide for the control of Diatraea sp. The present work aimed to evaluate potential virulence enhancers for fungal conidia and determine efficacy of formulated conidia under laboratory and field conditions. Three potential enhancers (glucose, Trichoderma sp. enzymatic extract and sodium lignosulfonate) were mixed with Bv062 conidia and evaluated to control D. saccharalis larvae. Glucose and an enzymatic extract increased three times the insecticidal activity of the conidia. Conidia were formulated using two delivery systems: emulsifiable concentrate (EC) and dispersible granulate (WG). Mean lethal concentration estimated in a laboratory bioassay was 1x10⁶ con/mL for both prototypes and this value decreased to $4x10^5$ con/mL, when conidia enhanced with Trichoderma sp. extract. The EC formulation presented the highest in plant efficacy under semi-controlled conditions and was selected to be evaluated in the field at three different application rates. Significant differences were not detected between doses, with 93% efficacy obtained with the lowest dose (1x10¹² con/ha). Results demonstrated the high potential of the isolate Bv062 to control the predominant pest in sugarcane crops for panela production.

Selection of Metarhizium sp isolates based on thermotolerance, endophytic colonization and a gene expression assay

MCn19 •277•

Allan F. Marciano 1,2, Deborah E. Henderson 1, Patrícia S. Golo², Vania R. E. P. Bittencourt ² Athena Williamson¹, Lisa Wegener¹

¹ Institute for Sustainable Horticulture, Kwantlen Polytechnic, University, Langley, Canada: ²Instituto de Veterinária Universidade Federal Rural do Rio de Janeiro, Seropédica, Brasil Corresponding author: allanfmarc@gmail.com

Entomopathogenic fungi can be endophytic-symbiotic with plant species, and can act against host plant pests. Their bi-functionality is characterized by the expression of different genes, and resistance to biotic, and abiotic stress factors. The aim of the present research was to study the thermotolerance. endophytic colonization of mung bean plants, and expression of the gene adhesin-like protein 2 (Mad2), in order to select Canadian isolates for field use. Metarhizium sp, isolates from soil of the South West region of British Columbia, Canada, were used for the thermotolerance experiment. Suspensions of 10⁵ conidia/ ml of each isolate were placed in water-baths at 40°C, 45°C, and a control temperature of 25°C, for 2, 4, 8, and 12h. Germination was evaluated after a 48h incubation in the dark at 28 ± 1 °C. For the endophytic colonization trial and gene expression analyses, four isolates were used: ISH-CC-90, ISH-CC-277, ISH-CC-188, and ISH-CC-133. Mung bean seeds were treated by immersion in fungal suspensions of 10⁶ conidia/ml of the respective isolates. Plants were treated with suspensions of 10⁸ conidia/ml as a foliar spray, and a soil drench. The controls for both treatment applications were water. Two weeks later, plants were harvested and their leaves, stems, and roots were sampled to evaluate endophytic fungal colonization. After passage of the isolates through the plant, the

relative expression of the gene Mad2 was profiled by qPCR, and compared with colonies cultivated in PDAY medium. Among the isolates submitted to the thermotolerance test, two showed more than 94% germination after 12h of heating at 40°C. For isolates exposed to 45°C for 2 hours, up to 73% germination was observed. All isolates tested for endophytic colonization were able to colonize all tissues analysed. The molecular results of the Mad2 gene expression are underway. Prior to field use it is essential to understand characteristics of new microbial agents and these results will be used in the selection of new isolates of Metarhizium sp., as well as contribute to the scientific knowledge about the ecology of this microorganism.

Lethality of the entomogenous fungus Beauveria bassiana Strain NI8 on Lygus lineolaris (Hemiptera: MCn20 •278•

Miridae) and its possible impact on beneficial arthropods

Maribel Portilla¹, Randall Luttrell¹, Gordon Snodgrass¹, Yu Cheng Zhu¹, Eric Riddick²

¹USDA-ARS South Insect Management Unit, 141 Experiment Station Road, Stoneville, MS 38732; ²USDA-ARS National Biological Control Laboratory, 59 Lee Road, Stoneville, MS 38732 Corresponding author: maribel.portilla@ars.usda.gov

Bioassays were conducted to examine the pathogenicity of the fungus Beauveria bassiana (Balsamo) Vuillemin (Ascomycota: Hypocreales), strain NI8, against Lygus lineolaris (Palisot de Beauvois) and its impact on beneficial arthropods including Apis mellifera L., Crysoperla rufrilabris Burmeister, Orius insidiosus Say, Hippodamia convergens Guérin-Méneville, Harmonia axyridis (Pallas), Coleomegilla maculata De Geer, and field-collected Aranea spiders: Salticidae and Thomisidae. Insects were treated with four concentrations of NI8 $(3.9 \times 10^4, 2.3 \times 10^5, 4.2 \times 10^6 \text{ and } 1.5 \times 10^7 \text{ spores/mL})$ directly via topical spray. Median lethal concentration (LC₅₀), sporulation response (SR₅₀), and resistance ratio (RR₅₀) were estimated for all species except for the two groups of spiders. No significant differences in mortality (10-d after application) were observed among L. lineolaris, A. mellifera and C. rufilabris. All three species were highly affected when exposed to the highest concentrations of B. bassiana with 99.0%, 98.2%, and 90.0% mortality, respectively. Between 35 and 45% of the tested populations of O. insidiosus, H. convergens, field-collected crab spiders, and C. maculata were killed at 1.5 x 10⁷ spores / mL; whereas only 22% and 27%, respectively, of the fieldcollected jumping spiders and H. axyridis were killed at 10-d with the same concentration. No significant differences were found between the LC₅₀s measured for L. lineolaris and C. rufilabris 10-d after application. Results suggested that C. rufilabris would be highly affected by the NI8 strain of B. bassiana when applied for control of L. lineolaris. In contrast, B. bassiana appeared to have lower effects on the other beneficial arthropods assayed at the concentrations targeted for L. lineolaris control.

Entomopathogens in conjunction with imidacloprid could be used to manage wireworms (Coleoptera: Elateridae) on spring wheat

MCn21 •279•



Frank B. Antwi¹. Govinda Shrestha¹. Gadi V.P. Reddy¹, Stefan T. Jaronski²

¹Department of Research Centers, Western Triangle Agricultural Research Center, Montana State University, Conrad MT 59425 USA; 2 USDA-ARS-PARL, 1500 N. Central Ave. Sidney MT 59270 USA Corresponding author: reddy@montana.edu

The soil-dwelling larvae of click beetles (wireworms) (Coleoptera: Elateridae) are serious pests of several agricultural crops worldwide. Hypnoidus bicolor and Limonius californicus are two major wireworm species damaging to spring wheat, particularly in the Golden Triangle, an important cerealgrowing region in Montana. Wireworm damage to this field crop in the region is increasing, as currently available pesticides provide only partial control and no alternative options have been developed. We examined the effect of biopesticides alone, their mixtures or in conjunction with a conventional pesticide (imidacloprid) against wireworms. Biopesticides tested were: 1) spinosad Saccharopolyspora spinosa, (2) Metarhizium brunneum, (3) M. brunneum F52, (4) Beauveria bassiana GHA, (5) azadirachtin, (6) B. bassiana ANT-03 (7) pyrethrin, (8) Chromobacterium subtsugae, and (9) Burkholderia spp. strain A396. The efficacy of

Mic22 •280•

MCn23 •281•

biopesticides was based on crop stand protection, wireworm larval populations and grain yield. In the 2015 study, we found that entomopathogenic fungi alone or their combinations with imidacloprid had significant impacts on plant stand protection compared to water control. Applications of B. bassiana or a combination of M. brunneum F52 with imidacloprid protected wheat stand seedlings from wireworm damage better than water control at the Ledger location, while a combination of B. bassiana with M. brunneum F52 provided similar protection at the Valier location. Unexpectedly, wireworm larvae populations were found significantly higher on plots treated with B. bassiana, spinosad, M. brunneum F52 + spinosad, and M. brunneum compared to water control at 14 or 28 days post applications at the Ledger location, but without effects at the other location. We found significantly higher grain yield when plots treated with imidacloprid + M. brunneum F52 and B. bassiana + azadirachtin over water control at the Ledger location. In the 2016 study, there were no significant effects of treatments on studied parameters as compared to water control.

Performance of three Indian isolates of *Beauveria* bassiana (Balsamo) Vuillemin and three commercial mycoinsecticides against three developmental stages of *Bactrocera dorsalis* (Hendel) (Diptera:Tephritidae)

Mohammed I. Elbashir¹, P. Bishwajeet², K. Shankarganesh², P. Sharma, ³, *

¹Department of Bio pesticides and Bio fertilizers – Environment and Natural Resources and Desertification Research Institute, PO Box 6096, Khartoum, Sudan; ²Biological Control Laboratory, Division of Entomology, Indian Agricutlure Research Institute, New Delhi 110012; ³Division of Plant Pathology, Indian Agricultural Research Institute, New Delhi 11002, India *Corresponding author: fataloope@yahoo.com

Oriental fruit fly, *Bactrocera dorsalis* (Hendel) (Diptera: Tephritidae), has been identified as one of the three most important agricultural pests in South East Asia. In India, it causes losses up to 80% on mango alone. In this study three isolates of *B. bassiana* and three commercial mycoinsecticides were screened against larva, pupa and adults of *B. dorsalis*. The isolates ITCC No. 6628 and B. NCIPM were found pathogenic to adult stage with LC_{50} 2.5×10^5 and 7.5×10^6 conidia /ml, respectively. While ITCC No. 6645 found effective against adult stage at 1.2×109 and 3rd larvae stage with LC_{50} 9×10^9 conidia /ml. However, the three commercial mycoinsecticides prove their efficacy only on adult stages with 26.6%, 40% and 46.6% mortality for, Biopower®, Bio-magic®, Bio-catch®, respectively. Significant differences between treatments and control were recorded accompanied by observation of mycosis on the cadavers.

Testing formulation improvements for application of the entomopathogenic fungus *Metarhizium brunneum* to control Asian longhorned beetles

Sana Gardescu¹, **Ann E. Hajek¹**, Tarryn A. Goble¹, Mark A. Jackson²

¹Dept. Entomology, Cornell University, Ithaca, New York, US, ²USDA, ARS, NCAUR, Crop Bioprotection Research Unit, 1815 N University Street, Peoria, Illinois, US Corresponding author: aeh4@cornell.edu

The Asian longhorned beetle (ALB) infestation in the US midwestern state of Ohio, first detected in 2011, is still under eradication by the USDA. In an ongoing effort by our lab group to assess the potential for biological control using insect-pathogenic fungi, in 2016 we tested a sprayable formulation of Metarhizium brunneum F52 microsclerotia within the Ohio ALB regulated zone, at East Fork State Park. Since ALB might be rare or absent, effectiveness was tested using adult beetles in our quarantine lab colony. Small wood veneer samples (attached to tree trunks with Velcro) were sprayed in June and then collected biweekly from June to August for use in bioassays with adult females. Additional samples were used for spore counts, since insect mortality depends on the density of infective spores produced by the microsclerotia. Spore production peaked at 4 weeks after spraying, resulting in a median ALB survival time (ST50) of 9.5 days after beetle exposure. The Ohio formulation included straw hydromulch and hydrogel to increase moisture retention. To test whether the gel provided a significant improvement over hydromulch alone, in a separate set of 4-week field trials in Ithaca, NY, three versions of the microsclerotial formulation were

compared (straw, gel, or straw+gel). There were no significant differences in beetle mortality from quarantine bioassays for samples collected for July, August and September replicates. In the August trial, a mere 2-day exposure of ALB was effective in killing all beetles within 10-11 days. The area experienced a severe drought in June and samples in the field during that period were not effective. When rainfall was adequate, any of the three formulations showed potential for development as a spray application onto tree trunks for use during summer when adult beetles would be active.

Coating of maize seeds with entomopathogenic STU MCn24 •282• fungi promotes rhizosphere colonization, endophytic ability and provides biocontrol of a pest and pathogen

F. Rivas ^{1,2}, T.A. Jackson³, N. Altier², M. Rostás¹, J. Hampton¹, T.R. Glare¹

¹BioProtection Research Centre (BPRC - Lincoln University, New Zealand),

²National Institute for Agricultural Research (INIA, Uruguay),

³AgResearch (AgR, New Zealand).

Corresponding author: Federico.RivasFranco@lincolnuni.ac.nz

Entomopathogenic fungi were originally identified as pathogens of insects, but recently some species have also been described as plant root colonizers and potentially endophytes. These fungal – plant associations could lead to benefits to the plant such as plant growth promotion or increase systemic resistance to pathogens. We evaluated the impact of different New Zealand isolates of Metarhizium anisopliae, M. robertsii, M. guizhouense, M. novozelandicum, Beauveria bassiana and Trichoderma harzianum, on maize after applying them in a seed coating. The seeds were sown in potting mix in the presence of Costelytra zealandica (root feeding grass grub larvae) and Fusarium graminearum (maize pathogen). Rhizosphere colonization, measured by quantification of fungal colony forming units, was greatest (P=0.019) from roots treated with M. robertsii (F447), followed by M. anisopliae (F672) with M. guizhouense (BK41) the lowest. The presence of grass grub larvae significantly reduced rhizosphere colonisation for all isolates while F. graminearum had no effect. No differences were noted in endophytic ability among the isolates, however when larvae were present the endophytism rate was reduced while F. graminearum increased the endophytic colonisation by the entomopathogenic fungi (P<0.05). Seedling emergence and shoot length were unaffected by the biocontrol fungal strains alone, but M. anisopliae (A1080) and M. guizhouense (BK41) treatments produced increased shoot length in the presence of F. graminearum. Grass grub larvae reduced root dry weight in all maize plants, although this reduction differed depending on the fungal strain used for the seed coating treatment. Bare seeds, without a biocontrol fungal coating, were the most affected by F. graminearum, while B. bassiana (Bb21), M. guizhouense (F16) and M. novozelandicum (F99) reduced the damaged caused to the roots by this fungal pathogen. Biocontrol fungi differed in pathogenicity towards grass grub larvae; with M. anisopliae (F672) being the most pathogenic while M. guizhouense (BK41) and M. robertsii (F447) the least. This study showed that it is possible to deliver entomopathogenic fungi via a seed coating to provide biocontrol of both a fungal pathogen and an insect pest.

Evaluation of pyrethrum, extract of Saccharopolyspora Mcn2 spinosa, Beauveria bassiana and Metarhizium anisopliae for the control of Chaetanaphothrips signipennis, a pest of banana

Mcn25 •283•

Alex Delgado¹, Richard Hall^{1,2}, Daniel Navia¹, William Viera¹, Francisco Báez¹, Mirian Arias¹, and **Trevor A. Jackson³**

¹Instituto Nacional de Investigaciones Agropecuarias (INIAP), Ecuador; ²Proyecto Prometeo (SENESCYT), Ecuador; ³ AgResearch, New Zealand Corresponding author: william.viera@iniap.gob.ec

The banana rust thrip, *Chaetanaphothrips signipennis*, is a significant pest of bananas in South America where it causes mancha roja (red ring disease) which leads to fruit discoloration and fruit rejection from the market. The problem is particularly significant for organic growers where chemical pesticides are prohibited. In this study we evaluated biological products and entomopatho-genic fungi isolated from Ecuadorian soils for control of thrips and prevention of fruit damage. A laboratory bioassay included the treatments of *Beauveria bassiana* (10⁸ conidia per ml) isolated from soil of

organic banana orchard located in El Oro, Metarhizium anisopliae (108 conidia per ml) isolated from a pasture field located in Sucumbios, Phyriplus (Pyrethum) 2 ml/L, Spinosad (Saccharopolyspora spinosa) 1 ml/L, Phyriplus + Spinosad 2 ml/L+ 1 ml/L and a control. In the bioassay, Pyrethrum and Spinosad produced 100% thrip mortality within 24 h and fruit was completely free of red ring discolouration following treatment. The fungi B. bassiana and M. anisopliae produced 23% mortality, significantly greater than the control (7%), but there was no damage reduction with 63 % red ring recorded in the fungal treatments and 25% in the control. However, it is possible that using the high rate of 10⁸ spores per ml killed the insects mechanically rather than through infective processes. Further assays were undertaken with seven strains of B. bassiana and ten strains of M. anisopliae using a maximum of 10⁷ conidia per ml, a concentration likely to be maximal in the field and which would, if pathogenic, kill insects by infection. None produced significant mortality. Pyrethrum and Spinosad have also been tested in the field and adopted for commercial production with applications at bunch formation and at closure of the covering bag. The rejection rate of fruit due to mancha roja has dropped from 90 % to 8 %. It appears that the biological products, Spinosad and Phyriplus, are effective controls for mancha roja for organic farmers and, at this time, the entomopathogenic fungi cannot be recommended for use by producers.

Evaluation of biocontrol potential, plant growth promoting activities and physiological effects of bacterial endophytes against major pathogens of rice in Eastern Himalaya Region of India.

Poster - Mcn26

Mohammed M. Abdelbaset Hassan¹, Dipali Majumder², Dwipendra Thakuria², Krishnappa Rangappa³

¹Microbial Culture Lab School of Crop Protection, College of Post Graduate Studies, Umiam, Meghalaya, India; ²Microbial Ecology Lab, School of Natural Resource Management, College of Post Graduate Studies, Umiam, Meghalaya, India; ³Division of Crop Production, ICAR Research Complex for NEH Region, Umiam, Meghalava, India Corresponding author: abdosayad97@gmail.com

Twenty bacterial endophytes obtained from. Out of 20 isolates tested by Gram staining, 4 were found as Gram+ve and remaining 16 as Gram-ve. Endophytes were tested for PGP attributes and, lytic enzymes and phosphate solubilization. 20 isolates were positive for IAA production (113.16 to 566.05 µg/ml), whereas none of the isolates were positive for phosphate solubilization and HCN production. One isolate (34WE) recorded positive for cellulase production. Three isolates (6WE, 11WE and 44E) were recorded for 100% germination. Rice seeds treated with isolate 34WE showed maximum shoot length (9.68cm) followed by 11WE (9.48 cm), which were statistically non-significant, whereas isolate 60E was recorded as best for root elongation (14.73 cm). Only one isolate i.e. 34WE was recorded as effective antagonist against P. grisea and X.oyzae pv. oryzae with inhibition of 20mm and 11mm respectively in dual culture assay. In-vivo study of endophytes for PGP attributes revealed that, rice seedlings treated with the isolates 8WE showed highest plant height (95.75 cm), followed by isolate 54E (93.75 cm) and 15WE (91.0 cm), which were found significant. All the isolates showed significant root length in comparison to control during in-vivo tests. Number of effective tillers/hill was recorded highest in case of isolate 60E (8.00) followed by isolate 19WE (7.00), which differed significantly. Number of filled grain/panicle was recorded highest in isolate 54E (335), which was statistically significant as compared to control (158.05). Rice seedlings treated with all bacterial endophytes showed significant seeds test weight in comparison to control during in-vivo evaluation. Out of 20 isolates inoculated into rice seedlings (CAUR1), only one isolate i.e. 26WE was observed as effective against the sheath blight pathogen.

WEDNESDAY, 8:00 – 10:00 pm

Fungi Division - Business meeting, Village West, Building 2. Rooms 2A/2B

Microsporidia Division - Business meeting / Workshop, Village West, Building 2, Room 2C

The Microsporidia as Eukaryotes

Microsporidia workshop •284•

Charles Vossbrinck

Department of Environmental Sciences, The Connecticut Agricultural Experiment Station, New Haven, CT, USA Corresponding author: Charles.Vossbrinck@ct.gov

We discuss, in plain terms, the relationship between phylogeny and taxonomy in the context of the Microsporidia as eukaryotes. The goal is not to make a statement about the position of the Microsporidia within the eukaryotes but to look at the data and to discuss the parameters we might weigh as new evidence and new analyses are presented. Factors to consider include: convergence, horizontal gene transfer, phylogenetic methods, long branch attraction, and apomorphic versus plesiomorphic character states. Controversial examples are taken from the vertebrates when considering the appropriateness of accepting paraphyletic taxa.

Virus Division - Business meeting

and tribute to Michael R Strand's election to the US National Academy of Sciences. Organiser: Elisabeth Herniou. Vote of Thanks by Peter Krell. Response by Michael Strand

Village Tower West, 15th fl., 15B

THURSDAY, 8:00 - 10:00 am

Theater

BACTERIA DIVISION SYMPOSIUM

What is Bt? Current perspectives

Organizer and Moderator: Neil Crickmore

Challenges in understanding the biology of Bt

8:00 •285•



Neil Crickmore

University of Sussex, Brighton, UK Corresponding author: n.crickmore@sussex.ac.uk

Despite the fact that Bacillus thuringiensis has been used successfully as a biopesticide for almost 80 years, and its toxin-encoding genes expressed in genetically modified crops for over 20 years, there is still much that we don't understand about this bacterium. Many of us have read, indeed written, papers that tell the world that Bt is a gram-positive, spore-forming, soildwelling, environmentally-friendly organism that produces toxins capable of forming pores in the gut of a target organism and thus killing it. Some of these we are sure about (it is gram-positive) but many of the other aspects remain controversial, or poorly understood. In this symposium we will look at various aspects of Bt and consider some of the broader topics related to its biology. Is Bt just B. cereus with a Cry toxin encoding plasmid? Why are the toxins on plasmids anyway? Where did the toxins come from, spore coat proteins or somewhere else? How are the toxins evolving? Can Bt target human cells? Can Bt even kill insects by itself? This first talk will introduce a number of these topics and will consider some of the difficulties encountered in defining many of the terms we commonly associate with Bt and its toxins.

Bacillus thuringiensis: A constellation of toxins

8:25 •286•

Daniel R. Zeialei

The Ohio State University, Columbus, Ohio, USA Corresponding author: zeigler.1@osu.edu

Bacillus thuringiensis has been traditionally defined as a B. cereus-like organism that produces one or more parasporal crystals composed of proteins toxic to insects and other invertebrates. Nearly four decades of molecular analysis have unveiled a constellation of proteins with widely diverse histories that have become mingled in these organisms and are now co-evolving with them. Many proteins may co-exist in crystals, while others may be synthesized and secreted by vegetatively growing cells. Detailed analysis of the best-studied of these protein families—the three-domain Cry proteins—has revealed that they themselves are composites pieced together from domains with independent histories of their own. There is abundant evidence that homologous recombination between cry genes has resulted in Cry proteins with shuffled domains and therefore a wide array of finely tuned differences in structure and function. Genomic studies confirm that cry genes and other toxin genes are associated with mobile elements, especially megaplasmids. Plasmids are mobile elements. They can be cured, resulting in a crystal-free bacterium, or they can be mobilized in conjugation, resulting in a bacterium with a new combination of toxin genes. There are now numerous examples of cry genes in distantly related endospore-forming species from outside the family Bacillaceae. These observations challenge the traditional definition of B. thuringiensis and raise difficult questions about the phylogeny of the B. cereus species group.

A genomic perspective on the definition, evolution and phylogenetics of Bacillus thuringiensis

Ben Raymond¹, Guillaume Meric², Jinshui Zheng³, Donghai Peng³, Ben Pascoe², Ming Sun³, Samuel K. Sheppard²

¹Department of Bioscience, University of Exeter, Penryn campus, Cornwall, UK; School of Life Science, University of Sussex, Brighton, UK; ²Department of Biology, University of Bath, Bath; 3State Key Laboratory of Agricultural Microbiology, Huazhong Agricultural University, Wuhan, China Corresponding author: b.raymond@exeter.ac.uk

Understanding how and why Bacillus thuringiensis is distinct from Bacillus cereus, an opportunistic pathogen of humans, is crucial when discussing whether Bt isolates make safe pesticides, and for studying the evolution of pathogenicity in the B. cereus group. Currently, using Cry toxins to define Bt imposes compli-cations because cry genes are typically carried on conjugative plasmids. Thus, diverse phylogenetic clades in the B. cereus group contain cry genes, while chromosomally similar strains can be either Bt or B. cereus: making Bt both polyphyletic and paraphyletic. Here I describe two population genomic studies that investigated the distribution of plasmids and virulence genes in the B. cereus group phylogeny. We found that strains carrying cry genes can be found in all the major B. cereus clades. Surprisingly, parasporin genes and cry genes originally associated with Lysinibacillus and Paenibacillus spp. are common and widely distributed. However, the clade containing the classic biopesticidal isolates (clade 2 or the 'kurstaki' clade), stands out for a number of reasons: Bt isolates in clade 2 carry more Cry toxins; toxins with clear invertebrate toxicity are concentrated in clade 2; and Cry toxins in clade 2 are also associated with a cladespecific plasmid. The tight association between virulence plasmids and bacterial chromosomes appears to have evolved several times in the specialized pathogens in the B. cereus group. Strong directional selection in a pathogenic niche could also account for the reduced allelic diversity in cry positive isolates. The variation in carriage of virulence genes also confirms earlier work highlighting the consider-able ecological differences between the major clades in the B. cereus group. We could better incorporate phylogenetics into the definition of Bt by restricting this species name to Cry expressing isolates in clade 2. This definition would exclude several Bt isolates of dubious safety, i.e., those closely related to Bacillus anthracis. An additional practical consequence would be that we would be able to use genes and plasmids unique to clade 2 to develop simple diagnostic tests for Bt.

General discussion

Microbial Control Division #4

Moderators: Roma Gwynn, Dietrich Stephan

Characterization of the efficacy and yield promoting effects of the nematopathogenic fungus Purpureocillium lilacinum: Greenhouse evaluation of BioAct DC (Purpureocillium lilacinum strain 251): understanding and predicting in-field efficacy and beneficial effects on yield

8:00 •288•

The Forum

Tracy K. Webb¹, Kristi R. Sanchez¹, Chun-Jou Hsiang¹, Kristen J. Rasmussen¹, Marc Rist², Malte Roemer², Punita Juneja¹, Matthew R. Tarver¹, Reed N. Royalty¹

¹Bayer, U.S., Crop Science, Biologics, West Sacramento, California, USA; ²Bayer AG, Research & Development, Crop Science, Monheim, Germany Corresponding author: tracv.webb@baver.com

BioAct DC, a novel liquid formulation of the fungus Purpureocillium lilacinum strain 251, has previously shown interesting efficacy in field tests. As is often the case with nematicides, the efficacy of BioAct DC was variable in these trials. However, a detailed cross-trial analysis of the trial results did show a consistent beneficial effect on crop yield. The subsequent development of in vitro plate and in planta bioassays (Sanchez et al.) was pivotal in understanding the fungal biology, plant-parasitic nematode infection process, and the effect of abiotic factors such as temperature on performance of Purpureocillium lilacinum. These lab assays also showed positive effects on growth of plants. In order to more fully understand the potential of BioAct DC as a useful nematode management tool for growers, it was necessary to transfer these lab results to the greenhouse. Bayer nematologists, greenhouse horticulturists, and biometricians worked together to develop an experimental design ideal for greenhouse evaluation of BioAct DC and other nematicide treatments. A latin-square design coupled with detailed measurements of environmental conditions allowed us to incorporate spatial effects into the experimental design. As a result, we created a bioassay method that has, with manageable replication, the statistical power necessary to differentiate among treatments. Our assays showed that data on dry root weight, total fruit weight, and egg counts reliably describe the activity of P. lilacinum and other bionematicides against Meloidogyne javanica. We demonstrated a clear dose-response with P. lilacinum. BioAct DC treatments outperformed competitive biological products, producing significantly higher yields and root weight in treated plants, and a decrease in root knot nematode reproduction. We also tested the potential of an integrated nematode management program featuring BioAct DC in combination with the chemical nematicide fluopyram (Velum Prime). By optimizing the experimental design of the greenhouse pot assays, we can now reliably predict and better understand the results of past and future field trials with P. lilacinum and other bionematicides.

Development of a laboratory testing cascade to predict the in-field activity of the nematopathogenic fungus Purpureocillium lilacinum

8:15 •289•

Kristi R. Sanchez¹, Marc Rist², Matthew R. Tarver¹, Reed N. Royalty¹

¹Bayer, U.S., Crop Science, Biologics, West Sacramento, California, USA; 2Bayer AG, Research & Development, Crop Science, Monheim, Germany Corresponding author: reed.royalty@bayer.com

Nematopathogenicity of the fungus Purpureocillium lilacinum strain 251 has been well documented in the literature. Several commercial products have been introduced, but like many fungal-based biologic products, they have enjoyed limited commercial success due to inconsistent performance. In 2013, Bayer developed BioAct DC, a novel liquid formulation of P. lilacinum. The product showed promising efficacy in field trials, but activity was inconsistent. Our efforts to explain variable field performance with traditional laboratory and greenhouse nematicide bioassays were unsuccessful. To support Bayer's efforts to launch BioAct DC, we developed a cascade of progressively higher-tiered bioassays that characterize the

9:45

performance of nemapathogenic fungi. Our in vitro assay describes the pathogenicity to nematode eggs and motile J2 larvae. With this assay, we made detailed observations of spore attachment, germination, production of appresoria, and subsequent growth of hyphae on infected nematodes. P. lilacinum consistently infected and killed Meloidogyne, Pratylenchus, and Heterodera species. P. lilacinus attacked J2 larvae and eggs of phytophagous nematodes, while the free-living nematode C. elegans was not attacked. These results suggest properties associated with obligate parasitism. We also quantified the effects of temperature on parasitism with this assay. We also developed a small 'jar' test to characterize the activity of nematopathogens against Meloidogyne spp. in planta. This assay was easy to set up, and of short-duration (ca. 2-3 weeks). We found that counting egg masses was a faster and more statistically-robust measure of efficacy than the 'traditional' method of counting galls. Consistent suppression of egg mass production and positive effects on the growth of the host tomato were observed in response to P. lilacinum. BioAct DC provided superior control and plant growth promoting effects relative to other P. lilacinum formulations. Differences in efficacy between strain 251 and other P. lilacinum strains were observed. The bioassay also described a clear dose-response for treatments. Methods developed in this in planta 'jar' test were subsequently incorporated into greenhouse pot trials, where BioAct DC is also showing performance superior to other nematopathogens. Data from our lab tests are being used to both explain results from past field trials, and to design protocols for future testing.

Effect of entomopathogenic fungus, Metarhizium anisopliae on South American tomato leafminer, Tuta absoluta (Meyrick), in the field in Tanzania

8:30 •290•

Never Mwambela², Thomas Dubois¹, Srinivasan Ramasamy¹

¹ World Vegetable Center, P.O. Box 10, Duluti, Arusha Tanzania ²Nelson Mandela African Institution of Science and Technology, P. O. Box 447 Arusha Corresponding author: never.mwambela@worldveg.org

Invasion of the tomato leafminer Tuta absoluta has become the major threat to tomato production in sub-Saharan Africa including Tanzania, Broad spectrum chemical pesticides are the primary mode of control locally, but mis- and overuse of these pesticides lead to development of resistance, environmental degradation and increasing production cost among smallholder farmers in Tanzania. The efficacy of alternative options, such as the use of biological control agents, has not yet been studied in sub-Saharan Africa. This study evaluated the efficacy of the commercially available biopesticide, Metarhizium anisopliae and pheromone lures for management of T. absoluta under field conditions in Tanzania during the rainy and dry seasons. M. anisopliae significantly increased growth and yield of tomato compared to untreated fields to a level that was on par with fields treated with pesticides, whereas pheromone traps were comparatively less effective in preventing damage. In fields treated with M. anisopliae, T. absoluta larval damage decreased by 79.3-85.2% across seasons whereas in fields treated with pheromone traps, damage decreased by 56.0-76.2% across seasons compared to untreated fields. This study demonstrates the biological control potential of M. anisopliae against T. absoluta on tomato in the highlands of Tanzania, and this biological control agent may be suitable for other regions of Africa, as well.

Successful mass production and application of *Metarhizium* 8:45 •291• anisopliae for rice pest control during the full growing season

Guoxiong Peng, Deyu Zeng, Mingsheng Hong, Yuxian Xia

Genetic Engineering Research Center, School of Life Science,
Chongqing University, Chongqing, 400030
corresponding author: yuxianxia@cqu.edu.cn

Pest outbreaks cause significant losses in rice production worldwide. To control various rice pests including the rice stem borer, the planthopper, the leaf roller and others, several chemical insecticides have to be used repeatedly during the growing season. Current dependence of chemical insecticides to control rice insect pests causes increasing public concerns on human health and the environment. We show that a broad spectrum strain of the entomopathogenic fungus *Metarhizium anisopliae*, CQMa421, can infect all the major rice insect pests. We report on the results of a 5 year field

trial study in different provinces in China that shows that an oil suspension of CQMa421 is effective in controlling the three major rice insect pests found, i.e., the stem borer, planthopper, and leaf roller. Although the efficacy of the CQMa420 oil suspension in terms of insect mortality to the three main insect pests were lower in the field trials as compared to use of chemical insecticides (Buprofezin, Chlorpyrifos, Thiamethoxam), overall rice yield in fungal versus chemical treated plots were not significantly different. The results were similar when the oil suspension of CQMa421 was applied repeatedly to control insect pests during the full growing season. Our results demonstrated that the broad-spectrum M. anisopliae strain CQMa421 can protect rice from major insect pests during the full growing season and that simple measurements of pest mortality may be a poor indicator of overall plant health and/or crop yield in agricultural applications. To produce enough of the fungal aerial conidia (and final formulated product) required for large-scale, economically feasible use, a solid-state fermenter was specially designed capable of producing 5 tons of formulated fungal spores within a month period. The successful mass production and application of M. anisopliae for rice pest control resolves a key technological hurdle in terms of production, application, and cost-effectiveness of mycoinsecticides for crop pest control.

Characterization of entomopathogenic fungi for microbial control of *Spodoptera exigua*

0 •292

Ji hee Han, Jeong Jun Kim, Sang Yeop Lee

Agricultural Microbiology Division, National Institute of Agricultural Sciences, Korea Corresponding author:bijouhee@korea.kr

Beet armyworm, *Spodoptera exigua* is one of the most destructive polyphagous pests for many economically important crops but this pest is difficult to control using chemical insecticides because of the development of insecticide resistance. Various control agents may use for eco-friendly beet armyworm managements, Entomopathogenic fungus can be used alternative to chemical control agent. In this study, we investigated characteristic of *Paecilomyces fumosoroseus* which is high virulent isolate against *S. exigua*. *P. fumosoroseus* caused 100% mortality 6 days after treatment at 1×10^4 conidia/ml against 2^{nd} instar larvae and effectively controlled the larvae at 20° 30°C and 45° 100% RH. Control efficacy of *P. fumosoroseus* against 4th-5th instar larvae was $96.7 \pm 2.1\%$ and $36.7 \pm 8.4\%$ and LT_{50} was 4.4 and 6.7 days. This isolate was also found to repel *S. exigua*. Adult of the beet armyworm avoided oviposition at Chinese cabbage treated with *P. fumosoroseus* compare to control. This result may be used to control and prevent the infestation of moth in crop production.

Comparison of different formulations of Beauveria brongniartii for control of white grubs of Melolontha melolontha in apple orchards

Maximilian Paluch, Joschua Göttmann, Dietrich Stephan

Julius Kühn-Institut, Federal Research Centre for Cultivated Plants Institute for Biological Control, Darmstadt, Germany Corresponding author: dietrich.stephan@julius-kuehn.de

White grubs of the common cockchafer, Melolontha melolontha can cause dramatic damage in apple orchards. Because no pesticides are registered to control white grubs in Germany new control methods are needed. Although Beauveria brongniartii is known to control white grubs, up to now no product is registered. This may be due to the high production and registration costs and the complicated application technique. To reduce the production and formulation costs we developed a liquid fermentation system for the B. brongniartii strain BIPESCO2 and coated submerged spores in a fluid bed dryer on millet as carrier. After 68 hours of fermentation time we got the maximum yield of 6.3x10⁸ submerged spores per ml and a dry weight of 4.9 mg per ml. 15 ml of a fungal suspension containing 2 % of biomass (dry weight) was used for coating 100 g millet. For quality control the millet was incubated on water agar and the fungal outgrowth was evaluated. When the coated millet was stored for at least 42 days at 20 °C, the fungus was growing on 100 % of the coated millet grain. No contamination was visible. Up to 1x10⁶ conidia per millet grain were produced on water-agar after 11 days. In a first bioassay all white grubs treated with the coated millet died within nine

weeks. In a field trial at an organic apple orchard three different formulations were compared. Before application soil samples were taken and based on the Galleria and Tenebrio bait method and using selective media, the occurrence of entomopathogenic fungi was determined. The commercial product Melocont and the coated millet were applied with an application rate of 50 kg per ha by slitting the turf and adding the granule underneath of the turf. The application is based on conventional direct sowing machines. Additionally, for one treatment a liquid suspension (7,5 I/ha with a concentration of around 1x10⁵ submerged spores/ml was poured in the slits. Six weeks after application again the occurrence of entomopathogenic fungi was evaluated. First results of this ongoing field trial will be presented and discussed.

Combined effect of Beauveria bassiana and Heterorhabditis bacteriophora against Rhynchophorus ferrugineus (Olivier)

Waqas Wakil¹, Muhammad Yasin^{1,2}

¹Department of Entomology, University of Agriculture, Faisalabad, Pakistan; 1,2 Department of Entomology, College of Agriculture, Bahauddin Zakariya University, Bahadur Sub-Campus, Layyah, Pakistan Corresponding author: waqaswakeel@hotmail.com

The interaction between entomopathogenic nematode Heterorhabditis bacterio-phora and entomopathogenic fungi Beauveria bassiana against fourth-instar of red palm weevil (Rhynchophorus ferrugineus) was studied in the laboratory experiment. Heterorhabditis bacteriophora was added simultaneously or 1, 2 and 3 weeks after application of B. bassiana. The results showed that integrated application of B. bassiana with H. bacteriophora increased larval mortality either in an additive or synergistic manners. To achieve stronger additive and synergistic interactions, larvae had to be exposed at least 2 or 3 weeks before the nematode application. In combined treatments increase in mortality, decrease in pupation, adult emergence and egg hatching was found with a direct relation to the application interval. Our study suggested that the combined use of B. bassiana with H. bacteriophora may offer an effective integrated approach for R. ferrugineus control.

Infection of invasive red palm weevil by endophytic Beauveria bassiana colonizing date palm in combination with Bacillus thuringiensis



Muhammad Yasin^{1,2}, Wagas Wakil²

¹Department of Entomology, College of Agriculture, Bahauddin Zakariya University, Bahadur Sub-Campus, Lavyah, Pakistan: ²Department of Entomology, University of Agriculture, Faisalabad, Pakistan Corresponding author: wagaswakeel@hotmail.com

The effect of Beauveria bassiana inoculated in date palm petioles alone and then with integrated application of Bacillus thuringiensis var. kurstaki (Bt-k) was evaluated against second-instar larvae of Rhynchophorus ferrugineus in laboratory studies. Both the alone and combine treatments were applied against R. ferrugineus larvae to monitor larval mortality, pupation, adult emergence and egg eclosion after treatment. Mortality was low in sole treatments, while in combined treatments increased mortality while decreased pupation, adult emergence and egg eclosion were found. Synergistic effect (CTF≥20) on the mortality was observed when larvae were exposed to simultaneous application of B. bassiana with 40 µg ml⁻¹ of Bt-k. It can be surmised from the findings that endophytic colonization of B. bassiana in combination with Bt-k has potential as an effective strategy to control R. ferrugineus.

Roosevelt Room

DISEASES OF BENEFICIAL INVERTEBRATES SYMPOSIUM

The Pathobiome Concept: An emerging view of microbes and diseases

Organizers and Moderators: David Bass, Helen Hesketh

The pathobiome concept: An emerging view of microbes and disease



David Bass^{1,2}, Ronny van Aerle¹, Lydia Doherty¹ Jamie Bojko¹, Dominique Chaput³, Grant Stentiford¹

¹Centre for Environment, Fisheries and Aquaculture Science (Cefas) Barrack Road, The Nothe, Weymouth, Dorset, DT4 8UB, UK: 2Department of Life Sciences. The Natural History Museum, Cromwell Road, London, SW7 5BD, UK; 3School of Biosciences, Geoffrey Pope Building, University of Exeter, Stocker Road, Exeter, EX4 4QD, UK Corresponding author: david.bass@cefas.co.uk

Recent studies have shown that pathogens, parasites, and related lineages are highly diverse in environmental and organismal matrices. While some of these are recognized disease causing agents, the role of others is less clear they may be asymptomatic or contribute directly or indirectly to a cumulative effect on host health. The pathobiome idea is starting to replace a one-pathogen-one-disease paradigm, based on the concepts of a host organism being the focus of a diverse community of microbial symbionts, multiple infections being the norm rather than exception, and synergistic effects occurring between infecting microbes. This talk will present the results of recent studies investigating microbial diversity associated with a range of invertebrate and plant hosts, and review broader findings supporting the pathobiome concept, a further case study being the microbiome associated with disease outbreaks in aquaculture pond systems.

Characterization of pathobiomes using high-throughput sequencing: Challenges and opportunities

8:30 •297•



Ronny van Aerle

¹European Union Reference Laboratory for Crustacean Diseases, Centre for Environment, Fisheries and Aquaculture Science (Cefas), Barrack Road, The Nothe, Weymouth, Dorset, DT4 8UB, UK Corresponding author: ronny.vanaerle@cefas.co.uk

Recently, it has been shown that many pathogens are part of complex microbial communities, collectively known as the pathobiome, in which they interact with other micro-organisms (e.g., bacteria, viruses, fungi, protozoa) to influence or drive disease processes in the host organism. In order to understand the role of the pathobiome in the disease process, it is important to study the diversity of organisms in this community. Recent advances in high-throughput sequencing technologies and the development of comprehensive bioinformatic tools facilitated the analysis of species diversity within pathobiomes based on sequence information. This presentation will provide an overview of the current approaches used in the characterization of pathobiomes, including metagenetics (e.g., 16S/18S rRNA amplicon sequencing) and metagenomics (shotgun DNA sequencing of environmental samples). The importance of study design (e.g., inclusion of relevant controls, sequencing coverage/depth) as well as challenges (e.g., limited sequence availability in public reference databases, false positives) and opportunities (e.g., single molecule sequencing, functional metagenomics) will be

Effects of the gut microbiota on susceptibility to pathogens in social bees

9.00



Nancy A. Moran

¹ University of Texas, 2506 Speedway, NMS 4.216, Stop A5000, Austin, Texas, 78712 Corresponding author nancy.moran@austin.utexas.edu:

Honey bees and bumble bees have a characteristic gut microbiota that has coevolved with them for millions of years. In honey bees, the adult gut is dominated by 9 bacterial species, each of which can be present as multiple strains with distinct capabilities. Early evidence for a protective effect of the gut microbiota came from bumble bee studies showing that the microbiota protected against trypanosomatid parasites. Later studies have shown that the gut microbiota of honey bees has a protective effect against opportunistic Gram negative bacterial pathogens. The routes of protection are not well understood, but colonization by the entire gut microbiota or by single strains of the core species results in upregulation of antimicrobial peptides of the host. There may also be direct interactions within the gut that enable the core microbiota species to kill opportunistic pathogens or to block their entry from the gut into bee cells or the body cavity. Additionally, the gut microbiota in bees causes a major shift in the physicochemical conditions within the gut, likely making the environment more hostile to invasive opportunists. Use of antibiotics in bee colonies, for prophylaxis against larval pathogens, may result in higher adult mortality due to disruption of the adult gut microbiota. Both honey bees and bumble bees are beneficial insects that appear to be undergoing increasing rates of mortality. Preservation of honey bee and bumble bee health will likely be enhanced by practices that help to sustain a healthy gut microbiota.

Competitive interactions and disease suppression in soil microbiomes

9:30 •299•

JP Dundore-Arias, Linda L. Kinkel

Department of Plant Pathology, University of Minnesota, Saint Paul, Minnesota $Corresponding\ author: jdundore@umn.edu$

Plants live in intimate association with diverse microbes, and these associations are essential for plant performance and survival. Our work explores the interacting roles of plants and microbial species interactions in determining the pathogen-suppressive capacity and composition of soil microbiomes, and the consequences for plant productivity. Using culturebased approaches we have found that plant community diversity plays a critical role in determining the likelihood of antagonistic arms race coevolution vs. niche differentiation among sympatric soil populations, with significant implications for plant disease suppression. Moreover, amplicon sequencing of rhizosphere communities associated with different plant hosts provide insights into non-cultured taxa associated with disease suppression. This presentation will cover what we have learned about the factors that contribute to determining the pathogen-suppressive and plant growthpromoting potential of indigenous soil microbes, and the possibility to create tailored approaches targeting species interactions to enhance plant health and productivity.

10:00-10:30 am

Coffee Break

THURSDAY, 10:30 am - 12:30 pm

Theater

Society for Invertebrate Pathology **Annual Business Meeting**

J. Jehle, President, SIP

THURSDAY, 12:30 - 1:30 pm

IMPORTANT: All posters must be removed before noon today.

Awards Committee member lunch (Green Room) Monique van Oers, Chair

THURSDAY, 1:30 - 3:30 pm

Theater

FUNGI, NEMATODES, AND MICROSPORIDIA DIVISIONS **CROSS-DIVISIONAL SYMPOSIUM**

Ecology of Invertebrate Pathogens

Organizers: David Shapiro-Ilan, Ann E. Hajek

Interaction networks of invertebrate pathogenic fungi: **Ecological consequences of multifunctional lifestyles** and community diversity

1:30 •300•

Nicolai V. Meyling¹, Ann E. Hajek²

¹University of Copenhagen, Dept. of Plant and Environmental Sciences. Frederiksberg C, Denmark; ²Cornell University, Dept. of Entomology, Ithaca, NY, USA Corresponding author: nvm@plen.ku.dk

Two main taxonomic groups include invertebrate pathogenic fungi, Entomophthorales and Hypocreales. These fungi interact with many hosts, are widespread in terrestrial ecosystems and are causing significant ecological effects. Recent studies have revealed an extraordinary diversity within hypocrealean invertebrate pathogenic fungi regarding both their genomic variability and ability to exploit hosts from different kingdoms as resources. The traditional view of fungal species as a single entity is therefore challenged as is the perception of the entomopathogenic hypocrealean fungi as solely acting as pathogens in nature. We give examples of recent knowledge of resource utilization and community diversity within the main genera Metarhizium and Beauveria to emphasize the complex interaction networks that these fungi may engage in. We discuss the significance of these interaction networks in an ecological context to predict the potential effects of the interactions as well as the possible causes for the complex community structure of the invertebrate pathogenic fungi observed in various ecosystems.

Dispersal, dispersion and disruption: where and why entomopathogenic nematodes move 1:50 •301•

Edwin Lewis¹, Ivan Hiltpold², David Shapiro-Ilan³

¹Department of Entomology, Plant Pathology and Nematology, University of Idaho, Moscow, ID, USA; ²Department of Entomology and Wildlife Ecology, University of Deleware, Newark, DE, USA; ³USDA-ARS, SEA, SE Fruit and Tree Nut Research Unit, Byron, GA, USA

Corresponding author: eelewis@uidaho.edu

Entomopathogenic nematodes (EPNs) are unusual as pathogens because of how much they move. Infective juveniles move to maximize the probability of encountering hosts. The strategies EPNs employ to find hosts have a great impact on which host species they ultimately infect. Many studies describe and measure EPN infective stages' movement toward hosts, or toward plants that are infested with potential hosts, but what stimulates and governs movement when hosts are absent? EPNs are active without being directly stimulated by host cues and their population structure does not depend entirely on host population structure. Thus, an intriguing question is: what makes an obligate parasite with no function other than to find a host move, other than the presence of a host? Dispersal of EPN IJs as they leave the cadaver is driven by conditions in the cadaver and the presence of semiochemicals, including ascaroside pheromones, at the end of an infection. In addition to stimulating dispersal from the host, these substances also influence infection behavior, tolerance to environmental extremes and lifespan of the IJs. Distribution of IJs in the soil is known to be patchy, but what is the advantage to staying in groups? Group dynamics studies suggest that there is a cost to being alone. For EPNs, that cost is the inability of a single IJ to successfully infect a host (assuming success is defined as producing offspring). Thus, not only do EPN IJ populations have a clumped distribution in the field, but they move en masse in the absence of a host as well. A disturbance can be defined as the interruption of a settled and peaceful condition. Soil conditions affect all aspects of EPN existence, especially dispersal and dispersion. Thus, layered on top of complex behaviors, physiology and infection dynamics are the temporally and

spatially variable conditions within the soil environment. Peace and happiness for EPNs in an unforgiving world remains a challenging goal.

The ecology of microsporidia - a balancing act between minimizing host damage and maximizing pathogen reproduction and transmission



Leellen Solter¹, Gernot Hoch²

¹Illinois Natural History Survey, Prairie Research Institute, University of Illinois, Champaign, IL, USA; ² BFW Austrian Research Centre for Forests, Department of Forest Protection, Vienna, Austria ${\it Corresponding\ author: } IsoIter@illinois.edu$

The Phylum Microsporidia, a monophyletic taxon of eukaryotic unicellular pathogens that appear to be ubiquitous in invertebrate populations, possess a suite of unique genetic and biological characteristics that determine interactions with their hosts. Similar to pathogens in other taxonomic groups, interactions between each microsporidian species and its host(s) can vary considerably from those of other microsporidian-host relationships, but the overall trend is a general chronicity that maximizes pathogen production, vertical transmission or both. Typically, microsporidia do not cause detectable harm to the cells of their hosts until sporulation begins. When sporulation is completed, horizontally transmitted species often cause high mortality in their hosts. Vertically transmitted species tend to be less virulent. Characteristics that best describe microsporidia include an uncertain taxonomic placement related to Fungi; obligate intercellular development; absence of many eukaryotic features, notably functional mitochondria; interactions with host mitochondria in infected cells that suggest direct energy (ATP) uptake; lack of plant and fungal hosts; and possession of a unique invasion apparatus consisting of a polar filament through which the infective form is injected from the spore into a host cell. Microsporidian life cycles range from simple to highly complex and, in some cases, include obligate intermediate hosts. As well, pathogen-host population dynamics and impacts on host populations are highly diverse. Here, we highlight some of the unique characteristics of microsporidia that govern the ecology of these primary pathogens of invertebrate animals.

The hidden ecology of bacterial entomopathogens

2:30 •303•

Trevor A. Jackson¹, Colin Berry² and Maureen O'Callaghan¹

¹AgResearch, New Zealand; ²Cardiff University, UK Corresponding author: trevor.jackson@agresearch.co.nz

This presentation will review the ecology of bacterial entomopathogens which are frequently reported from dead and diseased insects and include well known species, such as Bacillus thuringiensis, and more recently discovered pathogens like Yersinia entomophaga. Unlike other pathogen groups, bacteria are rarely obligate pathogens and often exist in commensal forms with an ecology outside the insect host. In spite of this, entomopathogens often show a high degree of host specificity suggesting an evolutionary advantage from the close host association. The pathogenicity genes of entomopatho-genic bacteria can be chromosomal or borne on transferable elements (plasmids) suggesting a gene flow (transfer) between pathogenic and non-pathogenic forms. During infections, entomopathogenic bacteria use the insect host as an energy source to multiply and once an epizootic has developed can become dominant forms in the microbiota. To take advantage of the relationship with the insect host, bacteria must have a mechanism for transmission between hosts and survival through periods with absence of susceptible hosts. This can be achieved through resistant stages (spores), or survival and multiplication in the host's environment. Vertical transmission through all stages of the insect life cycle is suggested for some species. Despite their abundance, the ecology of most species of bacterial entomopathogens is not well understood and they constitute a hidden resource for further study and exploitation. Bacterial entomopathogens are an important resource for pest management and knowledge of their ecology will aid the search for new forms/genes as well as guide strategies for their use and management after application.

Ecology of insect viruses: how viruses adapt to their insect host population

2:50



Madoka Nakai¹, Trevor Williams ¹

¹Institute of Agriculture, Tokyo University of Agriculture and Technology (TUAT). Fuchu Tokyo, Japan; ² Instituto de Ecologia AC (INECOL), Xalapa, Mexico Corresponding author: madoka@cc.tuat.ac.jp

Insect viruses cause both overt disease and covert infection in their hosts. One key characteristic that determines the ecology of insect viruses is the presence or absence of an occlusion body (OB). The OB is particularly important in the transmission of viruses that infect insects in terrestrial habitats. The best studied insect viruses from an ecological standpoint are baculoviruses because they can be used as biological insecticides against agricultural and forest pests. Parameters affecting the efficacy of baculoviruses as natural biological control agents are well documented involving infectivity, speed of kill, persistence in the environment and as covert infections in the host population and the range of host species that are infected naturally in a particular ecosystem. Other perspectives of baculovirus ecology include the study of cyclic population dynamics of some forest pests including the Gypsy moth (Lymantria dispar) or the Western tent caterpillar (Malacosoma californicum pluviale). In non-occluded insect viruses transmission often ocurrs directly from infected host to susceptible host through mating and cannibalism, or indirectly via an insect vector. Ascoviruses are unusual insect viruses that are transmitted via endoparasitoids. Overall the transmission, persistence and dispersal of insect viruses are modulated through host factors such as insect feeding and dispersal behavior, and by biotic and abiotic factors, however, these should not be viewed in isolation but rather as a set of interacting and interdependent processes.

Ecology of emerging infectious diseases of invertebrates

3:10 •305•

Colleen A. Burge, Natalie D. Rivlin, Amanda Shore-Maggio

Institute of Marine and Environmental Technology, University of Maryland Baltimore County, Baltimore, Maryland, United States Corresponding author: colleenb@umbc.edu

Infectious diseases are important natural drivers within ecosystems. However, epidemics of infectious disease have become more frequent, causing concern about the long-term impacts on ecosystem function, economic growth, and food security. Of particular concern are emerging infectious diseases (EIDs), defined as those which have rapidly increased in incidence or geographic range, recently moved into new host populations, and/or recently been discovered or are associated with a newly evolved pathogen(s). Even though invertebrates represent the greatest diversity of animals in both marine and terrestrial ecosystems, EIDs of invertebrates have been poorly defined and are not well understood, in part from a lack of baseline data, unresolved or complex etiologies, lack of specific diagnostic tools, and inherent difficulty of monitoring for disease outbreaks. The best information on invertebrate disease ecology, human influence on disease emergence, and best practices for disease management/prevention come from investigations of commercially important invertebrates (e.g., oysters, shrimp, and bees). More effort is being made to better understand diseases of ecologically important species (e.g. corals, sea stars). Disease emergence is often defined by the disease triad (host-pathogen-environment relationship) and how direct anthropogenic change and climate change shift the underlying homeostasis. Several case studies will be used to discuss EIDs of invertebrates in terrestrial and marine systems, specifically highlighting the basic ecology and etiology of the disease, the human activities or environmental covariates of disease emergence and/or transmission, and attempted or preventative measures. Given the high impact of invertebrate EIDs, we need to be ready to detect and to respond to the next invertebrate

The Forum

MICROBIAL CONTROL DIVISION SYMPOSIUM

Biopesticides III: Beyond entomopathogenicity -Reliable tools or just a novelty?

Organizers amd Moderators: Roma Gwynn, Travis Glare, Michael Brownbridge

We increasingly see papers showing effects of microbial inoculants on plant performance (improved stress tolerance, better growth, etc.), and evidence of effects of endophytes and epiphytes on plant genes and expression of compounds in response to stress (abiotic and biotic). Entomopathogenic fungi have long been considered as biocontrol agents based on their ability to directly infect and kill arthropods. More recently, non-lethal effects which complement crop protection efforts have been documented and their status as (part-time?) endophytes demonstrated. Effects of 'non-entomopathogens' on insect and other pests are also well recognized for true endophytes such as Epichloë, but not so much for many 'newer endophytic associations (are all entomopathogenic fungi really endophytic?). To take full advantage of the benefits these organisms can bring to crop protection efforts we need to be able to demonstrate the type of plant responses that can be expected from these associations, the level to which these are induced, and the consistency of these responses (on a commercial scale). In addition, cost-effective ways of deploying these technologies must be found, with attendant regulatory pathways defined and end-user buy-in confirmed before they can be fully commercialized. 'Biopesticides III' builds on prior Symposia at SIP, and brings together a mix of industry and academia to consider the end-game for these plant-microbe associations.

Biopesticides III - The final chapter?

1:30 •306•

Michael Brownbridge

Vineland Research and Innovation Centre, 4890 Victoria Ave N, Vineland Sta., ON Canada LOR 2E0 $Corresponding\ author:\ michael. brown bridge @vineland research. com$

Biopesticides are increasingly considered as replacements for chemical pesticides that have come under regulatory scrutiny, as essential components of resistance management, and as components of IPM systems where functional compati-bility with other biological control agents is essential. In the first Biopesticides Symposium, we considered the prevalence of insect-pathogenic fungi in nature, aspects related to improving the consistency of their performance, and how other characteristics shown by these organisms, e.g., as endophytes, may contribute to pest management. In the second chapter, we explored growing and evolving opportunities to use microbial biocontrol agents in plant protection. While these approaches are based on insect pathogens, they use their multiple ecologies, bioactives or new formulation and delivery systems to more fully exploit these beneficial microbes. In the third session of this series, we will focus on nonlethal effects of 'insect pathogens' which complement crop protection and assess their status as endophytes. Effects of 'non-entomopathogens' on insect and other pests are well recognized for true endophytes such as Epichloë, but not so much for entomopathogenic fungi such as Beauveria and Meta-rhizium. To take full advantage of these organisms in pest management we need to be able to demonstrate the type of plant responses that can be expected from these associations, the level to which these are induced, their impact on pest (and non-target) organisms, and the consistency of these responses on a commercial scale. In addition, costeffective ways of deploying these technologies must be found, with attendant regulatory pathways defined and end-user buy-in confirmed before they can be fully commercialized.

Plant associations of Metarhizium and Beauveria

2:00 •307•



Travis R. Glare, Federico Rivas, Aimee C. McKinnon, Maya Raad, Maria E. Moran-Diez, Mchael Rostás

> BioProtection Research Centre, Lincoln University, PO Box 84085, Lincoln, New Zealand Corresponding author: travis.alare@lincoln.ac.nz

Entomopathogenic fungi in the genera Metarhizium and Beauveria have been well studied as direct mortality agents against invertebrate pests. Many products are based on strains from these genera. The discovery of the ability by species of both genera to colonise plants has suggested exciting possibilities for further exploitation in pest control. A plethora of publications have indicated endophytic occurrence and other studies have shown root colonisation abilities. What is the true ecology of these insect pathogens and what is the potential for commercial and practical use of the ability to closely associate with plants? It seems that not all species or strains within the genera share plant association abilities, but there also does not appear to be

specialisation of strains within species. Are some species truly endophytes or are these largely opportunistic associations? The crucial aspect of these plant associations, whether they confer benefits to the plant in terms of pest and disease resistance and plant growth promotion, also appear to be variable. We will explore the evidence that is accumulating on plant associated entomopathogens from Beauveria and Metarhizium, including some of our own data. The response of plants to colonisation provides fascinating insights into resistance mechanisms. Is it now possible to develop crops with stable communities of associated entomopathogens?

Beauveria bassiana strain ATCC 74040 - does the understanding of its endophytic activity affect its use as a biocontrol agent?



Edith Ladurner¹, Massimo Benuzzi¹, Sergio Franceschini², Francesco Greco²

¹CBC (Europe) S.r.l. – BIOGARD Division Technical Area, Cesena, Italy, ²CBC (Europe) S.r.l. – BIOGARD Division, Grassobbio, Italy Corresponding author: eladurner@cbceurope.it

Beauveria bassiana strain ATCC 74040, the active ingredient of the formulated plant protection product Naturalis®, has been shown to be able to endo-phytically colonize some plant species, in particular grapevine and tomato. Furthermore, evidence exists that the microbial control agent, once endo-phytically established, can contribute to negatively affecting the survival and/or development of key insect pests of grapevine and tomato, respectively grape mealybug (Planococcus ficus) and tomato leaf miner (Tuta absoluta). In short, the likelihood exists that the endophytic activity of the fungal strain could be exploited to complement crop protection. The potential of the findings of the above-mentioned studies in affecting the use of B. bassiana strain ATCC 74040 as a biocontrol agent is discussed in consideration of several important aspects, such as application rates tested in the studies on endophytic activity of the antagonist versus authorized application rates of the microbial control agent, observed percentages and consistency of endophytic colonization, efficacy observed in the abovementioned studies versus efficacy considered acceptable for the purposes of registration of a plant protection product and by growers, etc.. An in-depth analysis of the possible effects and repercussions of the observed endophytic activity of B. bassiana strain ATCC 74040 on its use as a plant protection product is provided.

Approaches for the commercialisation of plant colonising microorganism: the importance of educating the gatekeepers



Roma Gwynn

Rationale, 1 Lintlaw Farm Cottages, Duns TD11 3QA, Scotland. UK Corresponding author: rgwynn@biorationale.co.uk

Microbials developed for crop protection for farmers and growers (for both for insects and diseases) have so far been based on well-known species and the interactions have been more or less binary, e.g., entomopathogenic fungi directly colonising and killing the host. Established commercial development pathways are in place for these technologies, including appropriate regulatory frameworks or approaches. Whereas, plant colonising microorganisms are complex, they have a wide range of interactions that are influenced not only by the interaction with the pests and diseases but also by the plant which in turn, is influenced by its biotic and abiotic environment. Capturing the net of all these effects and using them to develop a product that can be sold is largely unknown but probably equally complex. Applied research into plant colonising microorganisms for pest and disease management is moving forwards quickly but will bringing plant colonising microorganism onto the market be simple? Once it is established that the strain can be produced reliably on a large enough scale, commercialisation is a combination of does the product 'work', how to use it to obtain consistent benefits and how can it be regulated. In particular, how can their effectiveness be demonstrated and how can this be explained to end users (agronomists, crop consultants, sales personnel and farmers). It will be important to know what benefits the microbes deliver, whether they are always measureable and whether they can be demonstrated commercially. Sitting alongside this for commercial development companies, is that they need to know what the regulatory pathways are for plant colonising

microorganisms. This presentation will consider the commercialisation processes for plant colonising micro-organisms and how this will influence the ease of availability and adoption of these new technologies.

Plant microbe interactions, microbial solutions for invertebrate pests, and crop yield: Current and future focus of the crop protection industry

3:30 •310•



Reed N. Royalty, Dilara Ally, Magalie Guilhabert

Bayer, U.S., Crop Science, Biologics, West Sacramento, CA, USA

Corresponding author:reed.royalty@bayer.com

Many microbial-based pest and disease control products have been introduced into the agricultural marketplace. Although some of these products have been successful commercially, they are justifiably perceived as being less efficacious than alternative solutions. For this reason, there has traditionally been minimal R&D investment by the crop protection industry in microbial solutions, relative to the level of investment in synthetic chemistry and genetically-modified crops. However, due to the recent commercial success of microbial seed treatments, and to regulatory pressure on and public concerns about chemical pesticides and genetically-modified crops, the agrichemical industry is now investing significantly in microbial solutions. Increased investment is occurring both in major agrichemical companies and in smaller 'start up' firms. Research and development on microbial solutions differs from research on chemical and genetically modified plant solutions in that the research often focuses on the interactions between microbes, plants, the targeted pest, and the environment. This broader 'ecological' approach is viewed by many companies as having a greater likelihood of technical success than simply focusing on controlling a particular pest with a particular microbe. In this presentation we present our perspective on recent research activities by the agrichemical industry on endophytic and rhizosphericcompetent microbes.

Roosevelt Room

Virus Division #5

Moderators: Roma Gwynn, Dietrich Stephan

New viruses exclusive to the honey bee pathogen mite Varroa destructor

1:30 •311•



Nor Chejanovsky¹, Sofia Levin^{1,2}, Noa Sela¹,

¹Institute of Plant Protection, Agricultural Research Organization, Israel; ²Faculty of Agricultural, Food and the Environmental Quality Sciences, The Hebrew University of Jerusalem, Rehovot, Israel Corresponding author: ninar@volcani.agri.gov.il

Varroa destructor, the endoparasitic mite of Apis mellifera carries and/or promotes replication of viruses pathogenic to the honey bee like the Deformed wing virus, the Acute bee paralysis virus, the Israeli acute bee paralysis virus and the Kashmir bee virus. However, viruses exclusively associated with Varroa and not present in A. mellifera were not found. To search for viruses associated with- or infecting V. destructor we performed next generation sequencing (NGS) of RNA extracted from Varroa mites and honey bees in Varroa-infested colonies. Comparative bioinformatic analysis of the two separate contig-assemblies generated from the sequences' reads, annotated using Blastx, enabled identification of new viruses unique to Varroa and absent in A. mellifera: an Iflavirus and a virus with homology to Ixodes scapularis associated virus 2, that we named Varroa destructor virus 2 (VDV-2) and 3(VDV-3), respectively. Furthermore, we confirmed these findings sequencing the mite- and honey bee-viromes and in by Reverse transcription polymerase chain reaction in separate mites and honey bees randomly sampled. The genomes of VDV-2 and VDV-3 bear 9576 nucleotides and 4202 nucleotides, respectively. Phylogenetic analysis of VDV-3 suggests that it belongs to a new group of viruses. Our results open venues for investigating the pathogenicity of these new *V. destructor* viruses.

A microcarrier-based process for production of the Oryctes nudivirus in spinner-flask bioreactors

Charlotte Pushparajan¹, Juan Daniel Claus², Gabriel Alberto Visnovsky³

¹Laboratory for Evolution and Development, Department of Biochemistry, University of Otago, Dunedin, 9016, New Zealand; ²Department of Chemical and Process Engineering, University of Canterbury, New Zealand: ³Lab. Virologia, Facultad de Bioquimica y Ciencias Biologicas, Universidad Nacional del Litoral, Santa Fe, S3000, Argentina Corresponding author: aabriel.visnovskv@canterburv.ac.nz

The coconut rhinoceros beetle, an economically important palm pest, is effectively controlled by application of its natural pathogen, Oryctes nudivirus (OrNV). Production of OrNV can be performed in vivo by infecting larvae. In vivo production has several disadvantages, primarily, lack of sterility and inconsistency in virus titers produced. In vitro production of OrNV in the susceptible and permissive DSIR-HA-1179 host insect cell line, is an attractive alternative to produce pure, sterile virus at a standardized titer. Among other factors, the feasibility of producing viral biological control agents in insect cell cultures in bioreactors rests on the ability to grow and subsequently infect host cells in suspension culture. The anchoragedependent nature of the DSIR-HA-1179 cell line precludes its growth in free suspension, but microcarriers which provide a surface area on which cells can grow attached to, can be used to cultivate them within a suspension-like environment and provide a starting point to scale-up the system. This study evaluated OrNV production in DSIR-HA-1179 microcarrier cultures in spinner flask bioreactors. Cells were grown in TC-100 culture medium supplemented with 10% fetal bovine serum on three types of microcarrier, Cytodex-1, Cytodex-3 and Cultispher-G, in 60ml spinner flasks. Cells attached to Cytodex-1 and Cytodex-3 with efficiencies of 99.6% and 98.5% respectively, but failed to attach to Cultispher-G. Maximum cell density was dependent on bead type and concentration and cell-to-bead ratio. With a bead concentration of 1 mg/ml and cell-to-bead ratio of 30, a maximum cell density of 1.7 x 10⁶ cells/ml was reached on Cytodex-1 and 1.2 x 10⁶ cells/ml on Cytodex-3 microcarriers. Since it supported higher cell yields, Cytodex-1 was chosen to study the kinetics of OrNV production in this system. Infecting cells during the early exponential growth phase at a multiplicity of infection of 1 led to an OrNV yield of 1.3 x 10⁸ TCID₅₀/ml and cell-specific yield of 177 TCID₅₀/cell, which was an improvement over volumetric (6.5 x 10⁷ TCID₅₀/ml) and cell-specific yields (102 TCID₅₀/cell) obtained in infections in attached Tflask cultures. Future work will focus on optimization and scaling-up of OrNV production in the microcarrier system in stirred tank and airlift bioreactors.

Vago-like gene, BtSVC is involved in antiviral immune responses and antimicrobial peptides (AMPs) expression in bumblebee Bombus terrestris

2:00 •313•

Haidong Wang, Guy Smagghe and Ivan Meeus

Department of Crop Protection, Faculty of Bioscience Engineering, Ghent University, Coupure Links 653, 9000 Ghent, Belgium Corresponding author: Ivan. Meeus@ugent.be

The single von Willebrand factor C (SVCs) proteins are a recently discovered arthropods protein family with their functionalities poorly characterized. Recent study has shown that Vago, a member of SVCs can act as an antiviral modulator in *Drosophila melanogaster* and *Culex* mosquitos. Within hymenopteran species, we noticed a reduced repertoire of SVC protein family members compared to flies and mosquitos. Especially in most bee species, there is just one SVC which function is not clear. Therefore we asked if this SVC in bumblebee Bombus terrestris is an immune modulator. More specifically, we tested its antiviral function in relation to Israeli acute paralysis virus (IAPV). Our results showed that silencing BtSVC lead to increased viral titers in fat bodies. Silencing Dicer-2, a key player of the siRNA pathway resulted in a lower expression of BtSVC and increased viral titers. Interestingly, we also found that silencing BtSVC can lower the expression of four antimicrobial peptides (AMP)-coding genes. These results proved that BtSVC has a vago-like function and plays a role in the antiviral immunity of bees. Furthermore, the interaction of BtSVC with AMPs related pathway also shows its potential to be an immune modulator in the insect innate immune system.

Pesticide enhance the level of chronic bee paralysis virus

2:15 •314

Beibei Li^{1,2}, Shuai Deng^{1,2}, Fei Li^{1,2}, Qinqyun Diao^{1,2}, **Chunsheng Hou**^{1,2}

¹Institute of Apicutural Research, Chinese Academy of Agricultural Sciences; ²Key Laboratory of Pollination Insect Biology, Ministry of Agriculture Corresponding author: houchunsheng@caas.cn;dqyun1@126.com

Honey bee plays a vital role in pollination for plants and crops, especially for wild plants in mountain area in China. However, there are increasing reports about the large number of honey bee dead or unable to forage in many apiaries. Although a variety of emergent microbial pathogens were thought to be impact to honeybee colony losses, little is known about the interaction between viruses and environmental stressors, such as pesticides. To understand better the causes, we collected samples from several provinces and to screen the 8 common viruses and imidacloprid. The results shown that samples were widespread infected by CBPV as well as imidacloprid from environment. To confirm whether imidacloprid can elevate the level of CBPV under the natural condition, we infected healthy bee samples with crude CBPV at low level and then feed the imidacloprid, and found that imidacloprid can enhance the level of CBPV of honeybee. These indicated that pesticides from environment have impacts on honey bee healthy in a long term. This work was funded by the National Natural Science Foundation of China (31572471) and the Agricultural Science and Technology Innovation Program (CAAS-ASTIP-2017-IAR).

Shinobi tetravirus and Kunoichi rhabdovirus, latent viruses in mosquito cultured cell line, suppress multiplication of arboviruses

2:30 •315•

Ryosuke Fujita^{1,2}, Fumihiro Kato², Shigeru Tajima², Masayuki Saijo², Haruhiko Isawa², Kyoko Sawabe²

¹Creative Research Institution, Hokkaido University, Japan ²National Institute of Infectious Diseases, Tokyo, Japan Corresponding author: r-fujita@cris.hokudai.ac.jp

Viruses in blood-sucking insects can be categorized in following 3 groups; insecti-cidal viruses, human/animal-pathogenic viruses so called arthropodborne viruses (arboviruses), and other viruses without any symptoms in infected hosts. Recent technical developments such as next generation sequence analysis enable us to identify viruses without morphological changes in infected cells. We could successfully identify and isolate two viruses which latently infect to mosquito Aedes albopictus cultured cell line C6/36 obtained from a cell bank of JCRB. Shinobi tetravirus (STV) was a novel single-strand, positive sense RNA virus, and Kunoichi rhabdovirus (KRV) was a novel virus categolized in a family Rhabdo-viridae. These viruses were not found in the C6/36 cells obtained from a different cell bank. Using these cell lines, we compared growth of Zika virus (ZIKV), Dengue virus (DENV), Japanese encephalitis virus (JEV), Sindbis virus (SINV), and Getah virus (GETV). As a result, the growth of flaviviruses (ZIKV, DENV, and JEV) were dramatically suppressed in JCRB C6/36 cell lines whereas the differences of growth kinetics of alphaviruses (SINV and GETV) were not so significant. We cloned STV and KRV from JCRB C6/36 cell line, then re-infected them to another C6/36 cell line, resulting in reproduction of latent infection of each virus. ZIKV growth was also suppressed in STV and/or KRV re-infected C6/36 cells. We also determined the response of innate immune signaling pathway in viral infected C6/36 cells and found that IMD pathway was up-regulated by STV and/or KRV latent infection, although the contribution of IMD pathway to suppression of ZIKV was still obscure. This is the first demonstration that latently infected rhabdovirus and/or tetravirus suppressed flavivirus replication in mosquito cells.

Origin of supplementary capsid proteins in different AAV serotypes produced with the baculovirus/insect cell expression system

2:45 •316

Lionel Galibert^{1,2}, Adrien Savy^{1,3}, Yohann Dickx¹, Delphine Bonnin¹, Bérangère Bertin¹, Isidore Mushimiyimana⁴, Aurélien Jacob^{1,5}, Marjorie Boutin-Fontaine¹, Monique M. van Oers⁶, Philippe Moullier⁷, Fulvio Mavilio¹, **Otto-W. Merten**¹ ¹Généthon, Evry, France; ²FinVector Vision Therapies, Kuopio, Finland;

³Synpromics Ltd., Edinburgh, U.K.; ⁴University of Eastern Finland,
A.I. Virtanen Institute for Molecular Sciences, Kuopio, Finland; ⁵TIGET,
Ospedale San Raffaele – Milano, Milan, Italy; ⁶Wageningen University,
Laboratory of Virology, Wageningen, The Netherlands; ⁷INSERM UMR 1089,
Thérapie génique translationnelle des maladies neuromusculaires

et de la rétine, Nantes, France

Corresponding author: omerten@genethon.fr

The baculovirus/insect cell expression system is a common scalable production system for adeno-associated viral (AAV) vectors which are of great interest for in vivo gene therapy of rare genetic disorders. The generation and analysis of several AAV serotypes produced with this system has shown that, in addition to the three structural proteins (vp1, vp2, vp3) supplementary viral peptides could often be revealed by Western analysis. Moreover, for several serotypes, the expression level of vp1 was well below the expected level (which normally should be equivalent to that of vp2). More detailed analysis demonstrated that, in particular for AAV8, a part of these supplementary peptides observed in SDS-PAGE/Western blot was due to cleavage of vp1/vp2 by baculoviral cathepsin. The origin of other peptides was probably due to the codon-optimization performed during the development of the dual baculovirus system for the generation of AAV vectors (Smith et al. (2009) Mol. Ther. 17, 1888). Concerning the cleavage by cathepsin activity, we could show that the addition of the specific inhibitor E64 prevented the cleavage of vp1/vp2. A similar susceptibility to cathepsin cleavage was observed for AAV serotypes 1 and 6. The cleavage site was identified as L(189)-G(190)-E(191) (AAV8 amino acid numbering) and it could also be shown that the replacement of L by I or E by Q (as established for serotypes 2 and 9/10, for instance) led to resistance towards cleavage by cathepsin. Since it is not feasible to use E64 at a large scale, two strategies have been evaluated for alleviating this 'cleavage' problem: (i) mutation of single amino acids to render the cleavage site 'non-cleavable', and (ii) use of a cathepsin knock-out baculovirus. Whereas the first strategy led only to a shift of the problem to hidden downstream cleavage sites, the second approach completely alleviated this problem, resulting in AAV preparations with a higher (about 3x) in vivo potency in mice than when using the nonmodified baculovirus system. The paper will present details on the generation of supplementary peptides as seen in Western blot analysis either caused or not by baculoviral cathepsin activity.

A challenge of constructing insect/mammalian shuttle vector using two constitutive virus-based promoters

3:00 •317•

N. ElSahly, T.Z. Salem

Biomedical Sciences, University of Science and Technology, Zewail City, 12588, Giza, Egypt

Corresponding author: Isalem@zewailcity.edu.eg

It is quite often that a gene is required to be stably or transiently expressed in one of the baculovirus host cells such as Sf9, using the constitutive promoter OpIE2. In addition, a transient expression in mammalian cells is commonly driven by CMV promoter. However, two separate vectors would have been constructed if the same gene is required to be expressed in both types of cell lines (insect and mammalian cells). Here we are trying to construct a shuttle vector that can support the constitutive expression of transgenes in two cell lines of different species (Sf9 and Hela cells) using two of the aforementioned constitutive promoters. It was reported previously that the presence of ATGs in the second promoter of a shuttle vector eliminates its candidacy from being used. Accordingly, a truncated version of the OpIE2 includes only one ATG was cloned downstream of a wild type CMV promoter and upstream of the gene under test: enhanced green fluorescent protein (EGFP). The remained ATG was subjected to all possible nine single point mutations. Each of the mutated copies resulted in different expression level of the gene under test, in both sf9 and Hela cell lines. Although the nine mutated OpIE2 have no ATG remained, each mutated clone derived a variable expression level of EGFP in Sf9 and each has a negative, yet variable, impact on the gene expression in Hela cells. Further experiments are needed to decipher the reason for this variable expression patterns in both Sf9 and Hela cells.

Using recombinant baculovirus AcMNPV as a safe drug finding platform

3:15 •318•

3:30

Tzong-Yuan Wu¹, Chun-Chung Chen¹, Ming-Kun Liu²

Department of Bioscience Technology, Chung Yuan Christian University, Chungli; ²Institute of Bioinformatics and Structural Biology, National Tsing Hua University, Hsinchu 300, Taiwan Corresponding author: tywu@cycu.edu.tw

Previous studies have found that the addition of HDACis could significantly increase the baculovirus-mediated exogenous gene expression in mammalian cells. Based on this phenomena, we hypothesize that baculovirus could be designed to screen candidate drugs for HDACis. In this study, we test the hypothesis, using baculovirus-transduced cells and histone (acethyl Lys5, Lys8, Lys12, Lys16) antibody to identify a compounds that can enhance baculovirus-mediated gene expression in U2OS cells via stabilizing the hyperacetylation state of histone and implied that Bacmam could be a drug screening platform for HDACis. Besides, we also incorporated alphavirus structure proteins and replicon components into the Bacmam and used these recombinant baculoviruses to screen compounds that can interfere the entry or replication of alphavirus.

Inhibition of melanization by serpin-5 and -9 promotes nucleopolyhedrovirus infection in Helicoverpa armigera

Chuanfei Yuan¹, Longsheng Xing², **Manli Wang**¹, Xi Wang¹ Mengyi Yin¹, Qianran Wang¹, Zhihong Hu¹, Zhen Zou²

> ¹State Key Laboratory of Virology, Wuhan Institute of Virology, Chinese Academy of Sciences, Wuhan, China ² State Key Laboratory of Integrated Management of pest Insects nd Rodents, Institute of Zoology, Chinese Academy of Sciences, Beijing, China Corresponding author: huzh@wh.iov.cn, zouzhen@ioz.ac.cn

Melanization, an important insect defense mechanism, is mediated by clipdomain serine protease (cSP) cascades and is regulated by serpins. Here we show that proteolytic activation of prophenoloxidase (PPO) and PO-catalyzed melanization kill the nucleopolyhedrovirus (NPV) in vitro. Our quantitative proteomics revealed that NPV infection of the cotton bollworm, Helicoverpa armigera, reduced levels of most cascade members in the host hemolymph and PO activity. By contrast, serpin-9 and serpin-5 were sequentially upregulated after the viral infection. The H. armigera serpin-5 and serpin-9 regulate melanization by directly inhibiting their target proteases cSP4 and cSP6, respectively and cSP6 activates PPO purified from hemolymph. Furthermore, serpin-5/9-depleted insects exhibited high PO activities and showed resistance to NPV infection. Together, our results characterize a part of the melanization cascade in H. armigera, and suggest that natural insect virus NPV has evolved a distinct strategy to suppress the host immune system.

3:00-3:30 pm **Refreshments Break**

THURSDAY, 6:30 pm - 12:30 am

Estancia La Jolla Hotel

Cocktail Hour 6:30

7:30 Banquet

Awards Ceremony Live Band and Dancing

The Estancia La Jolla Hotel is a 15 minute walk from the Village (same distance as Price Center).

A shuttle will circle between the Village and the banquet starting at 6:15 from the Village.

Return shuttles from the banquet to the Village will run as needed, starting at about 9 pm. The last bus leaves the banquet site at 12:30 am.

We look forward to seeing you at the 51st Annual Meeting of SIP!

12-16 August 2018 Gold Coast, Australia

Contact: Caroline Hauxwell (caroline.hauxwell@qut.edu.au)

INDEX OF AUTHORS

Presenting authors and the abstracts they are presenting are listed in **BOLDFACE ITALICS**.

Papers lacking appended numbers can be found in the program by the Division oral paper session or by the Division poster number. Divisions abbreviations are the following **Bac** (Bacteria), **DBI** (Diseases of Beneficial Invertebrates), **Fun** (Fungi),

Mcr (Microbial Control), Msp (Microsporidia), Nem (Nematodes), and Vir (Virus).

AAA		Banerjee, R	137	Bonilla, CM	55
Abd-Alla, AMM	117, 118	Bannach, C	95	Bonnin, D	316
Abdelbasset Hassan, MM		Banyuls, N	120, 213	Bonning, B	209, 210
	Poster Fun24,	Barajas-Mendoza	234	Boogaard, B	161
	Poster Mcn26	Barelli, L	112	Booth, R	126
Abdelgaffar, H	74	Barrera, GP	183, 184	Borba, R	84
Abraham, A	125	Barreto, LP	244	Borgemeister, C	248
Abreo, E	14	Barros, MM	30	Boutin-Fontaine, M	316
Adegawa, S	212	Barry, J	126	Bowling, AJ	211
Ahmad, JN	268, Oral	Barthel, A	76	Bradshaw, C	169, Poster
•	Mcn#2	Bartholomay, LC	34	•	Bac26
Ahmad, SJN	Oral Mcn#2	Bashir, EM	Poster Fun23	Bravo, A	43, 44 , 47,
Ahmad, ST	268	Bass, D	115, 296	·	122, 207
Ahuja, I	Oral Mcn#2	Bateman, K	10, 66, 128	Bressan, A	220
Airs, PM	34	Bauchan, GR	20	Broadley, HJ	18
Aiuchi, D	265	Bayram, S	77, 162	Broglie, R	89
Akiau, DM	275	Becnel, JJ	3, 26, 127 ,	Brownbridge, M	91, 306
Akülke, AZI	149	Bednarz, H	269	Bruck, DJ	89
Al Souhail, Q	237	Bei, Y		Brutscher, L	38
· ·		Bei, Y Beilinson, V	120, 263	•	
Alli, H	49 51	•	151	Büchel, K	276 67
Allahyari, H	51	Beitzen-Heineke, E	92, 270, 271,	Buchon, N	67
Allen, C	95	5	272	Buisson, C	124
Ally, D	310	Beitzen-Heineke, W	92, 270, 271,	Burand, JP	18, 20
Almeida, ZG	201		272	Burge, CA	116, 305
Altier, N	14, 282	Belaich, MN	183, 184	Bush, M	220
Alves, DDS	255	Benatto, A	261	Butler, C	222
Alves, VS	255	Bennett, KA	Poster Bac23		
Anderson, T	53	Bennison, J	90	CCC	
Andrade, MC	192	Benuzzi, M	308	Cai, Wanzhi	80
Andrade, MS	190	Bergamini, C	231	Cai, Yi	99
Antwi, FB	279	Berger, M	245, 246	Caixeta, CF	201
Anyanga, MO	88	Bergh, JC	253	Cali, A	4
Aquilino, KM	113	Berlitz, DL	62	Caner, E	77
Arai, E	164	Bermudez, E	49	Carballar-Lejarazú, R	217
Arai, H	199	Bernardo, CC	244	Cardoso, CP	206
Araque, GA	276	Berón, CM	203	Carrière, Y	73, 150
Ardisson-Araújo, D	62, 93, 190	Berry, C	303	Carrillo, D	157
Arensburger, P	143	Bertin, B	316	Castillo-Esparza, JF	139
Arias, M	283	Beyhaut, E	14	Castrillo, LA	54
Arinto-Garcia, R	188	Beys-da-Silva, WO	245, 246	Cavallo, E	227
Aroian, RV	125	Bideshi, DK	75, 142 , 164	Cave, RD	157
Arruda, J	119	Bidochka, MJ	112	Chabaud, M-A	109
Arruda, W	244	Biganski, S	25	Chacha, M	52
Asano, S-I	265	Bigot, Y	142	Chakroun, M	120
· · · · · · · · · · · · · · · · · · ·			152	Chan, L	
Ashby, M	89	Bilgo, E			Poster Bac2
Asokan, R	140	Bishwajeet, P	280	Chandler, D	61, 90 , 155,
Avery, PB	157	Bissinger, B	88	Chang Dereit	174, 235
Awando, JA	21	Bittencourt, VREP	245, 275, 277	Chang, Dennis	30, 256
0.00		Bivián-Hernández, MdlA	192	Chang, Ju-Chun	193
BBB	202	Blackburn, M	210	Chang, Zi-Ting	19
Báez, F	283	Blissard, GW	67	Chaput, D	296
Bai, Shuxiong	80	Blum, LEB	216, 249, 250	Chateigner, A	16, 182
Baldo, GR	261	Boiteux, JJ	Oral Fun#3	Chawla, S	166
Baldwin, D	77, 181	Bojko, J	296	Chejanovsky, N	311
Daidwiii, D	•	• •			
Baldwin, J Balloni, A	30	Bojórquez-Ramos, C Bones, A	55 Oral Mcn#2	Cheng, Chen Chen, Chun-Chung	Oral Vir#1

Chen, Claurous	Chen, Chun	236	de Souza, ML	185, Poster	Feng, Guozhong	94
Chen, Yu-Bin 19	Chen, Guoqing	94		Vir20	Fernandes, EKK	106, 230, 242,
Chen, Ya-Bilin (burly) 82 Debits, 2009 Ferrer, J 200, 210 Chene, Ya-Will 164 det Rhuck-Castro, MC 192 Ferrer, J 208, 313 Chen, Xa-Willian 194 Deblabera Juna, 1 15 Ferreira, DG 255 Chol, Jack Young 214, 239, 258 Deligado, A 283 Ferreira, DG 255 Chol, Jack Young 114 Hall Chol, Jack Young 144 Filipii, T 144 144 Chol, Jack Young 144 Filipii, T 144 144 Chol, Jack Young 144 Filipii, T 144 144 Chor, Jack Young 144 Filipii, T 144 144 Chor, Jack Young 144 Pilipii, T 144 144 Pilipii, T	Chen, Jianwu	166, 168	de Souza, DA	233		244
Chen, Yaminq Indudy) 82 Debnort, MH \$1,259 Ferre, In 2011, 123, 208, 213 China, Keu Ping 19 Defalbrea Innor, 1 15 Ferreira, T 25 China, Keu Ping 214, 239, 258 Delagado, A 283 Ferreira, T 28 Claus, Jo 312 Deng, Shaul 314 Filink, C 45 Clerkin, S 205 Diao, Gingyan 314 Filink, C 46 Closek, G 11 Dioc, Say 316 Renniken, M 38 Contestino, L 48,49 Dioch, Sh 125 Portastin, I 72,227,228 Cong, R 48,49 Dieth, Sh 125 Portastin, I 92,227,228 Cong, R 48,49 Dieth, Sh 125 Portastin, I 92,127,228 Cong, R 48,49 Dieth, Sh 125 Portastin, I 92,227,228 Cong, R 48,49 Dieth, T 72 Frederication, I 92 Corristancia, TA 275 Debretty, L 296 Fred	Chen, Tzu-Han	19	Dedkhad, W	34	Fernandes, TA	255
Cent. No. Section Se	Chen, Ya-Bin	165	Deist, B209		Fernandez-Luna, MT	209, 210
Cinia (see Ping) 19 Debalbare Aumon.1 15 Ferrein, DC 25 Calson, DC 25 Calson, DC 28 Calson, DC 312 Deng, Shuai 314 Ferrein, DC 28 Calson, DC 44 Calson, DC 45 Calson, DC 45 Calson, DC 45 Calson, DC 45 Calson, DC 45 Calson, DC 45 Calson, DC 45 Calson, DC 45 Calson, DC 46 Calson, DC 46 Calson, DC 46 Calson, DC 46 Calson, DC 46 Calson, DC 46 Calson, DC 46 Calson, DC 46 Calson, DC 46 Calson, DC 46 Calson, DC 46 Calson, DC 46 Calson, DC 46 Calson, DC 46 Calson, DC 46 Calson, DC 46 Calson, DC 47 Calson, DC <td>Chen, Yanping (Judy)</td> <td>82</td> <td>Dekhordi, MH</td> <td>51, 259</td> <td>Ferre, J</td> <td>120, 121, 123,</td>	Chen, Yanping (Judy)	82	Dekhordi, MH	51 , 259	Ferre, J	120 , 121, 123,
Chol, Jan Young 214, 239, 258 Delgado, A 283 Forteria, T 284 Cleun, R 198 Diabate, A 152 Filia, C 43 Cleun, R 198 Diabate, A 152 Filia, C 43 Clerkin, S 205 Diac, Gingyun 314 Filoza, L 62 Closek, Cl 116 Dicks, Y 316 Filoza, L 24 Consert, G 238 Diego Nava, F 252 Folgardi, Pl 75, 727, 228 Consert, G 48 49 Dielin, SH 126 Fordestra, I Poster, Pl Consert, G 48 49 Dielin, SH 126 Fordestra, I Poster, BR Contest, G 183 Dillon, A 200 Francesthin, S 308 Contest, G 186 Dimitrui, T 72 Freeman, AM 241 Corries, TA 275 Doherty, L 296 Freeman, AM 201 201 Corries, TA 275 Doherty, L 296	Cheng, X-W	164	del Rincón-Castro, MC	192		208, 213
Chol, Jae Young 214, 239, 258 Delgado, A 283 Forteria, T 28 Cleux, R 198 Diabate, A 152 Filk, C 45 Cleur, R 198 Diabate, A 152 Filke, C 45 Clerkin, S 205 Diabo, Clingyun 314 Filke, C 42 Citton, EH 238 Diac Sainhez, M 168 Filegard, M 38 Coates, S 238 Dilego Nava, F 252 Folgardi, PJ 78, 727, 228 Cong, R 48, 49 Dilbin, SH 126 Fordestro, J Poster Boc27 Contentra-Gardulo, J 124 Dillian, AR 30, 256 Fowell, M 241 Conters, G 166 Dimitru, T 72 Fredenstro, S 308 Conters, G 167 Dimitru, T 72 Fredenstro, S 308 Conters, G 275 Dobrery, L 296 Fryan, G 119 Corres, G 275 Dobrery, L 292 Friegar, R 119	=	19	Delalibera Junor, I	15	Ferreira, DG	
Clause, No		214, 239, 258	·		·	
Clem, R 198 Diabate, A 152 Finke, C 45	_		_			
Certin, S 205 Diao, Qingyun 314 Fiuz, L 62 Clifton, EH 238 Diaz-Sánche, M 188 Fige, L M 24 Closek, CJ 116 Dicks, Y 316 Fige, L M 38 Closek, CJ 116 Dicks, Y 316 Fige, L M 38 Coates, 85 238 Diego Nava, F 252 Folgarati, PJ 37, 227, 228 Cong, R 48, 49 Diehn, SH 126 Folgarati, PJ 37, 227, 228 Cong, R 48, 49 Diehn, SH 126 Folgarati, PJ 37, 227, 228 Cong, R 48, 49 Diehn, SH 126 Forwell, M 241 Conternor, Gradulo, J 83 Dillon, A 260 Fraceschini, S 308 Contreras, E 166 Dimitriu, T 72 Fredensborg 64 Coon, KL 173 Dippel, C 270 Freeman, AM Oral DBI 42 Correia, TA 275 Doberty, L 296 Frey, M 119 Corson, E 49 Dorrington, RA 21, 22 Friedman, CS 114, 116 Courtes, ST 138 Downs, S 208 Frey, M 119 Courtes, ST 138 Downs, S 208 Fuertes, VB 157 Courton, T 251 Dubois, T 290 Fuertes, VB 157 Courton, T 251 Dubois, T 290 Fuertes, VB 157 Courton, T 251 Dubois, T 290 Fuertes, VB 157 Courton, T 251 Dubois, T 290 Fuertes, VB 215 Court, G 126 Duman, RH 157 Courthe, Downs, S 235 Galaini-Wraight, S 4 Couth, C 77 Dwyer, S 235 Galaini-Wraight, S 4 Cov, GA 259 Cov, GA 259 Cov, GA 259 Fuertes, B 207 Galaini-Wraight, S 4 Cov, GA 259 Fuertes, B 207 Gandro, P 217 Cov, Foster, D 260 Galaini-Wraight, S 28 Crosson, LM 114 Ellin, G 90 Correca-Media, M 159 Elbing, J 200 Gardro, P 218 Crosson, LM 114 Ellin, G 90 Correca-Media, M 159 Elbing, J 200 Gardro, P 218 Correca-Media, M 159 Elbing, J 200 Gardro, P 218 Correca-Media, M 159 Gardro, MA 219 Gardro, P 219 Correca-Media, M 259 Excise, B 207 Gardro, P 219 Correca-Media, M 259 Excise, B 207 Gardro, P 219 Correca-Media, M 259 Excise, B 200 Gardro, P 227 Correca-Media, M 259 Gardro, MA 250 Gardro, P 277 Corre	•		•		-	
Cirtion, EH 238 Diac Sachner, M 168 Flequit, TW 24 Coates, BS 238 Digco, Y 31 6 Flentiken, M 38 Coates, BS 238 Diepn, SH 126 Fordiagnit, PI 57, 227, 228 Connertar Gordufio, J 84 90 Dilon, A 250 Fordiagnit, PI 522 Contrera-Gordufio, J 83 Dillon, A 250 Fordiagnit, PI 231 Contress, E 166 Dimitriu, T 72 Fredemsborg 64 Corrica, TA 275 Debrety, L 296 Frey, M 119 Corrica, TA 275 Debrety, L 296 Frey, M 119 Corrica, TA 275 Debrity, L 296 Frey, M 119 Corrica, TA 49 Oprrington, RA 21, 22 Friedman, S 114, 116 Cortes, TA 49 Oprrington, RA 22, 22 Friedman, S 114, 116 Cotte, Stephanite, C 251 Obbotis, T 20 20 20 </td <td>•</td> <td></td> <td>•</td> <td></td> <td>· ·</td> <td></td>	•		•		· ·	
Closek, C. 116	•					
Coates, BS 238 Depo Nava, F 252 Folgarnit, PI 57, 227, 228 Cong, R 48, 49 Diehn, SH 126 Fornstrui, I 922 Consentino, L 124 Dilloma, AR 30, 256 Fowell, M 241 Conteres, Gradullo, J 83 Dilloma, AR 30, 256 Frowell, M 241 Conteres, Gradullo, J 133 Dilloma, AR 200 Freedensborg 64 Conne, M. 173 Dippel, C 270 Freedensborg 64 Corrol, S. 49 Dorrington, RA 21, 22 Friedman, C 113, 116 Corrol, S. 182 dos Santos, NA 262 Fritch, E 8 157 Court, S. 138 Dorbers, S 208 Fullow, R 315 157 Court, C. 231 Dubore, S 208 Fullow, R 315 157 Court, C. 241 Durran, RA 275 2245 Galaini-Wraight, S 54 Court, S. 238 <th< td=""><td></td><td></td><td>·</td><td></td><td>• •</td><td></td></th<>			·		• •	
Cong, R. 48, 49 Diehn, SH 126 Fordstro, I Poster Bac27 Consentino, I. 124 Dillman, AR 30, 256 Franceschini, S 308 Contrerae, E. 166 Dimitriu, T 72 Fredmontorg 6 Conor, K. 173 Dippe, C 270 Freeman, AM Oral DBI 92 Corrais, TA 275 Doherty, L 296 Frey, M 119 Corris, TA 292 Doherty, L 296 Frey, M 119 Corris, TA 252 dos Sontos, MA 262 Firthsh. E 85 Contrés-Martine, C 252 dos Sontos, RF 204, 205, 206 Fuentes, VB 137 Coute, G 182 dos Sontos, RF 294, 205, 206 Fuentes, VB 137 Coute, G 183 Downs, S 208 Fullaga, R 315 Coute, G 265 Douncan, RH 157 Full Blan, AR 315 Couti, C 70 Deg, S 235 Galain-Wraight, S 56 <td></td> <td></td> <td>•</td> <td></td> <td>· · · · · · · · · · · · · · · · · · ·</td> <td></td>			•		· · · · · · · · · · · · · · · · · · ·	
Consention L Contreva Sardardio, J 88 241 billman, AR 260 Franceschinl, S 308 Contreva Se Contreva Se 166 Dillman, AR 260 Franceschinl, S 308 Contreva Se Contreva Se 166 Dillmitriu, T 72 Fredensborg 64 Contreva Se Contreva	•		-		• •	
Contreros, Garduño, J 83 Dillon, A 260 Franceschini, S 308 Conteros, E 166 Dimitriu, T 72 Freemshorg 64 Con, KL 132 Dippel, C 270 Freeman, AM Oral Del Bt 2 Correia, TA 275 Doherty, L 296 Fey, M 119 Correia, TA 275 Dohor, S 208 Freeman, AM C 114, 116 Cortes, Martiner, C 252 dos Santos, NA 26, 22 Friedman, CS 114, 116 Courte, SC 138 Downs, S 208 Fulftee, R 315 Couter, T 251 Dubós, T 299 Fufftee, R 315 Coutro, Toster, D 60 de Silva, CSR 235 Galaini-Wrinjth, S 54 Cox, GA 299 Fufftee, R 316 36 Cox, GA 299 God Galaini-Wrinjth, S 54 Cox, GA 299 God Galaini-Wrinjth, S 54 Cox, Gaster, D 41,772,044 Elbing,	=		•		·	
Contrexs, E 166 Dimitriu, T 72 Fredensborg 64 Cono, KL 173 Dippel, C 270 Freeman, AM Oral DBI 42 Corson, E 49 Doherty, L 296 Frey, M 119 Corson, E 49 Dormigtor, RA 21, 22 Fritsch, E 85 Corv, J. S 182 dos Santos, RA 204, 205, 206 Fuentes, VB 157 Coutry, T 138 Downs, S 208 Fulfre, R 315 Coutron, T 251 Duncan, RH 157 Funchs, VB 59 Coutro, T 41 Duncan, RH 157 Cotribor, G GG G G Coutu, C 77 Dwer, S 235 Galain-Wrraight, S 54 Cox-Foster, D 60 de Silva, CSR 244 Galbert, L 316 Cox-GA 259 EE E E Galain-Wraight, S 54 Cox-Goster, D 126 EE E E Galain-Wraight, MRI 192 Crane, WC 127	·		· ·	•	•	
Coon, K. 173 Dippel, C 270 Freeman, AM Oral D8I H2 Correia, TA 275 Doherty, L 296 Free, M 119 Corson, E 49 Dorrington, RA 21, 22 Friedman, CS 114, 116 Cortes-Martinez, C 292 dos Santos, RA 262 Friedman, CS 114, 116 Corter, SC 138 Downs, S 208 Fluither, R 315 Court, T 41 Duncian, RH 157 Inithaus, A 59 Court, C 77 Dwyer, S 235 Galaini-Wraight, S 4 Cox, Ga 295 EST Galaini-Wraight, S 4 Cox, Ga 295 EST Galaini-Wraight, S 4 Cox, Ga 295 Ester Galaini-Wraight, S 4 <	· · · · · · · · · · · · · · · · · · ·		•		•	
Correia, TA 275 Oberty, L 296 Frey, M 119 Corson, E 49 Obrington, RA 21, 22 Friedman, CS 114, 116 Cortes, Martinez, C 252 dos Santos, NA 262 Fritsch, E 85 Corty, JS 182 dos Santos, RF 204, 205, 206 Fuentex, VB 157 Coute, C 138 Downs, S 208 Pulyto, R 315 Courton, T 211 Dubois, T 290 Furnhaus, A 35 Courton, T 41 Duncar, RH 157 Furnhaus, A 315 Coutin, C 77 Duyer, S 235 Galain-Wraight, S 54 Coutin, C 77 Duyer, S 235 Galmer, C 1316 Cox, GA 259 E Galain-Wraight, S 54 Cran, QC 117, 2,204, Bling, J EE Galing, J 63 64 Cran, WC 117, 2,204, Bling, J 8, 50, 64 Garcha-Arree, G 136 62 Cran, WC	· · · · · · · · · · · · · · · · · · ·		· · · · · · · · · · · · · · · · · · ·		•	
Corson, E 49 Operington, RA 21, 22 Friedman, CS 114, 116 Cortés-Martinez, C 252 dos Santos, RF 204, 205, 206 Fluentes, VB 157 Corter, SC 138 Downs, S 208 Fulthes, VB 315 Courter, SC 138 Downs, S 208 Fulthaus, A 59 Court, T 41 Duncan, RH 157 Turbles, VB Fullhaus, A 59 Coutlon-Rodrigues, C 245 Dundor-Arias, JP 299 GG G C Cout, C 77 Dwyer, S 235 Galain-Wraight, S 54 Cox, GA 299 Elster GG G Galvin, MFL 192 Crane, VC 126 EEE Galvin, MFL 192 Crase, VC 116 Elling, J 59 Gandra, P 211 Crosson, LM 114 Ellenberg, J 8, 50, 64 Garcia-Arrae, C 155 52, 522 Crosson, LW 126 Elsain, M 280 Garcia-Arrae, G			11 /		·	
Cortes-Martínez, C 252 dos Sontos, NA 262 Fritsch, E 85 Corty, S 182 dos Sontos, RF 204, 205, 206 Fuentes, VB 157 Coutron, T 251 Dubois, T 290 Furthaus, A 315 Coutron, T 41 Dundore, Arias, P 299 Ternhaus, A 5 Coutro, C 77 Dwe, S 235 Galialin-Wraight, S 54 Cox-Foster, D 60 de Silva, CSR 244 Galibert, L 316 Cox, GA 259 EEE Gamer, C 157 Crane, VC 126 EEE Gamer, C 157 Criskmore, N 1,172, 204 Ebling, J 59 Gandra, P 211 Crus-Lopez, V 188 Ekstein, B 207 Gangra, Yuanthu 187 Crus-Lopez, V 188 Ekstein, B 207 Garcia-Grutkfree, C 55, 252 Cuenca-Medina, M 153 Elbashir, MI 280 Garcia-Grutkfree, C 55, 252 Dariu,	•		• •		· ·	
Cory, IS 182 dos Santos, RF 204, 205, 206 Fuentes, VB 157 Coutro, T 251 Dubois, T 290 Fürfhaus, A 59 Coutro, T 251 Dubois, T 299 Fürfhaus, A 59 Coutu, C 77 Dwyer, S 235 Galaini-Wraight, S 54 Cox, Foster, D 60 de Silva, CSR 244 Galbert, L 316 Cox, GA 259 EEE Galvan, MFL 192 Crane, VC 126 EEE 59 Gandro, P 211 Crickmore, N 41, 72, 204 Eblenberg, J 8, 50, 64 Gorcia-Grave, C 157 Crosson, LM 114 Ellenberg, J 8, 50, 64 Gorcia-Gutlefree, C 55, 252 Cuenca-Media, M 153 Elbashir, MI 280 Gorcia-Gutlefree, C 55, 252 Cuenca-Media, M 153 Elkinton, JS 18 Gorcia-Arnace, G 136 Curcic, I 260 Elkinton, JS 18 Gorcia-Arnace, G 139	·		=		· ·	
Cotter, SC 138 Downs, S 208 Fujita, R 315 Coudron, T 251 Dubbois, T 299 Curry, T 41 Dundore, Arias, JP 299 GGG Coutry, T 77 Dwey, S 235 Gallain-Wraight, S 54 Cox, GA 259 - Gallain-Wraight, S 192 Crane, VC 126 EEE Gallain-Wraight, S 192 Crickmore, N 41, 72, 204, Bbling, J 59 Gandra, P 211 Crosson, LM 114 Ellenberg, J 8, 50, 64 Gorcia-Arrae, G 136 Cuenca-Medina, M 153 Elbshir, MI 280, Garcia-Montelongo, M 139 Cuenci, I 260 Poster Fun24 Garcia-Suare, R 139 Dahiru, E 115 Ellishir, MI 280 Garcia-Montelongo, M 139 Cuenca-Medina, M 153 Elbashir, MI 280 Garcia-Montelongo, M 139 Dahiru, E 1 Elishir, M 317 Gauthier, D 225 <	· · · · · · · · · · · · · · · · · · ·		<u>-</u>		•	
Coudron, T 251 Dubois, T 290 Fünfhaus, A 59 Coutty, T 41 Duncan, RH 157 Coutt, C 77 Dwyer, S 235 Galaini-Wraight, S 54 Cox-Foster, D 60 de Sliva, CSR 244 Galibert, L 316 Cox, GA 259 EE E Galibert, L 316 Crane, VC 126 EE E Gamdra, P 211 Crickmore, N 41, 72, 204 Ebling, J 59 Gandra, P 211 Crosson, LM 114 Ellenberg, J 8, 50, 64 Garcia-Graver, G 136 Cruz-López, V 168 Ekesi, S 248 Garcia-Guerierre, C 55, 252 Cuenca-Nedina, M 153 Elbashir, MI 280 Garcia-Guerierre, C 55, 252 Cuenca-Nedina, M 153 Elbashir, MI 280 Garcia-Guerierre, C 55, 252 Damode, H 126 Elsahir, MI 317 Garcia-Montellong, MI 139 Damiu, E 41			<u>-</u>			
Courty, T 41 Dundan, RH 157 Coutin, C 77 Dunder-Arias, JP 299 GGG Courto,	•		•			
Coutin, C Coutin, C Coutin, C Coutin, C Coutin, C Coutin, C Coutin, C Cox Foster, D Coutin, C Cox, Galani-Wraight, S Cox Galani-Wraight, S Cox Galani-Wraight, S Cox Galani-Wraight, S Cox Galani-Wraight, S Cox Galani-Wraight, S Galani-Wraight, M I S Galani-Wraight, M I S Galani-Wraight, M I S Galani-Wraight, M I S Galani-Wraight, M I S Galani-Wraight, S Galani-Wra	•		,		Funthaus, A	59
Cour. Cox. Foster, D 60 de Silva, CSR 235 Galaini-Wraight, S 54 Cox. Foster, D 60 de Silva, CSR 244 Galibert, L 316 Cox, GA 259 EE E Galvian, MFL 192 Crane, VC 126 EE E Gamer, P 211 Crickmore, N 41, 72, 204, Ebling, J 59 Gandra, P 211 Crosson, LM 114 Ellenberg, J 8, 50, 64 Garcia-Arreac, G 136 Crour-Lope, V 168 Ekes, S 248 Garcia-Gutièrrez, C 55, 252 Cuenca-Medina, M 153 Elbashir, MI 280 Garcia-Montelongo, M 139 Curcic, I 260 Elkinton, JS 18 Garcia-Montelongo, M 139 Dairu, E 41 Ellis, CB 90 Gassman, A 238 Dairu, E 41 Ellis, CB 90 Gassman, A 238 Dairu, E 41 Ellis, CB 90 Gassman, A 238 <td>•</td> <td></td> <td>·</td> <td></td> <td>666</td> <td></td>	•		·		666	
Cox, Foster, D 60 de Silva, CSR 244 Galbert, L 316 Cox, GA 259 EEE Galván, MFL 192 Crane, VC 126 EEE Galván, MFL 192 Cricknore, N 41,72, 204, Ebling, J 59 Gandra, P 211 Crosson, IM 114 Elenberg, J 8,50,64 García-Muntelongo, M 136 Cruz-López, V 168 Ekesi, S 248 García-Gutiérrez, C 55, 252 Cuenca-Medina, M 153 Elbashir, MI 280 García-Montelongo, M 139 Curcic, I 260 Elkinton, JS 18 García-Montelongo, M 139 Dahiru, E 41 Ellis, CB 90 Gassman, AJ 238 Dama, SS 53, 87 Endo, H 46, 212 Gascub, D 225 Dara, SS 53, 87 Endo, H 46, 212 Gaszola, D 125 Dara, SS 53, 87 Endoson, MA 17, 65, 68, 77 Geiser, R 251	o ,		· · · · · · · · · · · · · · · · · · ·			
Cox, GA 259 EEE Galván, MFL 192 Crane, VC 126 EEE Gamez, C 157 Crickmore, N 41,72, 204, 205, 285 Eckstein, B 207 Gan, Vuanzhu 187 Crosson, LM 114 Bilenberg, J 8, 50, 64 Garcia-Aracez, G 136 Cruz-López, V 168 Ekesi, S 248 Garcia-Mortlelongo, M 139 Cucric, I 260 Poster Fun23, Poster Fun23, Poster Fun24 Garcia-Mortlelongo, M 139 Louric, I 153 Elbashir, MI 280, Garcia-Mortlelongo, M 139 Dariu, E 260 Poster Fun24, Garcia-Suare, R 139 Dariu, E 41 Ellis, CB 90 Gassmann, AI 238 Dahiru, E 41 26 ElSahly, N 317 Gauthier, D 225 Dara, SS 53, 87 Endo, H 46, 212 Gazcia, D 125 Dara, SS 53, 87 Erlandson, MA 17, 65, 68, 77, Geibert, R Geibert, R 251 Da	· ·		• •		<u> </u>	
Crane, VC 126 EFF Gámez, C 157 Crickmore, N 41, 72, 204, 205, 206, 285 Eckstein, B 207 Gao, Yuanzhu 187 Crosson, LM 114 Eilenberg, J 8, 50, 64 Garcia-Arroez, G 136 Cruz-López, V 168 Eksi, S 248 Garcia-Montelongo, M 139 Curcic, I 260 Elbashir, MI 280 Garcia-Montelongo, M 139 Curcic, I 260 Elkinton, JS 18 Garcia-Joure, R 139 Dahiru, E 41 Ellis, CB 90 Gassmann, AI 238 Damude, H 126 ElSishhy, N 317 Gauther, D 255 Dara, SS 53, 87 Erladson, MA 17, 65, 68, 77, Geisert, R 251 Dara, SS 53, 87 Erladson, MA 17, 65, 68, 77, Geisert, R 251 Dara, SSR 53, 87 Erladson, MA 17, 65, 68, 77, Geisert, R 251 Dara, SSR 53, 87 Erladson, MA 17, 65, 68, 77, Geisert, R 26lbit, 1 217	· ·		de Silva, CSR	244	•	
Crickmore, N 41, 72, 204, 205, 285 Eckstein, B 207 Gandra, P 211 Crosson, LM 114 Ellenberg, J 8, 50, 64 Garcia-Arraez, G 136 Cruz-López, V 168 Ekesi, S 248 Garcia-Gutiérrez, C 55, 252 Cuenci, 1 260 Elbashir, MI 280, Garcia-Suare, R 139 Curcic, I 260 Elkinton, JS 18 Gardes-Cu, S 281 Curcic, I 260 Elkinton, JS 18 Gardia-Jurado, I 179, 243 Dahiru, E 41 Elkinton, JS 18 Gardia-Jurado, I 179, 243 Dahiru, E 41 Elkinton, JS 18 Gardia-Jurado, I 179, 243 Damude, H 126 ElSahly, N 317 Gastmann, AJ 238 Dara, SS 53, 87 Erdad, H 46, 212 Gazzola, D 125 Dara, SS 53, 87 Erlandson, MA 17, 65, 68, 77, Geisert, R 251 Dari, SY 53, 87 Escaba, S 182 Genersch, E					· ·	
Crosson, LM 205, 206, 285 Eckstein, B 207 Gao, Yuanzhu 187 Crosson, LM 114 Ellenberg, J 8, 50, 64 Garcia-Arraez, G 136 Cruz-López, V 168 Ekesi, S 248 Garcia-Gutierrez, C 55, 525 Cuenca-Medina, M 153 Elbashir, MI 280 Garcia-Suare, R 139 Curcic, I 260 Elkinton, JS 18 Garrido-Jurado, I 179, 243 Dahiru, E 41 Elkinton, JS 18 Garrido-Jurado, I 179, 243 Dahiru, E 41 Elkinton, JS 18 Garrido-Jurado, I 179, 243 Dahiru, E 41 216 ElSahly, N 317 Gauthier, D 225 Dara, SS 53, 87 Endo, H 46, 212 Gazzola, D 125 Dara, SSR 53, 87 Endo, H 46, 212 Gazzola, D 125 Dara, SSR 53, 87 Endo, H 46, 212 Gazzola, D 125 Dara, SSR 53, 87 Escasa, S <th< td=""><td>· · · · · · · · · · · · · · · · · · ·</td><td></td><td></td><td></td><td>-</td><td></td></th<>	· · · · · · · · · · · · · · · · · · ·				-	
Crosson, LM 114 Ellenberg, J 8, 50, 64 Garcia-Arraez, G 136 Cruz-López, V 168 Ekesi, S 248 García-Montelongo, M 139 Curcic, I 260 Fibashir, MI 280, García-Montelongo, M 139 Curcic, I 260 Poster Fun23, Poster Fun23, Poster Fun24 García-Suare, R 139 Dahiru, E 41 Ellishton, JS 18 Garrido-Jurado, I 179, 243 Damude, H 126 ElSahly, N 317 Gauthier, D 225 Dara, SK 53, 87 Endo, H 46, 212 Gazzola, D 125 Dara, SS 53, 87 Erlandson, MA 17, 65, 68, 77, Geisert, R 251 Dara, SSR 53, 87 Erlandson, MA 161, 181, 182 Gelbic, 1 217 da Silva, EYY 200, 201 Escasa, S 182 Genersch, E 10, 27, 59 Dautel, H 271 Espinel, C 183, 276 Gill, 181, 182 Gelbic, 1 183, 184 Davids, M Poster Bac23 Estrela, JLV	Crickmore, N		=		· · · · · · · · · · · · · · · · · · ·	
Cruz-López, V 168 Ekesi, S 248 García-Gutiérrez, C 55, 252 Cuenca-Medina, M 153 Elbashir, MI 280 García-Montelongo, M 139 Curcic, I 260 Poster Fun23, Poster Fun23, García-Suare, R 139 Dariu, I D D D Elkinton, JS 18 Gardescu, S 281 Dahiru, E 41 Ellis, CB 90 Gassmann, AJ 238 Damude, H 126 ElSahly, N 317 Gauthier, D 225 Dara, SK 53, 87 Erlandson, MA 46, 212 Gazzola, D 125 Dara, SS 53, 87 Erlandson, MA 47, 65, 68, 77, Geisert, R 251 Ga Silva, EYY 200, 201 Escasa, S 182 Genbic, I 217 da Silva, EYY 200, 201 Escabar, BF 209, 210 Gerber, R 10, 27, 59 Daytan, M 259 Escobar, BF 209, 210 Gerber, R 126 Daughenbaugh, K 38 Escriche, B 120, 121, 263 Ghiringhelli, PD		205, 206, 285	Eckstein, B	207	Gao, Yuanzhu	187
Cuenca-Medina, M 153 Elbashir, MI 280, Poster Fun23, Poster Fun23, Garcia-Suare, R 139 Curcic, I 260 Elkinton, JS 18 Garcia-Suare, R 139 D D D Elkinton, JS 18 Gardrido-Jurado, I 179, 243 Dahiru, E 41 Ellis, CB 90 Gassmann, AI 238 Damude, H 126 ElSahly, N 317 Gauthier, D 225 Dara, SK 53, 87 Endo, H 46, 212 Gazzola, D 125 Dara, SS 53, 87 Endo, H 46, 212 Gesert, R 251 Dara, SSR 53, 87 Erlandson, MA 17, 65, 68, 77, Geisert, R Cellic, I 217 da Silva, EYY 200, 201 Escas, S 182 Genersch, E 10, 27, 59 Dastranj, M 259 Escobar, BF 209, 210 Gerber, R 126 Dautel, H 271 Espinel, C 183, 276 Gill, SS 47, 166, 168 Davids, IW Poster Bac23 Evans, JD 82 Gill, SS	Crosson, LM	114	Eilenberg, J	8, 50 , 64	Garcia-Arraez, G	
Curcic, I 260 Foster Fun23, Poster Fun24 Garcia-Suare, R Garcia-Sua	Cruz-López, V	168	Ekesi, S	248	García-Gutiérrez, C	55 , 252
Poster Fun24 Gardescu, S 281 D D D D Elkinton, JS 18 Garrido-Jurado, I 179, 243 Dahiru, E 41 Ellis, CB 90 Gassman, AJ 238 Damude, H 126 ElSahly, N 317 Gauthier, D 225 Dara, SK 53, 87 Endo, H 46, 212 Gazzola, D 125 Dara, SS 53, 87 Erlandson, MA 17, 65, 68, 77, Geisert, R 251 Dara, SSR 53, 87 Erlandson, MA 17, 65, 68, 77, Geisert, R 251 Dara, SSR 53, 87 Escaba, S 182 Genersch, E 10, 27, 59 da Silva, EYY 200, 201 Escasa, S 182 Genersch, E 10, 27, 59 Dastranj, M 259 Escobar, BF 209, 210 Gerber, R 126 Dautel, H 271 Espinel, C 183, 276 Gill, SS 47, 166, 168 David, M Poster Bac23 Estrela, JLV 185 Giolo, SR 261 Davis, J 88 Eva	Cuenca-Medina, M	153	Elbashir, MI	280,	Garcia-Montelongo, M	139
Elkinton, JS 18 Garrido-Jurado, I 179, 243 Dahiru, E 41 Ellis, CB 90 Gassmann, AJ 238 Damude, H 126 Ellis, CB 90 Gasthier, D 225 Dara, SK 53, 87 Endo, H 46, 212 Gazzola, D 125 Dara, SS 53, 87 Erlandson, MA 17, 65, 68, 77, Geisert, R 251 Dara, SSR 53, 87 Erlandson, MA 17, 65, 68, 77, Geisert, R 251 Dara, SSR 53, 87 Erlandson, MA 17, 65, 68, 77, Geisert, R 251 Dard Silva, EYY 200, 201 Escasa, S 182 Genersch, E 10, 27, 59 Dastranj, M 259 Escobar, BF 209, 210 Gerber, R 126 Dautel, H 271 Escriche, B 120, 21, 263 Ghiringhelli, PD 183, 184 David, M Poster Bac23 Estrela, JLV 185 Gill, SS 47, 166, 168 Davis, IW Poster Bac23 Evans, D 82 Glare, TR<	Curcic, I	260		Poster Fun23,	Garcia-Suare, R	139
Dahiru, E 41 Ellis, CB 90 Gassmann, AJ 238 Damude, H 126 ElSahly, N 317 Gauthier, D 225 Dara, SK 53, 87 Endo, H 46, 212 Gazzola, D 125 Dara, SS 53, 87 Erlandson, MA 17, 65, 68, 77, Geisert, R 251 Dara, SSR 53, 87 Erlandson, MA 17, 65, 68, 77, Geisert, R 251 da Silva, EYY 200, 201 Escasa, S 182 Genersch, E 10, 27, 59 Dastranj, M 259 Escobar, BF 209, 210 Gerber, R 126 Daughenbaugh, K 38 Escriche, B 120, 121, 263 Ghiringhelli, PD 183, 184 Dautel, H 271 Espinel, C 183, 276 Gill, SS 47, 166, 168 David, M Poster Bac23 Estrela, JLV 185 Giolo, SR 261 Davis, J Poster Bac23 Evans, D 61 Gisder, S 27 Davis, J 88 6 Goll, P 282, 307				Poster Fun24	Gardescu, S	281
Damude, H 126 ElSahly, N 317 Gauthier, D 225 Dara, SK 53, 87 Endo, H 46, 212 Gazzola, D 125 Dara, SS 53, 87 Erlandson, MA 17, 65, 68, 77, Geisert, R 251 Dara, SSR 53, 87 161, 181, 182 Gelbic, I 217 da Silva, EYY 200, 201 Escasa, S 182 Genersch, E 10, 27, 59 Dastranj, M 259 Escobar, BF 209, 210 Gerber, R 126 183, 184 Daughenbaugh, K 38 Escriche, B 120, 121, 263 Ghiringhelli, PD 183, 184 David, M Poster Bac23 Estrela, JLV 185 Giol, SR 47, 166, 168 Davidson, EW 1 Evans, D 61 Gisder, S 27 Davis, JW 9 1 Evans, JD 82 Globe, TA 281 Davis, J 8 5 FFFF Goble, TA 281 29, 275, 227, 228 de Bortoli, SA 204, 205, 206 Falcio, R 185	DDD		Elkinton, JS	18	Garrido-Jurado, I	<i>179, 243</i>
Dara, SK 53, 87 Endo, H 46, 212 Gazzola, D 125 Dara, SSR 53, 87 Erlandson, MA 17, 65, 68, 77, Geisert, R 251 Dara, SSR 53, 87 161, 181, 182 Gelbic, I 217 da Silva, EYY 200, 201 Escasa, S 182 Genersch, E 10, 27, 59 Dastranj, M 259 Escobar, BF 209, 210 Gerber, R 126 Dautel, H 38 Escriche, B 120, 121, 263 Ghirringhelli, PD 183, 184 David, M Poster Bac23 Estrela, JLV 185 Giolo, SR 47, 166, 168 Davids, JW Poster Bac23 Evans, JD 82 Glare, TR 282, 307 Davis, JW Poster Bac23 Evans, JD 82 Glare, TR 282, 307 de Bortoli, CP 204, 205, 206 FFFF Goble, TA 281 281 de Bortoli, SA 204, 205, 206 Fabrick, JA 73, 150 Golo, P 275, 227, 228 de Buyn, M-M 21, 22 Fan, Jiangbin 86	Dahiru, E	41	Ellis, CB	90	Gassmann, AJ	238
Dara, SS 53, 87 Erlandson, MA 17, 65, 68, 77, 161, 181, 182 Geisert, R 251 Dara, SSR 53, 87 161, 181, 182 Gelbic, I 217 da Silva, EYY 200, 201 Escasa, S 182 Genersch, E 10, 27, 59 Dastranj, M 259 Escobar, BF 209, 210 Gerber, R 126 Daughenbaugh, K 38 Escriche, B 120, 121, 263 Ghiringhelli, PD 183, 184 Dautel, H 271 Espinel, C 183, 276 Gill, SS 47, 166, 168 David, M Poster Bac23 Estrela, JLV 185 Giolo, SR 261 Davis, IW Poster Bac23 Evans, D 61 Gisder, S 27 Davis, J 88 Evans, JD 82 Glare, TR Goble, TA 281 de Bortoli, CP 204, 205, 206 FFF Goble, TA 281 281 de Bruyn, M-M 21, 22 Fan, Jiangbin 86 Gomen, ACMM Poster Vir20 de Carvalho, VFP 262 Fang, Ving <td>Damude, H</td> <td>126</td> <td>ElSahly, N</td> <td>317</td> <td>Gauthier, D</td> <td>225</td>	Damude, H	126	ElSahly, N	317	Gauthier, D	225
Dara, SSR 53, 87 161, 181, 182 Gelbic, I 217 da Silva, EYY 200, 201 Escasa, S 182 Genersch, E 10, 27, 59 Dastrani, M 259 Escobar, BF 209, 210 Gerber, R 126 Daughenbaugh, K 38 Escriche, B 120, 121, 263 Ghiringhelli, PD 183, 184 Dautel, H 271 Espinel, C 183, 276 Gill, SS 47, 166, 168 David, M Poster Bac23 Estrela, JLV 185 Giolo, SR 261 Davis, IW Poster Bac23 Evans, D 61 Gisder, S 27 Davis, IW Poster Bac23 Evans, D 82 Glare, TR 282, 307 Davis, J 88 FF F Golbe, TA 281 de Bortoli, CP 204, 205, 206 FF F F Goffré, D 57, 227, 228 de Bortoli, SA 204, 205, 3 Fabrick, JA 73, 150 Golo, P 275, 277 de Bruyn, M-M 21, 22 Fan, Jiangbin 86 Gomes, ACMM Poster Valderra	Dara, SK	53, 87	Endo, H	46 , 212	Gazzola, D	125
Dara, SSR 53, 87 161, 181, 182 Gelbic, I 217 da Silva, EYY 200, 201 Escasa, S 182 Genersch, E 10, 27, 59 Dastrani, M 259 Escobar, BF 209, 210 Gerber, R 126 Daughenbaugh, K 38 Escriche, B 120, 121, 263 Ghiringhelli, PD 183, 184 Dautel, H 271 Espinel, C 183, 276 Gill, SS 47, 166, 168 David, M Poster Bac23 Estrela, JLV 185 Giolo, SR 261 Davis, IW Poster Bac23 Evans, D 61 Gisder, S 27 Davis, IW Poster Bac23 Evans, D 82 Glare, TR 282, 307 Davis, J 88 FF F Golbe, TA 281 de Bortoli, CP 204, 205, 206 FF F F Goffré, D 57, 227, 228 de Bortoli, SA 204, 205, 3 Fabrick, JA 73, 150 Golo, P 275, 277 de Bruyn, M-M 21, 22 Fan, Jiangbin 86 Gomes, ACMM Poster Valderra	Dara, SS	53 , 87	Erlandson, MA	17, 65 , 68, 77,	Geisert, R	251
da Silva, EYY 200, 201 Escasa, S 182 Genersch, E 10, 27, 59 Datsranj, M 259 Escobar, BF 209, 210 Gerber, R 126 Daudel, H 38 Escriche, B 120, 121, 263 Ghiringhelli, PD 183, 184 David, M 271 Espinel, C 183, 276 Gill, SS 47, 166, 168 David, M Poster Bac23 Estrela, JLV 185 Giolo, SR 261 Davidson, EW 1 Evans, D 61 Gisder, S 27 Davis, JW Poster Bac23 Evans, JD 82 Glare, TR 282, 307 Davis, J 88 FFF Goffle, D 57, 227, 228 de Bortoli, CP 204, 205, 206 FFF Goffle, D 57, 227, 228 de Bortoli, SA 204, 205, 3 Fabrick, JA 73, 150 Golo, P 275, 277 de Bruyn, M-M 21, 22 Fan, Jiangbin 86 Gomes, ACMM Poster Vir20 de Carvalho, VFP 262 Fang, Yang 94 Gomez, U			·			217
Dastronj, M 259 Escobar, BF 209, 210 Gerber, R 126 Daughenbaugh, K 38 Escriche, B 120, 121, 263 Ghiringhelli, PD 183, 184 Dautel, H 271 Espinel, C 183, 276 Gill, SS 47, 166, 168 David, M Poster Bac23 Estrela, JLV 185 Giolo, SR 261 Davis, IW Poster Bac23 Evans, D 82 Glare, TR 282, 307 Davis, J 88 Goble, TA 281 281 de Bortoli, CP 204, 205, 206 FFFF Goffré, D 57, 227, 228 de Bortoli, SA 204, 205, 7 Fabrick, JA 73, 150 Golo, P 275, 277 de Bortoli, SA 204, 205, 7 Fabrick, JA 73, 150 Gomendoza, KL 98 de Bruyn, M-M 21, 22 Fan, Jiangbin 86 Gomes, ACMM Poster Vir20 de Carvalho, VFP 262 Fang, Longfa 122 Gómez-Valderrama, J 276 de Laurentis, VL 262 Fang, Ying 214, 239, 258			Escasa. S			
Daughenbaugh, K 38 Escriche, B 120, 121, 263 Ghiringhelli, PD 183, 184 Dautel, H 271 Espinel, C 183, 276 Gill, SS 47, 166, 168 David, M Poster Bac23 Estrela, JLV 185 Giolo, SR 261 Davis, IW Poster Bac23 Evans, D 61 Gisder, S 27 Davis, J 88 Goble, TA 282, 307 206, 206 282 Goble, TA 281 de Bortoli, CP 204, 205, 206 FFF Goffré, D 57, 227, 228 66 60fré, D 57, 227, 228 66 60fo, P 275, 277 282 66 60fo, P 275, 277 283 66 Gomendoza, KL 98 66 60fore, P 275, 277 276 62 62 62 62 63 64 63 63 64 64 64 64 64 64 64 64 64 64 64 64 64 64 64 64 64 64 64		•				
Dautel, H 271 Espinel, C 183, 276 Gill, SS 47, 166, 168 David, M Poster Bac23 Estrela, JLV 185 Giolo, SR 261 Davidson, EW 1 Evans, D 61 Gisder, S 27 Davis, JW Poster Bac23 Evans, JD 82 Gole, TR 282, 307 Davis, J 88 FFF Goble, TA 281 de Bortoli, CP 204, 205, 206 FFF Goffré, D 57, 227, 228 de Bortoli, SA 204, 205, , Fabrick, JA 73, 150 Golo, P 275, 277 de Bruyn, M-M 21, 22 Fan, Jiangbin 86 Gomes, ACMM Poster Vir20 de Carvalho, VFP 262 Fang, Longfa 122 Gómez-Valderrama, J 276 de Castro, MT 216, 233, 249, Fang, Yang 94 Gomez, I 43, 183 de Laurentis, VL 262 Fannon, J 61 Gomez, Lebolla, J 121, 123, 208 De Ley, IT 33 Fanzolin, M 185 González-Mas, N 153	•		•		•	
David, M Poster Bac23 Estrela, JLV 185 Giolo, SR 261 Davidson, EW 1 Evans, D 61 Gisder, S 27 Davis, IW Poster Bac23 Evans, JD 82 Glare, TR 282, 307 Davis, J 88 FFF Goble, TA 281 de Bortoli, CP 204, 205, 206 FFF Goffré, D 57, 227, 228 de Bortoli, SA 204, 205, 7 Fabrick, JA 73, 150 Golo, P 275, 277 de Bruyn, M-M 21, 22 Fan, Jiangbin 86 Gomendoza, KL 98 de Carvalho, VFP 262 Fang, Longfa 122 Gómez, Valderrama, J 276 de Castro, MT 216, 233, 249, 249, 249 Fang, Yang 94 Gomez, I 207 250 Fang, Ying 214, 239, 258 Gómez, J 43, 183 de Laurentis, VL 262 Fannon, J 61 Gomis-Cebolla, J 121, 123, 208 De Ley, IT 33 Fanzolin, M 185 González-Mas, N 153	• •				_	
Davidson, EW 1 Evans, D 61 Gisder, S 27 Davis, IW Poster Bac23 Evans, JD 82 Glare, TR 282, 307 Davis, J 88 Goble, TA 281 de Bortoli, CP 204, 205, 206 FFF Goffré, D 57, 227, 228 de Bortoli, SA 204, 205, , Fabrick, JA 73, 150 Golo, P 275, 277 de Bruyn, M-M 21, 22 Fan, Jiangbin 86 Gomendoza, KL 98 de Carvalho, VFP 262 Fang, Longfa 122 Gómez-Valderrama, J 276 de Castro, MT 216, 233, 249, Fang, Yang 94 Gomez, I 207 de Laurentis, VL 262 Fang, Ying 214, 239, 258 Gómez, J 43, 183 de Laurentis, VL 262 Fannon, J 61 Gomis-Cebolla, J 121, 123, 208 De Ley, IT 33 Fanzolin, M 185 González-Mas, N 153 DeSantis, TZ Poster Bac23 Federici, BA 6, 11, 75, 142, Gonzalez, A 232 <td>•</td> <td></td> <td>·</td> <td>•</td> <td></td> <td></td>	•		·	•		
Davis, IW Poster Bac23 Evans, JD 82 Glare, TR 282, 307 Davis, J 88	•		-		·	
Davis, J 88 Goble, TA 281 de Bortoli, CP 204, 205, 206 FFF Goffré, D 57, 227, 228 de Bortoli, SA 204, 205, , Fabrick, JA 73, 150 Golo, P 275, 277 de Bruyn, M-M 21, 22 Fan, Jiangbin 86 Gomendoza, KL 98 de Carvalho, VFP 262 Fang, Longfa 122 Gómez-Valderrama, J 276 de Castro, MT 216, 233, 249, Fang, Yang 94 Gomez, I 207 de Laurentis, VL 262 Fang, Ying 214, 239, 258 Gómez, J 43, 183 de Laurentis, VL 262 Fannon, J 61 Gomis-Cebolla, J 121, 123, 208 De Ley, IT 33 Fanzolin, M 185 González-Mas, N 153 DeSantis, TZ Poster Bac23 Federici, BA 6, 11, 75, 142, Gonzalez, A Gonzalez, Villareal, SE 139 de Silva, CSR 244 143, 164 Gonzalez, A 232	•		•			
de Bortoli, CP 204, 205, 206 FFF Goffré, D 57, 227, 228 de Bortoli, SA 204, 205, , Fabrick, JA 73, 150 Golo, P 275, 277 de Bruyn, M-M 206, 262 Falcão, R 185 Gomendoza, KL 98 de Carvalho, VFP 262 Fan, Jiangbin 86 Gomes, ACMM Poster Vir20 de Castro, MT 216, 233, 249, Fang, Longfa 122 Gómez-Valderrama, J 276 de Laurentis, VL 250 Fang, Yang 94 Gomez, I 207 De Ley, IT 33 Fannon, J 61 Gomis-Cebolla, J 121, 123, 208 De Santis, TZ Poster Bac23 Federici, BA 6, 11, 75, 142, Gonzalez-Willareal, SE 139 de Silva, CSR 244 143, 164 Gonzalez, A 232	•		Evans, 35	02		
de Bortoli, SA 204, 205, , Fabrick, JA 73, 150 Golo, P 275, 277 206, 262 Falcão, R 185 Gomendoza, KL 98 de Bruyn, M-M 21, 22 Fan, Jiangbin 86 Gomes, ACMM Poster Vir20 de Carvalho, VFP 262 Fang, Longfa 122 Gómez-Valderrama, J 276 de Castro, MT 216, 233, 249, Fang, Yang 94 Gomez, I 207 de Laurentis, VL 262 Fang, Ying 214, 239, 258 Gómez, J 43, 183 de Laurentis, VL 262 Fannon, J 61 Gomis-Cebolla, J 121, 123, 208 De Ley, IT 33 Fanzolin, M 185 González-Mas, N 153 DeSantis, TZ Poster Bac23 Federici, BA 6, 11, 75, 142, Gonzalez-Villareal, SE 139 de Silva, CSR 244 143, 164 Gonzalez, A 232	•		FFF			
206, 262 Falcão, R 185 Gomendoza, KL 98 de Bruyn, M-M 21, 22 Fan, Jiangbin 86 Gomes, ACMM Poster Vir20 de Carvalho, VFP 262 Fang, Longfa 122 Gómez-Valderrama, J 276 de Castro, MT 216, 233, 249, Fang, Yang 94 Gomez, I 207 250 Fang, Ying 214, 239, 258 Gómez, J 43, 183 de Laurentis, VL 262 Fannon, J 61 Gomis-Cebolla, J 121, 123, 208 De Ley, IT 33 Fanzolin, M 185 González-Mas, N 153 DeSantis, TZ Poster Bac23 Federici, BA 6, 11, 75, 142, Gonzalez-Villareal, SE 139 de Silva, CSR 244 143, 164 Gonzalez, A 232	•			73 150	•	
de Bruyn, M-M 21, 22 Fan, Jiangbin 86 Gomes, ACMM Poster Vir20 de Carvalho, VFP 262 Fang, Longfa 122 Gómez-Valderrama, J 276 de Castro, MT 216, 233, 249, Fang, Yang 94 Gomez, I 207 250 Fang, Ying 214, 239, 258 Gómez, J 43, 183 de Laurentis, VL 262 Fannon, J 61 Gomis-Cebolla, J 121, 123, 208 De Ley, IT 33 Fanzolin, M 185 González-Mas, N 153 DeSantis, TZ Poster Bac23 Federici, BA 6, 11, 75, 142, Gonzalez-Villareal, SE 139 de Silva, CSR 244 143, 164 Gonzalez, A 232	ue Borton, SA		· ·	•		
de Carvalho, VFP 262 Fang, Longfa 122 Gómez-Valderrama, J 276 de Castro, MT 216, 233, 249, Fang, Yang 94 Gomez, I 207 250 Fang, Ying 214, 239, 258 Gómez, J 43, 183 de Laurentis, VL 262 Fannon, J 61 Gomis-Cebolla, J 121, 123, 208 De Ley, IT 33 Fanzolin, M 185 González-Mas, N 153 DeSantis, TZ Poster Bac23 Federici, BA 6, 11, 75, 142, Gonzalez-Villareal, SE 139 de Silva, CSR 244 143, 164 Gonzalez, A 232	do Bruyo M M		-		•	
de Castro, MT 216, 233, 249, Fang, Yang 94 Gomez, I 207 250 Fang, Ying 214, 239, 258 Gómez, J 43, 183 de Laurentis, VL 262 Fannon, J 61 Gomis-Cebolla, J 121, 123, 208 De Ley, IT 33 Fanzolin, M 185 González-Mas, N 153 DeSantis, TZ Poster Bac23 Federici, BA 6, 11, 75, 142, Gonzalez-Villareal, SE 139 de Silva, CSR 244 143, 164 Gonzalez, A 232	·		• •			
Ley, IT 33 Fancy In Mark 61 Gomez, J 43, 183 De Ley, IT 33 Fanzolin, M 185 González-Mas, N 153 DeSantis, TZ Poster Bac23 Federici, BA 6, 11, 75, 142, Gonzalez-Villareal, SE 139 de Silva, CSR 244 143, 164 Gonzalez, A 232					·	
de Laurentis, VL 262 Fannon, J 61 Gomis-Cebolla, J 121, 123, 208 De Ley, IT 33 Fanzolin, M 185 González-Mas, N 153 DeSantis, TZ Poster Bac23 Federici, BA 6, 11, 75, 142, Gonzalez-Villareal, SE 139 de Silva, CSR 244 143, 164 Gonzalez, A 232	ae Castro, MT				·	
De Ley, IT 33 Fanzolin, M 185 González-Mas, N 153 De Santis, TZ Poster Bac23 Federici, BA 6, 11, 75, 142, Gonzalez-Villareal, SE 139 de Silva, CSR 244 143, 164 Gonzalez, A 232					•	
DeSantis, TZ Poster Bac23 Federici, BA 6 , 11, 75, 142, Gonzalez-Villareal, SE 139 de Silva, CSR 244 143, 164 Gonzalez, A 232	· · · · · · · · · · · · · · · · · · ·				·	
de Silva, CSR 244 143, 164 Gonzalez, A 232	• • • • • • • • • • • • • • • • • • • •		•		·	
			Federici, BA			
Feist, SW 115 Goodman, C Oral Mcn#2	de Silva, CSR	244				
			Feist, SW	115	Goodman, C	Oral Mcn#2

Görg, L	50, 270	Horchler, L	27	Johnston, B	Oral-Msp #1
Göttmann, J	293	Hosseininaveh, V	263	Jones, S	241
Greco, F	308	Hotopp, JCD	102	Joseph, L	41
Green, M	115	Hou, Chunsheng	314	Juneja, P	289
Gross, J	50, 270	Hou, Jingtong	48, 49	Jurat-Fuentes, JL	42 , 74, 137
Grzywacz, D	260	Hou, Zhenglin	48, 49		
Gueli Alletti, G	85	Hsiang, Chus-Jou	289	KKI	
Guevara, J	184	Hu, Huajun	236	Kabaluk, T	266
Guide, BA	255	Hu, Jia	163	Kadono-Okuda, K	144
Guilhabert, M	310	Hu, Jian-Sheng	165	Kagimu, N	28
Guimarães, JA	245, 246	Hu, Sha Sha	112	Kahn, TW	220
Guan, Xiong	217	Hu, Yan	125	Kain, W	208
Guo, Shuyuan	78	Hu, Yaqiang	236	Kakani, K	48
Guo, Ya	67	Hu, Zhihong	7, 70 ,	Kamminga, K	222
Gwak, WonSeok	194, 224, 273,	ria, ziiiiong	Oral Vir#1,	Kanlidere, Z	149
GWak, Wonseek	274, Poster		Oral Vir#5	Kannost, MR	237
		Huang liamian	187	Kariithi, HM	117
	Msp06,	Huang, Jiamiao		•	
6	Poster Vir21	Huang, Wei-Fone	58	Kassa, A	126
Gwynn, R	90, 309	Huang, Yu-Feng	19	Kato, F	315
		Huang, Ying	Poster Bac24	Kaya, HK	9
ннн		Huang, Zhihong	96 , 97	Keddie, A	181
Ha, Pan Jung	180	Huarte-Bonnet, C	106, 176 , 242	Keith, L	54
Haas, JA	Poster Bac23	Hughes, GE	22	Kelly, R	151
Haigh, AM	257	Hughes, L	188	Keyhani, NO	81 , 111
Haile, F	126	Humber, RA	8 , 229 , 230,	Keyser, CA	88
Hajek, AE	26 , 281 , 300		231, 232	Khamis, F	248
Hall, AAG	172	Humbert, P	92, 270, 271,	Khan, IA	268
Hall, D	210		272	Khorramnejad, A	263
Hall, R	283			Kim, Dong Jun	194, 224, 273,
Hampton, J	282	111		, 6	274, Poster
Han, Ji hee	292	Ibarra, JE	139		Msp06,
Han, Yue	159 , 160	lmai, T	109		Poster Vir21
Haramboure, M	133	Ince, IA	149 , 164	Kim, InHui	224, Poster
Harding, K	49	Inoue, MN	100, 164, 197,	Killi, Illiai	Msp06
Harrison, RL	17, 20 , Oral	mode, will	199	Kim, Jae Su	19, 107, 177,
riuriison, KL	Mcn#2	Irons, S	188	Kiiii, Jue Ju	180 , 193, 239,
Heelen I	137	•			240,
Hasler, J		Isawa, H	315		•
Hassanlouei, RT	51	Ishii, H	164		Oral Fun#3,
Hata, T	109	Ito, K	144	12.	Poster Fun25
Hawes, C	188	Itsathitphaisarn, O	24	Kim, Jeong Jun	292
He, Jianguo	187	Iwai, S	Poster Bac23	Kim, Jong Cheol	19, 107, 177,
He, Kanglai	79 , 80				180, 240,
Heckel, DG	76 , 77	111			Poster Fun25
Heckert, M	126	Jackson, MA	281	Kim, Jong Hoon	214, 239, 258
Hegedus, D	77, 162	Jackson, TA	14 , 241, 282,	Kim, Sihyeon	107, 177 , 180,
Henderson, DE	277		283, 303		240,
Henry, JE	130	Jacob, A	316		Oral Fun#3,
Hepat, RP	17	Jacobs-Schoenwandt, D	156		Poster Fun25
Hermann, K	272	Jacobson, R	90	Kim, Tae Hoon	180
Hernandez-Martinez, P	121	Jafir, M	268, Oral	Kim, Woo Jin	18, 214, 239,
Hernández-Rodríguez, CS	213		Mcn#2		258
Herniou, EA	16, 182	Jain, S	Poster Bac23	Kim, Yonggyun	31, 215, 267,
Hesketh, H	10	Jaroenlak, P	24		Oral Mcn#2
Hibbard, B	251	Jaronski, ST	5 , 238, 264,	Kimber, MJ	34
Hice, R	143	•	279	Kimemia, JW	248
Hietala, L	137	Je, Yeon Ho	214, 239, 258	King, LA	95, 69 , 188
Hiltpold, I	32 , 301	Jehle, JA	25, 71 , 85 , 86,	Kinkel, LL	299
Hirano, T	199	Jeme, Jr	100,	Klikel, LL Kleeberg, I	269
		lansan AP		=	25
Hoch, G	302	Jensen, AB	50, 64	Kleespies, RG	
Hodgson, EW	238	Jensen, AH	50	Klinger, E	60 365
Hodgson, J	67	Jiang, Jian	Poster Bac25	Koike, M	265
Holdbrook, R	138	Jimenez, N	126	Krell, PJ	3 , 94, 189
Hong, Mingsheng	291	Jiwaji, M	21, 22	Krell, V	156
Hoover, K	196	Johnson, AJ	Poster–Bac23	Kristmundsson, A	Oral DBI#2
Hopper, JV	223	Johnson, KN	39	Kritunyalucksana, K	24

Large, CE 133 Lun, A 126 Monnerat, RG 200, 201, 20 Larsen, J 234 Luo, Kol-Jun 165 216, 218, 25 Lassner, M 49 Luttrell, R 74, 278 249, 250 Lau, S 48 Luz, C 106, 229, 230, Monroy, DM 276 Lae, Bo Ram 214, 239, 258 Wontalva, C 229, 231, 22 Lee, BiHoon 194, 224, 273, Montalvão, SCL 233, 249, 25 Lee, JiHoon 194, 224, 273, More, ID Ma, Jiln 194, 224, 273, Morore, ID 131 Lee, Mi Rong 107, 177, 180, Ma, Zhi 274, Poster Moron, N 298 298 Lee, Mi Rong 107, 177, 180, Macedo, C 207 Morgado, FdaS 93 Poster Fun25 Macchietto, M 30 Morrow, JL 172 Lee, Sea Jin 19, 107, 177, Mackenstedt, U 271 Mosley, G 253 Lee, Seok Ju 180, 193, 240, Madoodi, A 68 Mowery, J 20 Lee, Seok-Hee 214, 239, 258 Malan, AP 28 Moullier, P 316	Kroner, M	151	Lottsfeldt, NS	114	Meki, IK	117
Kuman, P	Krska. D	59	Lovett. B	152	Melo. FL	62. 190
Kunimi, Y 100, 197, 199	•				-	•
Kusia, E 248	=				,	
Lipeng	•		•		·	
Lit	·		, ,		•	
Lucrospec 15	куеї-Роки, G	225	_	•		
Labrouse, C			•		•	
Lodumer, Ε 308 Lukovsky, A 157 Mireva, M 101 Lang, C, Ε 133 Lule-Chaver, AN 139 Mohamed, AI Poster FunZ Larsen, J 234 Lun, Ro-Jun 165 152, 18, 25 Lassner, M 49 Lutr, CR 74, 278 Monroy, DM 276 Larate, JN 203 Luz, C 106, 229, 231, 224 Monrow, DM 276 Lee, HHoon 194, 224, 273, Poster Mo, Jili 194, 224, 273, More, Jili Mornor, Dill, Mire 233, 249, 22 Lee, MI Rong 107, 177, 180, Poster Mo, Jili 194, 224, 273, More, Jili More, Jili More, Jili More, Jili 307 274, Poster Moran, Dile, ME 307 238, 249, 22 233, 249, 22 220 More, Jili 194, 224, 273, More, Jili More, Jili More, Jili 194, 224, 273, More, Jili More, Jili 307 Morra, Jili 307 233, 249, 22 233, 249, 22 230 More, Jili 307 Morra, Jili 100 200 224, 273, More, Jili More, Jili 100			Lucero, NF	228	Mikaia, N	
Lai, Qinying	Labrousse, C	16	Luevano-Borroel, J	139	Mirza, Z	125
Longe, € Larsen, J 234 Lum, A 126 Monnerat, RS 200, 201, €1, 218, 221 Lassen, J 234 Luto, Kol-lun 165 266, 218, 221 Lasus, S 48 Lut, C 106, 229, 230, Monroy, DM 249, 250 Larate, JN 203 231, 244 Montoke, C 229, 231, 224 Lee, Bo Ram 194, 224, 273, Monro, D 194, 224, 273, Monro, D Montoke, C 233, 249, 22 Lee, HHoon 194, 224, 273, Monro, D 194, 224, 273, Monro, D 133, 249, 22 Lee, HR Rong 107, 177, 180, Moro, D Vir21 Moreau, Y 16 Lee, MR Rong 107, 177, 180, Moro, D Vir21 Morogan, C fdds 39 Lee, Sang Yeop 292 Machietto, M 30 Morogan, C fdds 39 Lee, Sang Yeop 292 Mach, RI 118 Mortazavi, A 30 Lee, Seok Ju 180, 193, 240, Mordoo, J 129 Moullier, P 316 Lee, Seok Ju 180, Poster Millouder, D 292 Machonerat, Maghodia, A 68 Mowere, D </td <td>Ladurner, E</td> <td>308</td> <td>Lukowsky, A</td> <td>157</td> <td>Mitreva, M</td> <td>101</td>	Ladurner, E	308	Lukowsky, A	157	Mitreva, M	101
Longe, CE 133 Lum, A 126 Monnerat, RG 200, 201, 25, 18, 25 215, 218, 25 215, 218, 25 215, 218, 25 215, 218, 25 215, 218, 25 215, 218, 25 215, 218, 25 220, 201, 224 Monroy, DM 276 249, 250 Lazarte, IN 203 Lee, Bo Ram 212, 239, 258 MIM M Monstrake, J 220 Monthoko, C 229, 231, 224 Monthoko, C 229, 231, 224 Monthoko, C 233, 249, 22 240, Monthoko, SCL 240, Monthoko, SCL	Lai, Qinying	195	Lule-Chavez, AN	139	Mohamed, Al	Poster Fun23
Jassen	Lanae. CE	133	· · · · · · · · · · · · · · · · · · ·	126	Monnerat. RG	200, 201, 207,
Jassner, M			·		,	
Lau, S 48 Luz, C 106, 229, 230, and monroy, DM Monroy, DM 276 Lee, BR Bam 214, 239, 258 231, 244 Monrative, C 229, 231, 244 Lee, BHoon 194, 224, 273, and monroy, DM Monrollyo, CC 229, 231, 234, 222 John Spins MSP06, and	•					
Lazarte, IN Lee, Bo Ram	•		•	,		
Lee, Bo Ram 194, 224, 273, 274 274, Poster Abgolo, Poster Vir.21 Lee, Mi Rong 107, 177, 180, Poster Fun25 Mach, R. R. R. R. R. R. R. R. R. R. R. R. R.			Luz, C		**	
Lee, JiHoon 194, 224, 273, 273, 249, 22 M M M M M M M M M Sp06, 274, Poster Moran-Diez, ME 307 307 Lee, Mi Rong 107, 177, 180, 240, Machine March Moran Diez, ME 307, 177, 180, 240, Machine March Moran Machine March Machine March Machine M	Lazarte, JN	203		231, 244	Monserrate, J	220
May May	Lee, Bo Ram	214, 239, 258			Montalva, C	229, 231 , 232
Lee, Mi Rong Msp06, Poster Wi 21 274, Poster Msp06, Poster Moran-Diez, Msp06, Poster Moran-Diez, Msp06, Poster Moran, N 298 Lee, Mi Rong 107, 177, 180, 240, Ms, 2hi 58 Morgado, FdaS 93 Oral Funif3, Macedo, C 207 Morgan, T 208 Lee, Sang Yeop 292 Mach, Rt 118 Mortzawi, A 30 Lee, Sang Yeop 19, 107, 177, Mackenstedt, U 271 Mosley, G 253 Lee, Sang Yeop 292 Machdow, J 129 Moullier, P 316 Lee, Seek Jin 180, 193, 240, Maddow, J 129 Moullier, P 316 Lee, Seek Ju 180, Poster Malgocka, J 68 Moweny, J 20 Lee, Seek Hee 118, Opester Malagocka, J 64 Mushtaq, R 42 Lee, Sang Yeop 292 Manktelow, CI 72, 202 Lee, Sang Yeop 292 Marcinon, AF 277 Mwembela, N 290 Lee, Sang Yeop 292 Marcinon, AF 277 Nagoshi, R 137 <	Lee, JiHoon	194 , 224 <i>, 273,</i>	MMM		Montalvão, SCL	233, 249 , 250
Lee, Mi Rong Msp06, Poster Wi 21 274, Poster Msp06, Poster Moran-Diez, Msp06, Poster Moran-Diez, Msp06, Poster Moran, N 298 Lee, Mi Rong 107, 177, 180, 240, Ms, 2hi 58 Morgado, FdaS 93 Oral Funif3, Macedo, C 207 Morgan, T 208 Lee, Sang Yeop 292 Mach, Rt 118 Mortzawi, A 30 Lee, Sang Yeop 19, 107, 177, Mackenstedt, U 271 Mosley, G 253 Lee, Sang Yeop 292 Machdow, J 129 Moullier, P 316 Lee, Seek Jin 180, 193, 240, Maddow, J 129 Moullier, P 316 Lee, Seek Ju 180, Poster Malgocka, J 68 Moweny, J 20 Lee, Seek Hee 118, Opester Malagocka, J 64 Mushtaq, R 42 Lee, Sang Yeop 292 Manktelow, CI 72, 202 Lee, Sang Yeop 292 Marcinon, AF 277 Mwembela, N 290 Lee, Sang Yeop 292 Marcinon, AF 277 Nagoshi, R 137 <		274, Poster	Ma, Jiln	194, 224 , 273,	Moore, JD	113
Lee, Mi Rong Poster Vil21 Mespol6, Poster Paraly Moreau, Y 128 Lee, Mi Rong 107, 177, 180, Ma, Zhi 58 Morgan, T 208 Jonal Funil S, Doster Fun25 Macchietto, M 30 Morrow, J. 172 Lee, Sang Yeop 292 Mach, RL 118 Mortazavi, A 30 Lee, Seln 19, 107, 177, Mackenstedt, U 271 Mostley, G 253 180, 193, 240, Maddox, J 129 Moullier, P 316 Lee, Seok Ju 180, 193, 240, Maghodia, A 68 Mowen, J 20 Lee, Seok Ju 180, Poster Malagacka, J 68 Mowen, J 20 Lee, Seok Hee 19, 239, 28 Malan, AP 28 Muskat, L 270 Lee, Seok Hee 212, 239, 258 Malan, AP 28 Muskat, L 270 Lee, Seok Hee 213 Marcin Marcin, M 268 Muskat, L 270 Lee, Seok Hee 213 Marcin M 277 Nag, M 191 <					·	307
Lee, MI Rong 107, 177, 180, Vir21 Moreau, Y 16 240, Ma, 2hi 58 Morgad, FdaS 93 Lee, Sang Yeop 292 Mach, RL 118 Morraw, IL 172 Lee, Se Jin 19, 107, 177, Mackenstedt, U 271 Mosley, G 253 Lee, Se Jin 19, 107, 177, Mackenstedt, U 271 Mosley, G 253 Lee, Seok Jiu 180, 193, 240, Maddow, J 129 Molliller, P 316 Lee, Seok Jiu 180, Poster Majumder, D Poster Minals Malan, AP 42 Lee, Seok-Hee 180, Poster Majumder, D Poster Minals Muskata, L 270 Lee, Sang Yeop 292 Mankelow, CI 72, 202 12 Marciano, A 28 Mwambela, N 290 Lee, Seok-Hee 136 Marciano, A 277 Nager 183 137 18 137 18 18 18 18 18 18 18 18 18 18 <td< td=""><td></td><td>• •</td><td></td><td>•</td><td>•</td><td></td></td<>		• •		•	•	
240	Lee Mi Pona			• •		
Poster Fun25 Macchel to, M	Lee, IVII NOIIG		NA - 71 '		•	
Lee, Sang Yeop 292 Machietto, M Lee, Sang Yeop 292 Mach, RL 118 Mortazavi, A 30 Lee, Se Ilin 19, 107, 177, Mackenstedt, U 271 Mosley, G 253 180, 193, 240, Maddox, J 129 Moullier, P 316 Oral Funal's, Maghodia, A 68 Mowery, J 20 Oral Funal's, Maghodia, A 68 Mowery, J 20 Dester Fun25 Mahadeva swamy, HM 140 Mushimiyimana, I 316 Lee, Seok Ju 180, Poster Majiumder, D Poster Mcn26 Mushimiyimana, I 316 Lee, Seok Ju 180, Poster Majiumder, D Poster Mcn26 Mushimiyimana, I 316 Lee, Seok Hee 214, 239, 258 Malan, AP 28 Mwambela, N 290 Lee, Sang Yeop 292 Manktelow, C 72, 202 Lei, Chengfeng 163 Manzoor, M 268 Lemaitre, B 136 Marciano, AF 277 Nagos, Leenanter, S 220 Marcomini, MC 255 Nagoshi, R 137 Lenzen, S 220 Marcomini, MC 255 Nagoshi, R 137 Lenzen, S 311 Marros, MM 30 Najiva-Shin 19, 193 Letherer, T 199 Marion-Poll, F 109 Nai, Yu-Shin 19, 193 Letwin, S 311 Marros, MM 30 Najiva-Shin 19, 193 Letwin, S 311 Marros, MM 30 Najiva-Shin 19, 193 Letwin, S 311 Marros, MM 30 Najiva-Shin 19, 193, 304 Lewis, F 301 Marros, MM 30 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 30 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 30 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 30 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 30 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 30 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 30 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 30 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 301 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 301 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 301 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 301 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 301 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 301 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 301 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 301 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 301 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 301 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 301 Najiva-Shin 19, 193, 304 Lewis, F 301 Marrios, MM 301 Najiva-Sh		•	•		• .	
Lee, Sang Yeop 292 Mach, R. 118 Mortazavi, A. 30 Lee, Se Jin 19, 107, 177. Mackenstedt, U 271 Mosley, G 253 Lee, Se Jin 19, 193, 240, Maddox, J 129 Moullier, P 316 Der, Seok Ju 180, Poster Majhodia, A 68 Mover, J 316 Lee, Seok Ju 180, Poster Majdoka, J 68 Mushtal, R 42 Lee, Seok Hee 180, Poster Majagocka, J 64 Muskat, L 270 Lee, Sang Yeop 292 Manktelow, CI 72, 202 Meritan, A 28 Mwambela, N 290 Lee, Sang Yeop 292 Manktelow, CI 72, 202 N. N. Lee, Sang Yeop N. N. N. N. Lee, Sang Yeop N. N.		•	•		=	
Lee, Se Jin 19, 107, 177, 180, 180, 193, 240, 180, 193, 240, 180, 193, 240, 180, 193, 240, 180, 193, 240, 180, 193, 240, 180, 193, 240, 180, 193, 240, 180, 193, 240, 180, 193, 240, 180, 193, 240, 180, 193, 240, 180, 193, 240, 180, 193, 240, 180, 193, 240, 180, 193, 240, 180, 193, 240, 180, 180, 180, 180, 180, 180, 180, 18			Macchietto, M	30	·	
180, 193, 240, Maddow, J 129 Moullier, P 316 Poster Fun25 Malpodia, A 68 Mowery, J 20 20 20 20 20 20 20	Lee, Sang Yeop	292	Mach, RL	118	Mortazavi, A	30
Coral Funitā, Maghodia, A 68 Mowery, J 20 Lee, Seok Ju 180, Poster Madadeva swamy, HM 140 Mushiniyimana, I 316 Lee, Seok Ju 180, Poster Majumder, D Poster Mcn26 Mushtaq, R 42 Lee, Seok-Hee 214, 239, 258 Malan, AP 28 Mwambela, N 290 Lee, Seng Yeop 292 Mantelow, CI 72, 202 Let, Chengferg 163 Marcano, AF 277 Number N 290 Lemaitre, B 136 Marcano, AF 277 Nagoshi, R 137 180 187 180	Lee, Se Jin	19, 107, 177,	Mackenstedt, U	271	Mosley, G	253
Lee, Seok Ju Poster Fun25 Mahadeva swamy, HM 140 Mushitaq, R 3 16 Lee, Seok Ju 19,0 Poster Majumder, D Poster Mcn26 Mushtaq, R 42 Lee, Seok Hee 214, 239, 258 Malan, AP 28 Mammbela, N 290 Lee, Sang Yeop 292 Manzoor, M 268 Total Control N N N Lemaitre, B 136 Marcomini, MC 255 Nagoshi, R 137 Lemand, K 208 Marfetán, A 57 Nagy, E 189 Letherer, T 119 Marion-Poll, F 109 Nai, Yu-Shin 191 Levin, S 301 Marrosh, MM 30 Nojer-Rincon, MB 234 Lewis, F 302 Marshall, S 241 Nakayinga, R 21 Li-Leger, E 303 Marshall, S 241 Nakayinga, R 21 Li-Leger, E 314 Martine, EM 32 Nakayinga, R 21 Li, Beibei 314 Martine, EM 228 Nakayinga, R		180, 193, 240,	Maddox, J	129	Moullier, P	316
Lee, Seok Ju Poster Fun25 Mahadeva swamy, HM 140 Mushitan, R 3 16 Lee, Seok Ju 19,0 poster Majumder, D Poster Mcn26 Mushtan, R 42 Lee, Seok-Hee 214, 239, 258 Malan, AP 28 Mwambela, N 290 Lee, Sang Yeop 292 Manzoor, M 268 Total Control N N Lematre, B 136 Marcomin, MC 255 Nagoshi, R 137 Lenard, K 208 Marfedin, A 57 Nagy, E 189 Letherer, T 119 Marion-Poll, F 109 Nai, Yu-Shin 191 Levin, S 301 Marrosh, MM 30 Nojer-Rincon, MB 234 Lewin, S 301 Marshall, S 241 Nakai, M 100, 164, 19 Li-Leger, E 302 Marshall, S 241 Nakai, M 100, 164, 19 Li-Leger, E 304 Martino, EM 38 Nakayinga, R 21 Li, Beibei 314 Martino, EM 38 Nayai, SA <td></td> <td></td> <td>·</td> <td>68</td> <td>·</td> <td>20</td>			·	68	·	20
Lee, Seok Ju 180, Poster Funds Majumder, D Malagocka, J Poster Mcn26 Mushta, R 42 Lee, Seok-Hee 214, 239, 258 Malagocka, J 64 Muskat, L 270 Lee, Seag Yeop 292 Mankelow, CI 72, 202 Lee, Chengfeng 163 Manzoor, M 268 Lemaitre, B 136 Marcomini, MC 255 Nagoshi, R 13 Lemand, K 208 Marfetan, A 57 Nagy, E 189 Lebenard, K 208 Marfetan, A 57 Nagy, E 189 Lebenter, T 119 Marion-Poll, F 109 Nai, Yu-Shin 19, 193 Lewin, S 311 Marros, MM 30 Naigy, E 189 Lewis, E 301 Marshman, BC 111 Noka, M 100, 164, 16 Li-leger, E 266 Marshman, BC 113 Nakayinga, R 21 Li-leger, E 266 Marshman, BC 113 Nava, N 199, 304 Li-ger, E 266 <td< td=""><td></td><td>•</td><td>•</td><td></td><td>• •</td><td></td></td<>		•	•		• •	
Fun25	Loo Sook lu		•		•	
Lee, Seok-Hee 214, 239, 258 Malan, AP 28 Mwambela, N 290 Lee, Sang Yeop 292 Mankelow, CI 72, 202 Lei, Chengfeng 163 Manzoor, M 268 Lemzen, S 220 Marcomini, MC 255 Nagshi, R 137 Leonard, K 208 Marfetán, A 57 Nagy, E 189 Lebrer, T 119 Marion-Poll, F 109 Nai, Yu-Shin 19, 193 Lewis, F 301 Marros, MM 30 Najer-Rincon, MB 234 Li-Leegr, E 301 Marshall, S 241 Makal, M 100, 164, 19 Li, Belbei 314 Martine, LM 228 Nakayinga, R 21 Li, Fel 314 Martine, EM 98 Naqvi, SA 268 Li, Hanchen 125 Martine, E 207 Narva, K 119, 137, 21 Li, Alingham 155 Martine, E 207 Navia, D 252, 226 Li, Hanchen 125 Martine, E	Lee, Seok Ju		• •		•	
Lee, Sang Yeop 292 Manktelow, CJ 72, 202 Lei, Chengfeng 153 Manzoor, M 268 Lemaitre, B 136 Marciano, AF 277 NR SN NN Lemzen, S 220 Marcomini, MC 255 Nagoshi, R 137 Lebnerg, T 119 Marion-Poll, F 109 Nai, Yu-Shin 19, 193 Lewin, S 311 Maros, MM 30 Najera-Rincon, MB 234 Lewis, E 301 Marshall, S 241 Nakoi, M 100, 164, 16 Li, Beibei 314 Martinez, LA 228 Nakayinga, R 21 Li, Fel 314 Martinez, LA 228 Nayavi, SA 268 Li, Hanchen 125 Martins, E 207 Narva, K 119, 137, 21 Li, Ming 165 Masson, F 136 Navia, D 283 Li, Zhaofei 67 Mathis, J 45 Nelson, M 52, 226 Lia, Deman, B 158 Mathew, LG 34 Neves,			• .			
Lei, Chengfeng 163 Marzoor, M 268 Lemaitre, B 136 Marciano, AF 277 NN N Lenzen, S 220 Marcomini, MC 255 Nagoshi, R 137 Leonard, K 208 Marfetán, A 57 Nagy, E 189 Levin, S 119 Marion-Poll, F 109 No., Yu-Shin 19, 193 Levin, S 311 Maros, MM 30 Najera-Rincon, MB 234 Lewis, E 301 Marshall, S 241 Nakai, M 100, 164, 15 Li, Belbei 314 Marshall, S 241 Nakai, M 100, 164, 15 Li, Belbei 314 Martine, LM 228 Nakayinga, R 21 Li, Hanchen 125 Martine, LM 207 Narva, K 119, 137, 21 Li, King 165 Masson, F 136 Navia, D 283 Li, Janchun 73 Mathew, LG 73 Ndakidemi, PA 52, 226 Li, Zhofei 170 Matisoff, M <td>•</td> <td></td> <td></td> <td></td> <td>Mwambela, N</td> <td>290</td>	•				Mwambela, N	290
Lemaitre, B 136 Marciano, AF 277 N N N Lenzen, S 220 Marcomini, MC 255 Nagoshi, R 137 Leonard, K 208 Marfetán, A 57 Nagy, E 189 Leonard, K 208 Marfetán, A 57 Nagy, E 189 Levin, S 311 Marion-Poll, F 109 Nai, Yu-Shin 19, 193 Levin, S 311 Marros, MM 30 Najera-Rincon, MB 234 Lewis, E 301 Marshall, S 241 Nakai, M 100, 164, 15 Li-Leger, E 266 Marshall, S 241 Nakai, M 100, 164, 15 Li, Fei 131 Martinez, LA 228 Nakayinga, R 21 Li, Fei 134 Martinez, LA 228 Nakayinga, R 21 Li, Fei 131 Martinez, LA 228 Nakayinga, R 21 Li, Jene 145 Martinez, LA 228 Nakayinga, R 21 Li, Fei 154 </td <td>Lee, Sang Yeop</td> <td>292</td> <td>Manktelow, CJ</td> <td>72, 202</td> <td></td> <td></td>	Lee, Sang Yeop	292	Manktelow, CJ	72, 202		
Lenzen, S 220 Marcomini, MC 255 Nagoshi, R 137 Leonard, K 208 Marfetán, A 57 Nagy, E 189 Letherer, T 119 Marion-Poll, F 109 Nai, Yu-Shin 19, 193 Levin, S 311 Marros, MM 30 Nojera-Rincon, MB 234 Levis, E 301 Marshall, S 241 Nakai, M 100, 164, 19 Li, Belbei 314 Marshman, BC 113 Nakayinga, R 21 Li, Fei 314 Martinez, LA 228 Nakayinga, R 21 Li, Fei 314 Martinez, EA 228 Nayori, SA 268 Li, Hanchen 125 Martine, E 207 Narva, K 119, 37, 21 Li, Mianchen 155 Martine, E 207 Navai, D 283 Li, Alanchen 165 Masson, F 136 Navai, D 283 Li, Manchen 158 Matson, J 45 Nelson, M 45, 126 L	Lei, Chengfeng	163	Manzoor, M	268		
Leonard, K 208 Marfetán, A 57 Nagy, E 189 Letherer, T 119 Marion-Poll, F 109 Nai, Yu-Shin 19, 32 Lewin, S 311 Marros, MM 30 Najer-Rincon, MB 234 Lewis, E 301 Marshall, S 241 Nakai, M 100, 164, 16 Li-Leger, E 266 Marshman, BC 113 199, 304 199, 304 Li, Beibei 314 Martino, EM 98 Naqvi, SA 268 Li, Hanchen 125 Martins, E 207 Narva, K 119, 137, 21 Li, Ming 165 Masson, F 136 Navia, D 283 Li, Zhaofei 67 Mathis, J 45 Nelson, M 45, 126 Liao, Wenli 170 Matisoff, M 222 Nelson, T 241 Lin, Bairong 112 Mayilio, F 316 Neves, PMOJ 255 Liscombe, D 112 McCarthy, EM 33 Nielaus, K 269	Lemaitre, B	136	Marciano, AF	277	NNN	
Leonard, K 208 Marfetán, A 57 Nagy, E 189 Letherer, T 119 Marion-Poll, F 109 Nai, Yu-Shin 19, 19 Lewin, S 311 Marron, MM 30 Najer-Rincan, MB 234 Lewis, E 301 Marshall, S 241 Nakai, M 100, 164, 19 Li-Leger, E 266 Marshman, BC 113 199, 304 199, 304 Li, Beibei 314 Martino, EM 98 Naqvi, SA 268 Li, Fei 314 Martino, EM 98 Naqvi, SA 268 Li, Hanchen 125 Martins, E 207 Nava, N 119, 137, 21 Li, Ming 165 Masson, F 136 Navia, D 283 Li, Janchen 73 Ndakidemi, PA 52, 226 Li, Janeie 67 Mathis, J 45 Nelson, M 45, 126 Liao, Wenli 170 Matisoff, M 222 Nelson, M 261 Lin, Bairong 217 Ma	Lenzen, S	220	Marcomini, MC	255	Nagoshi, R	137
Letherer, T 119 Marion-Poll, F 109 Nai, Yu-Shin 19, 193 Lewin, S 311 Marros, MM 30 Najera-Rincon, MB 234 Lewis, E 301 Marshanl, S 241 Nakai, M 100, 164, 19 Li-Leger, E 266 Marshman, BC 113 199, 304 199, 304 Li, Beibei 314 Martinez, LA 228 Nakayinga, R 21 11, 19, 193 11, 193 11, 193 11, 193 11, 193 11, 193 11, 193 11, 193 11, 193	•	208	· ·		=	189
Levin, S 311 Marros, MM 30 Najera-Rincon, MB 24 Lewis, E 301 Marshall, S 241 Nakai, M 100, 164, 151 Li-Leger, E 266 Marshall, S 241 Nakai, M 100, 164, 151 Li, Beibei 314 Martinez, LA 228 Nakayinga, R 21 Li, Fei 314 Martine, EM 98 Naqvi, SA 268 Li, Hanchen 125 Martins, E 207 Narva, K 119, 137, 23 Li, Ming 165 Masson, F 136 Navia, D 283 Li, Xianchun 73 Mathew, LG 73 Ndakidemi, PA 52, 226 Li, Zhaofei 67 Mathis, J 45 Nelson, M 45, 126 Liao, Wenli 170 Matisoff, M 222 Nelson, M 45, 126 Liao, Wenli 170 Mavilio, F 54 Neves, de Oliveira, MC 261 Lin, Bairong 173 Mavilio, F 316 Neves, PMOJ 255	,		·			
Lewis, E 301 Marshall, S 241 Nakai, M 100, 164, 165 Li-Leger, E 266 Marshman, BC 113 199, 304 199, 304 Li, Beibei 314 Martino, EM 98 Naqvi, SA 268 Li, Hanchen 125 Martins, E 207 Narva, K 119, 137, 23 Li, Ming 165 Masson, F 136 Navia, D 283 Li, Xianchun 73 Mathew, LG 73 Ndakidemi, PA 52, 226 Liao, Wenli 170 Matisoff, M 222 Nelson, M 45, 126 Liao, Wenli 170 Matsumoto, TK 54 Neves, PMOJ 255 Liao, Wenli 170 Matsumoto, TK 54 Neves, PMOJ 255 Liscombe, D 112 Mbega, ER 52, 226 Niehaus, K 269 Little, NS 74 McCarthy, EM 195 Nielsen-LeRoux, C 78, 124 Liu, Liu 126 McKinney, DA 173 Noon, J 125	•		•		·	
Li-Leger, E 266 Marshman, BC 113 199, 304 4 Li, Beibei 314 Martinez, LA 228 Nakayinga, R 21 Li, Fei 314 Martino, EM 98 Naqvi, SA 268 Li, Hanchen 125 Martins, E 207 Narva, K 119, 137, 25 Li, Ming 165 Masson, F 136 Navia, D 283 Li, Xianchun 73 Mathew, LG 73 Ndakidemi, PA 52, 226 Li, Zhaofei 67 Mathis, J 45 Nelson, M 45, 126 Liao, Wenli 170 Matisoff, M 222 Nelson, T 241 Libeberman, B 158 Matsumoto, TK 54 Neves, PMOJ 255 Liscombe, D 112 Mbega, ER 52, 226 Niehaus, K 269 Litu, Jisheng 170 McCarthy, EM 195 Nielsen-Lenux, C 78, 124 Liu, Jisheng 170 McDonnell, RI 33 Niou L 74 Liu, Jisheng 170 McKinnery, DA 173 Noon, J			•		•	
Li, Beibei 314 Martinez, LA 228 Nakayinga, R 21 Li, Fei 314 Martino, EM 98 Naqvi, SA 268 Li, Hanchen 125 Martins, E 207 Narva, K 119, 137, 21 Li, Ming 165 Masson, F 136 Navia, D 283 Li, Xianchun 73 Mathew, LG 73 Ndakidemi, PA 52, 226 Li, Zhaofei 67 Matisoff, M 222 Nelson, M 45, 126 Liao, Wenli 170 Matisoff, M 222 Nelson, T 241 Lieberman, B 158 Matsumoto, TK 54 Neves de Oliveira, MC 261 Lin, Bairong 217 Mavilio, F 316 Neves, PMOJ 255 Liscombe, D 112 Mbega, ER 52, 226 Niehaus, K 269 Little, NS 74 McCarthy, EM 195 Nielsen-LeRoux, C 78, 124 Liu, Jisheng 170 McCoarthy, EM 173 Noon, J 125 Liu, Jing-Kun 136 McKinney, DA 307 Nowatzki, TM			•		Nakai, IVI	
Li, Fei 314 Martino, EM 98 Naqvi, SA 268 Li, Hanchen 125 Martins, E 207 Narva, K 119, 137, 21 Li, Ming 165 Masson, F 136 Navia, D 283 Li, Xianchun 73 Mathew, LG 73 Ndakidemi, PA 52, 226 Li, Zhaofei 67 Mathis, J 45 Nelson, M 45, 126 Liao, Wenli 170 Matisoff, M 222 Nelson, T 241 Lieberman, B 158 Matsumoto, TK 54 Neves, de Oliveira, MC 261 Lin, Bairong 217 Mavilio, F 316 Neves, PMOJ 255 Liscombe, D 112 Mbega, ER 52, 226 Niehaus, K 269 Little, NS 74 McCarthy, EM 195 Nielsen-LeRoux, C 78, 124 Liu, Jisheng 170 McConnell, RJ 33 Niu L 74 Liu, Ming-Kun 318 McKinney, DA 173 Noon, J 125 Liu, Ming-Kun 318 McKinstry, M 0ral-Msp #1 Nunes, GdaS	•		· · · · · · · · · · · · · · · · · · ·			-
Li, Hanchen 125 Martins, E 207 Narva, K 119, 137, 21 Li, Ming 165 Masson, F 136 Navia, D 283 Li, Xianchun 73 Mathew, LG 73 Ndakidemi, PA 52, 226 Li, Zhaofei 67 Mathis, J 45 Nelson, M 45, 126 Liao, Wenli 170 Matisoff, M 222 Nelson, T 241 Libebrman, B 158 Matsumoto, TK 54 Neves de Oliveira, MC 261 Lin, Bairong 217 Mavilio, F 316 Neves, PMOJ 255 Liscombe, D 112 Mbega, ER 52, 226 Niehaus, K 269 Little, NS 74 McCarthy, EM 195 Nielsen-LeRoux, C 78, 124 Liu, Jusheng 170 McConnell, RJ 33 Niu L 74 Liu, Jusheng 170 McKinney, DA 173 Noon, J 125 Liu, Ming-Kun 318 McKinney, DA 307 Nowatzki, TM 126 Liuza, L 62 McKinney, DA 38 Nunes, GdaS	Li, Beibei		Martinez, LA		Nakayinga, R	
Li, Ming 165 Masson, F 136 Navia, D 283 Li, Xianchun 73 Mathew, LG 73 Ndakidemi, PA 52, 226 Li, Zhaofei 67 Mathis, J 45 Nelson, M 45, 126 Liao, Wenli 170 Matisoff, M 222 Nelson, T 241 Lieberman, B 158 Matsumoto, TK 54 Neves, PMOJ 255 Liscombe, D 112 Mavilio, F 316 Neves, PMOJ 255 Liscombe, D 112 Mbega, ER 52, 226 Niehaus, K 269 Little, NS 74 McCarthy, EM 195 Nielsen-LeRoux, C 78, 124 Liu, Jisheng 170 McCorthy, EM 195 Niel Sen-LeRoux, C 78, 124 Liu, Jisheng 170 McKinner, DA 173 Noon, J 125 Liu, Ming-Kun 318 McKinner, DA 307 Nowatzki, TM 126 Liuza, L 62 McKinstry, M 0ral-Msp #1 Nunes, GdaS 206 Lopez, RP 203 McMalen, JG 103 <td< td=""><td></td><td>314</td><td>Martino, EM</td><td></td><td></td><td></td></td<>		314	Martino, EM			
Li, Xianchun 73 Mathew, LG 73 Ndakidemi, PA 52, 226 Li, Zhaofei 67 Mathis, J 45 Nelson, M 45, 126 Liao, Wenli 170 Matisoff, M 222 Nelson, T 241 Lieberman, B 158 Matsumoto, TK 54 Neves de Oliveira, MC 261 Lin, Bairong 217 Mavillo, F 316 Neves, PMOJ 255 Liscombe, D 112 Mbega, ER 52, 226 Niehaus, K 269 Little, NS 74 McCarthy, EM 195 Nielsen-LeRoux, C 78, 124 Liu, Jisheng 170 McDonnell, RJ 33 Niu L 74 Liu, Lu 126 McKinney, DA 173 Noon, J 125 Liu, Ming-Kun 318 McKinney, DA 173 Nowatzki, TM 126 Liuza, L 62 McKinstry, M Oral-Msp #1 Nunes, GdaS 206 Loeher, MM 113 McMelannin, A 38 Nuñez-Valdez, ME 168 López Plantey, RJ 56, Oral McQuade, R 169, Poster <	Li, Hanchen	125	Martins, E	207	Narva, K	119, 137, 211
Li, Xianchun 73 Mathew, LG 73 Ndakidemi, PA 52, 226 Li, Zhaofei 67 Mathis, J 45 Nelson, M 45, 126 Liao, Wenli 170 Matisoff, M 222 Nelson, T 241 Lieberman, B 158 Matsumoto, TK 54 Neves de Oliveira, MC 261 Lin, Bairong 217 Mavillor, F 316 Neves, PMOJ 255 Liscombe, D 112 Mbega, ER 52, 226 Niehaus, K 269 Little, NS 74 McCarthy, EM 195 Nielsen-LeRoux, C 78, 124 Liu, Jisheng 170 McDonnell, RJ 33 Niu L 74 Liu, Lu 126 McKinney, DA 173 Noon, J 125 Liu, Ming-Kun 318 McKinnon, AC 307 Nowatzki, TM 126 Liuza, L 62 McKinstry, M Oral-Msp #1 Nunes, GdaS 206 Loeher, MM 113 McMelan, JG 103 López Plantey, RJ 56, Oral McQuade, R 169, Poster López-Dias, JA <td< td=""><td></td><td>165</td><td>Masson, F</td><td></td><td></td><td></td></td<>		165	Masson, F			
Li, Zhaofei 67 Mathis, J 45 Nelson, M 45, 126 Liao, Wenli 170 Matisoff, M 222 Nelson, T 241 Lieberman, B 158 Matsumoto, TK 54 Neves de Oliveira, MC 261 Lin, Bairong 217 Mavilio, F 316 Neves, PMOJ 255 Liscombe, D 112 Mbega, ER 52, 226 Niehaus, K 269 Little, NS 74 McCarthy, EM 195 Nielsen-LeRoux, C 78, 124 Liu, Jisheng 170 McDonnell, RJ 33 Niu L 74 Liu, Liu 126 McKinney, DA 173 Noon, J 125 Liu, Ming-Kun 318 McKinney, DA 173 Nowatzki, TM 126 Liuza, L 62 McKinstry, M Oral-Msp #1 Nunes, GdaS 206 Loeher, MM 113 McMenamin, A 38 Nuñez-Valdez, ME 168 López Plantey, RJ 56, Oral McQuade, R Bac26, Poster López-Dias, JA 47 Meagher, R Bac27 137 Lore	-					
Liao, Wenli 170 Matisoff, M 222 Nelson, T 241 Lieberman, B 158 Matsumoto, TK 54 Neves de Oliveira, MC 261 Lin, Bairong 217 Mavilio, F 316 Neves, PMOJ 255 Liscombe, D 112 Mbega, ER 52, 226 Nielaus, K 269 Little, NS 74 McCarthy, EM 195 Nielsen-LeRoux, C 78, 124 Liu, Jisheng 170 McDonnell, RJ 33 Niu L 74 Liu, Lu 126 McKinney, DA 173 Noon, J 125 Liu, Ming-Kun 318 McKinnon, AC 307 Nowatzki, TM 126 Liuza, L 62 McKinstry, M 0ral-Msp #1 Nunes, GdaS 206 Loeher, MM 113 McMenamin, A 38 Nuñez-Valdez, ME 168 López, Plantey, RJ 56, Oral McQuade, R 169, Poster López-Dias, JA 47 8ac26, Poster López-Dias, JA 47 8ac27 Lorenz, SC 271 Medina, P 256	•		·		-	
Lieberman, B 158 Matsumoto, TK 54 Neves de Oliveira, MC 261 Lin, Bairong 217 Mavilio, F 316 Neves, PMOJ 255 Liscombe, D 112 Mbega, ER 52, 226 Nielaus, K 269 Little, NS 74 McCarthy, EM 195 Nielsen-LeRoux, C 78, 124 Liu, Jisheng 170 McDonnell, RJ 33 Niu L 74 Liu, Lu 126 McKinney, DA 173 Noon, J 125 Liu, Ming-Kun 318 McKinnon, AC 307 Nowatzki, TM 126 Liuza, L 62 McKinstry, M Oral—Msp #1 Nunes, GdaS 206 Loeher, MM 113 McMenamin, A 38 Nuñez-Valdez, ME 168 López, RP 203 McMullen, JG 103 López Plantey, RJ 56, Oral McQuade, R 169, Poster López-Dias, JA 47 8ac26, Poster López-Dias, JA 47 McGarthy, E 137 Lorenz, SC 271 Medina, P 256	•		· ·		•	
Lin, Bairong 217 Mavilio, F 316 Neves, PMOJ 255 Liscombe, D 112 Mbega, ER 52, 226 Niehaus, K 269 Little, NS 74 McCarthy, EM 195 Nielsen-LeRoux, C 78, 124 Liu, Jisheng 170 McDonnell, RJ 33 Niu L 74 Liu, Lu 126 McKinney, DA 173 Noon, J 125 Liu, Ming-Kun 318 McKinnon, AC 307 Nowatzki, TM 126 Liuza, L 62 McKinstry, M Oral-Msp #1 Nunes, GdaS 206 Loeher, MM 113 McMenamin, A 38 Nuñez-Valdez, ME 168 Lopez, RP 203 McMullen, JG 103 López Plantey, RJ 56, Oral McQuade, R 169, Poster López-Dias, JA 47 Bac26, Poster López-Dias, JA 47 Meagher, R 137 Lorent, SC 271 Medina, P 256			•			
Liscombe, D 112 Mbega, ER 52, 226 Niehaus, K 269 Little, NS 74 McCarthy, EM 195 Nielsen-LeRoux, C 78, 124 Liu, Jisheng 170 McDonnell, RJ 33 Niu L 74 Liu, Lu 126 McKinney, DA 173 Noon, J 125 Liu, Ming-Kun 318 McKinnon, AC 307 Nowatzki, TM 126 Liuza, L 62 McKinstry, M Oral-Msp #1 Nunes, GdaS 206 Loeher, MM 113 McMenamin, A 38 Nuñez-Valdez, ME 168 Lopez, RP 203 McMullen, JG 103 Nuñez-Valdez, ME 168 López Plantey, RJ 56, Oral McQuade, R 169, Poster López-Dias, JA 47 Bac27 Lorchler, L 27 Meagher, R 137 Lorenz, SC 271 Medina, P 256	-		-			
Little, NS 74 McCarthy, EM 195 Nielsen-LeRoux, C 78, 124 Liu, Jisheng 170 McDonnell, RJ 33 Niu L 74 Liu, Lu 126 McKinney, DA 173 Noon, J 125 Liu, Ming-Kun 318 McKinnon, AC 307 Nowatzki, TM 126 Liuza, L 62 McKinstry, M Oral-Msp #1 Nunes, GdaS 206 Loeher, MM 113 McMenamin, A 38 Nuñez-Valdez, ME 168 Lopez, RP 203 McMullen, JG 103 López Plantey, RJ 56, Oral McQuade, R 169, Poster López-Dias, JA 47 Meagher, R Bac27 Lorchler, L 27 Meagher, R 137 Lorenz, SC 271 Medina, P 256	_		·			
Liu, Jisheng 170 McDonnell, RJ 33 Niu L 74 Liu, Lu 126 McKinney, DA 173 Noon, J 125 Liu, Ming-Kun 318 McKinnon, AC 307 Nowatzki, TM 126 Liuza, L 62 McKinstry, M Oral-Msp #1 Nunes, GdaS 206 Loeher, MM 113 McMenamin, A 38 Nuñez-Valdez, ME 168 Lopez, RP 203 McMullen, JG 103 López Plantey, RJ 56, Oral McQuade, R 169, Poster López-Dias, JA 47 Bac26, Poster Lorchler, L 27 Meagher, R 137 Lorenz, SC 271 Medina, P 256	Liscombe, D		Mbega, ER		Niehaus, K	
Liu, Lu 126 McKinney, DA 173 Noon, J 125 Liu, Ming-Kun 318 McKinnon, AC 307 Nowatzki, TM 126 Liuza, L 62 McKinstry, M Oral-Msp #1 Nunes, GdaS 206 Loeher, MM 113 McMenamin, A 38 Nuñez-Valdez, ME 168 Lopez, RP 203 McMullen, JG 103 Vinez-Valdez, ME 169 López Plantey, RJ 56, Oral McQuade, R 169, Poster López-Dias, JA 47 Bac27 Lorchler, L 27 Meagher, R 137 Lorenz, SC 271 Medina, P 256	Little, NS	74	McCarthy, EM	195	Nielsen-LeRoux, C	78, 124
Liu, Ming-Kun 318 McKinnon, AC 307 Nowatzki, TM 126 Liuza, L 62 McKinstry, M Oral–Msp #1 Nunes, GdaS 206 Loeher, MM 113 McMenamin, A 38 Nuñez-Valdez, ME 168 Lopez, RP 203 McMullen, JG 103 López Plantey, RJ 56, Oral McQuade, R 169, Poster López-Dias, JA 47 Bac26, Poster Lorchler, L 27 Meagher, R 137 Lorenz, SC 271 Medina, P 256	Liu, Jisheng	170	McDonnell, RJ	33	Niu L	74
Liu, Ming-Kun 318 McKinnon, AC 307 Nowatzki, TM 126 Liuza, L 62 McKinstry, M Oral–Msp #1 Nunes, GdaS 206 Loeher, MM 113 McMenamin, A 38 Nuñez-Valdez, ME 168 Lopez, RP 203 McMullen, JG 103 López Plantey, RJ 56, Oral McQuade, R 169, Poster López-Dias, JA 47 Bac27 Lorchler, L 27 Meagher, R 137 Lorenz, SC 271 Medina, P 256		126	McKinney, DA	173	Noon, J	125
Liuza, L 62 McKinstry, M Oral–Msp #1 Nunes, GdaS 206 Loeher, MM 113 McMenamin, A 38 Nuñez-Valdez, ME 168 Lopez, RP 203 McMullen, JG 103 López Plantey, RJ 56, Oral McQuade, R 169, Poster López-Dias, JA 47 Bac26, Poster Lorchler, L 27 Meagher, R 137 Lorenz, SC 271 Medina, P 256	Liu, Lu		-			
Loeher, MM 113 McMenamin, A 38 Nuñez-Valdez, ME 168 Lopez, RP 203 McMullen, JG 103 López Plantey, RJ 56, Oral McQuade, R 169, Poster López-Dias, JA 47 Bac26, Poster Lorchler, L 27 Meagher, R 137 Lorenz, SC 271 Medina, P 256	•				· · · · · · · · · · · · · · · · · · ·	
Lopez, RP 203 McMullen, JG 103 López Plantey, RJ 56, Oral McQuade, R 169, Poster López-Dias, JA 47 Bac26, Poster Lorchler, L 27 Meagher, R 137 Lorenz, SC 271 Medina, P 256	Liu, Ming-Kun	318	•	Oral–Msn #1		
López Plantey, RJ 56, Oral McQuade, R 169, Poster López-Dias, JA 47 Bac26, Poster Lorchler, L 27 Meagher, R 137 Lorenz, SC 271 Medina, P 256	Liu, Ming-Kun Liuza, L	318 62	McKinstry, M	•		
Fun#3 Bac26, Poster López-Dias, JA 47 Bac27 Lorchler, L 27 Meagher, R 137 Lorenz, SC 271 Medina, P 256	Liu, Ming-Kun Liuza, L Loeher, MM	318 62 113	McKinstry, M McMenamin, A	38		
López-Dias, JA 47 Bac27 Lorchler, L 27 Meagher, R 137 Lorenz, SC 271 Medina, P 256	Liu, Ming-Kun Liuza, L Loeher, MM Lopez, RP	318 62 113 203	McKinstry, M McMenamin, A McMullen, JG	38 103		
Lorchler, L 27 Meagher, R 137 Lorenz, SC 271 Medina, P 256	Liu, Ming-Kun Liuza, L Loeher, MM Lopez, RP	318 62 113 203 56, <i>Oral</i>	McKinstry, M McMenamin, A McMullen, JG	38 103 169 , Poster		
Lorenz, SC 271 <i>Medina, P</i> 256	Liu, Ming-Kun Liuza, L Loeher, MM Lopez, RP	318 62 113 203 56, <i>Oral</i>	McKinstry, M McMenamin, A McMullen, JG	38 103 169 , Poster		
Lorenz, SC 271 <i>Medina, P</i> 256	Liu, Ming-Kun Liuza, L Loeher, MM Lopez, RP López Plantey, RJ	318 62 113 203 56, <i>Oral</i> <i>Fun#3</i>	McKinstry, M McMenamin, A McMullen, JG	38 103 169 , Poster Bac26, Poster		
	Liu, Ming-Kun Liuza, L Loeher, MM Lopez, RP <i>López Plantey, RJ</i> López-Dias, JA	318 62 113 203 56, <i>Oral</i> <i>Fun#3</i> 47	McKinstry, M McMenamin, A McMullen, JG <i>McQuade, R</i>	38 103 169 , Poster Bac26, Poster Bac27		
toriday, 1 order bucks (Miccus, 1 of order bucks)	Liu, Ming-Kun Liuza, L Loeher, MM Lopez, RP <i>López Plantey, RJ</i> López-Dias, JA Lorchler, L	318 62 113 203 56, <i>Oral</i> <i>Fun#3</i> 47	McKinstry, M McMenamin, A McMullen, JG <i>McQuade, R</i> Meagher, R	38 103 169 , Poster Bac26, Poster Bac27 137		
02	Liu, Ming-Kun Liuza, L Loeher, MM Lopez, RP <i>López Plantey, RJ</i> López-Dias, JA Lorchler, L Lorenz, SC	318 62 113 203 56, <i>Oral</i> <i>Fun#3</i> 47 27	McKinstry, M McMenamin, A McMullen, JG <i>McQuade, R</i> Meagher, R <i>Medina, P</i>	38 103 169, Poster Bac26, Poster Bac27 137 256		

000	0	Praça, LB	200 , 201 , 207	
D'Callaghan, M	303	Prieto, E	176	
h, JongMin	194, 224	Prince, G	90	
.,,8	Poster Msp06	Przyklenk, M	92, 270, 271,	
nkawa, T	37	,,	272	
nta, R	197	Pushparajan, C	312	
Oliveira, AS	105			
mbura, LO	248	QQ	Q	
ppert, B	208	Qi, Jiaheling	265	
ral, J	35	Qi, Xiuli	126	
Ordonez-Acevedo, LG	139	Qin, Fujun	163	
Stroff, G	125	Qin, Weiwen	191	
Dzgen, A	149	Qiu, Jianxiang	99	
zşahin, E	189	Quan Yudong	79	
		Queiroz, P	207	
PPF		Quesada-Moraga, E	153, 179, 243	
icheco, S	122	Qureshi, N	166	
ne, TD	33			
xao, FRS	106, 242	RRI	₹	:
uch, M	293	Raad, M	307	S
n, Mengjia	96	Ramachandran, P	Poster Bac23	Sa
ura, D	Oral Fun#3	Ramalho, DG	262	Sa
edes, JC	136	Ramaswamy, S	290	Sa
k, Dong Hwan	214, 239, 258	Ramsden, M	90	Sc
k, Hyun-Woo	<i>75</i>	Randall, JL	138	Scl
rk, Min Gu	214 , 239, 258	Rangappa, K	Poster Mcn26	Sch
rk, So Eun	180, 240 ,	Rangel, DEN	105 , 106	Sco
	Poster Fun25	Rasmussen, KJ	289	Seat
rk, Tae Hyun	180	Rauscher, G	45	Seha
rker, AG	117, 118	Ravulapalii, R	59	Sela,
rker, BL	51	Raya-Ortega MdC	243	Shako
iscoe, B	287	Raymond, B	72 , 202, 287	Shank
ssarelli, AL	98	Reavey, CE	138	Shank
tel, AV	50, 92 , 156,	Reddy, GVP	264, 279	Shapri
	269, 270, 271,	Rejasse, A	124	Sharm
	272	Reall, T	Oral Mcn#2	Shelby
ten, P	49	Reyes, L	174	Sheppa
Irini, N	106, 176, 242	Riaño, LG	276	Shi, Jia
r-Cardeña, A	43	Riberiro, BM	62, 93 , 190	Shikano
ce, HE	211	Rice, A	260	Shilova
dell, S	198	Rice, J	126	Shin, Ta
, Donghai	219, 287	Ricietto, APS	123	Shore-N
, Guoxiong	291	Riddick, E	278	Short, JF
, Yan	217	Riegler, M	172, 257	Shrestho
e-Ranney, C	88	Ringbauer, JA	Oral Mcn#2	Shu, Cho
ra-Junior, RA	106	Ríos-Moreno, A	179, 243	Cia mfui
ra, C	275 7 4	Riquelme, AE	Oral Fun#3	Siegfried
era, OP	74 245	Rist, M <i>Rivas, F</i>	288, 289 14 , 282 , 307	Siehl, DL Sihler, W
notto, WMS				Sinier, w
y, D	155	Rivlin, ND	116 , 305	Silva, CE
erson, BF	103	Roberts, DW	2 , 106, 275 29	*
er, F	27	Roder, A		Šima, P
ce, CA	74 126	Rodrigues, J	230 , 231	Simmon
ner, C , JF	126 261	Rodriguez-Segura, Z	168	Simpsor
		Rodriguez, P	125	Slavicek
uolo, PH chuk, S	Oral Fun#3	Roemer, M	289 181	Smaggh
	133	Rohrmann, G		Smid, HN
datz, J		Rojas, E	232	Smith, S
	56		40 150 160	Cnadaur
	204, 205, 206	Ros, VID	40, 159 , 160	•
e, JC	204, 205, 206 176	Ros, VID Rosen, B	35	Snow, JW
anczyk, RA nce, JC nuraj, J	204, 205, 206 176 73	Ros, VID Rosen, B Ross, S	35 115	• .
ce, JC nuraj, J celevé, C	204, 205, 206 176 73 16	Rosen, B Roses, S Rostás, M	35 115 282, 307	Soares, CM
ce, JC nuraj, J televé, C tilla, M	204, 205, 206 176 73 16 278	Ros, VID Rosen, B Ross, S Rostás, M Rowley, DL	35 115 282, 307 20	Snow, JW
e, JC uraj, J elevé, C	204, 205, 206 176 73 16	Rosen, B Roses, S Rostás, M	35 115 282, 307	Snow, JW Soares, CM

Solter, L	11 , 26, 302	Upasani, SM	141	Wei, Jun-Zhi	35
Sommer, RJ	104	Urrutia, A	115	Weihrauch, B	85
Song, Fuping	78	Uzel, GD	118	Weinmaier, T	Poster Bac23
Sopko, M	119			Weiss, LM	132
Sosa-Gomez, DR	190, 261	VVV		Welch, MD	37
Sowa, JN	221	Vacari, AM	204, 206, 262	Weng, Shaoping	187
Spears, T	142	Valicente, FH	12	Wennmann, JT	85, 86, 100
Spieth, P	269	Valle, NB	234	Willcoxon, MI	48
Spooner-Hart, RN	257	Valenzuela, E	232	Williams, BAP	24
Sritunyalucksana, K	24	Valzania, L	173	Williams, RJ	Poster Bac23
Stanley, D	Oral Mcn#2	van Aerle, R	296, 297	Williams, T	304
St. Leger, RJ	110 , 152	van Houte, S	160	Williamson, A	277
Steinwender, B	91	van Lent, JWM	161	Willis, LG	17, 68
Stentiford, G	10, 24, 66,	van Oers, MM	40 , 68, 117,	Wilson, K	138, 260
,	128, 296	,	159, 160, 161,	Wiredu-Boakye, D	24
Stephan, D	293		316	Woo, Jin Kim	258
Stevens, GN	253, 254	van Rie, J	213	Woo, Ra Mi	214, 239 , 258
Stock, SP	29, 103, 169,	Vélez, A	13	Woo, SooDong	194, 224, 273,
,	Poster Bac26,	Verduzco-Rosas, LA	139	,	274, Poster
Poster Bac27	,	Vidal, S	92, 156, 272		Msp06,
Strand, Mark R	173	Viera, W	283		Poster Vir21
Strand, Michael R	173	Villamizar, L	183, 184, 241 ,	Wraight, SP	54
Strange, JP	63	· · · · · · · · · · · · · · · · · · ·	276	Wu, Gusui	45, 49, 126
Subramanian, S	248	Visnovsky, GA	312	Wu, Lijuan	94
Suijkerbuijk, H	159	Viswanathan, VK	Poster Bac27	Wu, Pei-Chi	148
Sun, Ming	219 , 287	Vlak, JM	7 , 117	Wu, Songqing	217
Sun, Xiulian	163	Vogel, KJ	173	Wu, Tzong-Yuan	318
Swevers, L	170	Volkman, LE	37	Wu, Wenbi	96, 97, 99,
		Vossbrinck, BA	23	,	195
TTT		Vossbrinck, CR	23, 135, 284	Wu, Y	Poster Bac23
Tabashnik, BE	73, 150	Vreysen, MJB	117, 118	Wu, Yueh-Lung	148
Tajima, S	315	,	,	Wu,Gusui	35
Takahash, D	237	wwv	V		
		Wakil. W	294 . 295	XXX	(
Takahashi, N	265	<i>Wakil, W</i> Walsh. T	294 , 295 208	X X X X	
Takahashi, N Takamatsu, T	265 199	Walsh, T	208	Xia, Yuxian	111, 291
Takahashi, N Takamatsu, T Talaei-Hassanloui, R	265	Walsh, T Wang, Chung-Hsiung	208 19	Xia, Yuxian Xie, Weiping	111, 291 126
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee	265 199 263	Walsh, T Wang, Chung-Hsiung Wang, Dun	208 19 86	Xia, Yuxian Xie, Weiping Xing, Longsheng	111, 291 126 Oral Vir#5
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping	265 199 263 119	Walsh, T Wang, Chung-Hsiung	208 19	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali	111, 291 126
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S	265 199 263 119 142 46	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan <i>Wang, Haidong</i>	208 19 86 211 313	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui	111, 291 126 Oral Vir#5 145 163
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin	265 199 263 119 142 46 99, 195	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong	208 19 86 211 313 163	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan	111, 291 126 Oral Vir#5 145 163 23
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR	265 199 263 119 142 46	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin	208 19 86 211 313	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui	111, 291 126 Oral Vir#5 145 163
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D	265 199 263 119 142 46 99, 195 288, 289 237	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB	208 19 86 211 313 163 Oral Vir#1	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei	111, 291 126 Oral Vir#5 145 163 23 217
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D	265 199 263 119 142 46 99, 195 288, 289	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen	208 19 86 211 313 163 Oral Vir#1 110	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan	111, 291 126 Oral Vir#5 145 163 23 217
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Taylor, CG	265 199 263 119 142 46 99, 195 288, 289 237 125 35	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Y Y Y	111, 291 126 Oral Vir#5 145 163 23 217
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen	208 19 86 211 313 163 Oral Vir#1 110	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Yalpani, N Yamamoto, T	111, 291 126 Oral Vir#5 145 163 23 217
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68 , 161,	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#1,	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Yalpani, N Yamamoto, T Yanagawa, A	111, 291 126 Oral Vir#5 145 163 23 217
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#1, Oral Vir#5	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Yalpani, N Yamamoto, T	111, 291 126 Oral Vir#5 145 163 23 217
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D Theilmann, DA	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68, 161, 181, 182	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli Wang, Ping Wang, Qianqian	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#1, Oral Vir#5 208	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Yalpani, N Yamamoto, T Yanagawa, A	111, 291 126 Oral Vir#5 145 163 23 217 126 48, 49 109 96, 97, 99 ,
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D Theilmann, DA	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68, 161, 181, 182	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli Wang, Ping Wang, Qianqian Wang, Qianran	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#1, Oral Vir#5 208	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Yalpani, N Yamamoto, T Yanagawa, A Yang, Kai Yang, Ming-Jun	111, 291 126 Oral Vir#5 145 163 23 217 126 48, 49 109 96, 97, 99 , 195
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D Theilmann, DA	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68, 161, 181, 182 182	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli Wang, Ping Wang, Qianqian	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#1, Oral Vir#5 208 187 Oral Vir#5	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Yalpani, N Yamamoto, T Yanagawa, A Yang, Kai	111, 291 126 Oral Vir#5 145 163 23 217 126 48, 49 109 96, 97, 99 , 195 165
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D Theilmann, DA Thézé, J Thiéry, D Tokarev, Y	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68, 161, 181, 182 182 56, Oral Fun#3	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli Wang, Qianqian Wang, Qianran Wang, Xiujuan Wang, Yueqin	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#5 208 187 Oral Vir#5 119	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Yalpani, N Yamamoto, T Yanagawa, A Yang, Kai Yang, Ming-Jun Yang, Shili Yang, Yang	111, 291 126 Oral Vir#5 145 163 23 217 126 48, 49 109 96, 97, 99 , 195 165 163
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D Theilmann, DA Thézé, J Thiéry, D	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68, 161, 181, 182 182 56, Oral Fun#3 135 78	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli Wang, Qianqian Wang, Qianran Wang, Xiujuan Wang, Yueqin Wang, Zengxia	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#5 208 187 Oral Vir#5 119 79	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Yalpani, N Yamamoto, T Yanagawa, A Yang, Kai Yang, Ming-Jun Yang, Shili	111, 291 126 Oral Vir#5 145 163 23 217 126 48, 49 109 96, 97, 99 , 195 165 163 165 107
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D Theilmann, DA Thézé, J Thiéry, D Tokarev, Y Tong, Zongxing	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68, 161, 181, 182 182 56, Oral Fun#3 135	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli Wang, Qianqian Wang, Qianran Wang, Xiujuan Wang, Yueqin	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#5 208 187 Oral Vir#5 119 79 80	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Yalpani, N Yamamoto, T Yanagawa, A Yang, Kai Yang, Ming-Jun Yang, Shili Yang, Yang Yang, Yi-Ting	111, 291 126 Oral Vir#5 145 163 23 217 126 48, 49 109 96, 97, 99 , 195 165 163 165
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D Theilmann, DA Thézé, J Thiéry, D Tokarev, Y Tong, Zongxing Toprak, U	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68, 161, 181, 182 182 56, Oral Fun#3 135 78 77, 162, 186	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli Wang, Ping Wang, Qianqian Wang, Qianran Wang, Xiujuan Wang, Yueqin Wang, Zengxia Wang, Zeyu Wang, Zhenying	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#5 208 187 Oral Vir#5 119 79 80 122	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Yalpani, N Yamamoto, T Yanagawa, A Yang, Kai Yang, Ming-Jun Yang, Shili Yang, Yang Yang, Yi-Ting Yao, Qin Yasin, M	111, 291 126 Oral Vir#5 145 163 23 217 126 48, 49 109 96, 97, 99 , 195 165 163 165 107 145, 146, 147 294, 295
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D Theilmann, DA Thézé, J Thiéry, D Tokarev, Y Tong, Zongxing Toprak, U Torres-Quintero, MC	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68, 161, 181, 182 182 56, Oral Fun#3 135 78 77, 162, 186 47	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli Wang, Ping Wang, Qianqian Wang, Qianran Wang, Xiujuan Wang, Yueqin Wang, Zengxia Wang, Zeyu	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#5 208 187 Oral Vir#5 119 79 80 122 79, 80	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Yalpani, N Yamamoto, T Yanagawa, A Yang, Kai Yang, Ming-Jun Yang, Shili Yang, Yang Yang, Yi-Ting Yao, Qin	111, 291 126 Oral Vir#5 145 163 23 217 126 48, 49 109 96, 97, 99 , 195 165 163 165 107 145, 146, 147
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D Theilmann, DA Thézé, J Thiéry, D Tokarev, Y Tong, Zongxing Toprak, U Torres-Quintero, MC Trimble, J	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68, 161, 181, 182 182 56, Oral Fun#3 135 78 77, 162, 186 47 88	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli Wang, Ping Wang, Qianqian Wang, Qianran Wang, Xiujuan Wang, Yueqin Wang, Zengxia Wang, Zeyu Wang, Zhenying Ward, GM	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#5 208 187 Oral Vir#5 119 79 80 122 79, 80 115	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Yalpani, N Yamamoto, T Yanagawa, A Yang, Kai Yang, Ming-Jun Yang, Shili Yang, Yang Yang, Yi-Ting Yao, Qin Yasin, M Yates, JR	111, 291 126 Oral Vir#5 145 163 23 217 126 48, 49 109 96, 97, 99 , 195 165 163 165 107 145, 146, 147 294, 295 245, 246
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D Theilmann, DA Thézé, J Thiéry, D Tokarev, Y Tong, Zongxing Toprak, U Torres-Quintero, MC Trimble, J Tripodi, AD	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68, 161, 181, 182 182 56, Oral Fun#3 135 78 77, 162, 186 47 88 63	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli Wang, Ping Wang, Qianqian Wang, Qianran Wang, Xiujuan Wang, Yueqin Wang, Zengxia Wang, Zeyu Wang, Zhenying Ward, GM	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#5 208 187 Oral Vir#5 119 79 80 122 79, 80 115 Oral Vir#1,	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Y Y Y Yalpani, N Yamamoto, T Yanagawa, A Yang, Kai Yang, Ming-Jun Yang, Shili Yang, Yang Yang, Yi-Ting Yao, Qin Yasin, M Yates, JR Ye, Sudan Yin, Mengyi	111, 291 126 Oral Vir#5 145 163 23 217 126 48, 49 109 96, 97, 99 , 195 165 163 165 107 145, 146, 147 294, 295 245, 246 236
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D Theilmann, DA Thézé, J Thiéry, D Tokarev, Y Tong, Zongxing Toprak, U Torres-Quintero, MC Trimble, J Tripodi, AD Troemel, ER Tsuruta, K	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68, 161, 181, 182 182 56, Oral Fun#3 135 78 77, 162, 186 47 88 63 134, 221 100	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli Wang, Ping Wang, Qianqian Wang, Qianran Wang, Xiujuan Wang, Yueqin Wang, Zengxia Wang, Zeyu Wang, Zhenying Ward, GM Wang, Xi	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#5 208 187 Oral Vir#5 119 79 80 122 79, 80 115 Oral Vir#1, Oral Vir#5 98	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Yalpani, N Yamamoto, T Yanagawa, A Yang, Kai Yang, Ming-Jun Yang, Shili Yang, Yang Yang, Yi-Ting Yao, Qin Yasin, M Yates, JR Ye, Sudan Yin, Mengyi Yokoyama, T	111, 291 126 Oral Vir#5 145 163 23 217 126 48, 49 109 96, 97, 99 , 195 165 163 165 107 145, 146, 147 294, 295 245, 246 236 Oral Vir#5 144
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D Theilmann, DA Thézé, J Thiéry, D Tokarev, Y Tong, Zongxing Toprak, U Torres-Quintero, MC Trimble, J Tripodi, AD Troemel, ER	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68, 161, 181, 182 182 56, Oral Fun#3 135 78 77, 162, 186 47 88 63 134, 221	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli Wang, Ping Wang, Qianqian Wang, Qianran Wang, Xiujuan Wang, Yueqin Wang, Zengxia Wang, Zeyu Wang, Zhenying Ward, GM Wang, Xi	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#5 208 187 Oral Vir#5 119 79 80 122 79, 80 115 Oral Vir#1, Oral Vir#1, Oral Vir#1	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Yalpani, N Yamamoto, T Yanagawa, A Yang, Kai Yang, Ming-Jun Yang, Shili Yang, Yang Yang, Yi-Ting Yao, Qin Yasin, M Yates, JR Ye, Sudan Yin, Mengyi Yokoyama, T Yoshimura, T	111, 291 126 Oral Vir#5 145 163 23 217 126 48, 49 109 96, 97, 99 , 195 165 163 165 107 145, 146, 147 294, 295 245, 246 236 Oral Vir#5
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D Theilmann, DA Thézé, J Thiéry, D Tokarev, Y Tong, Zongxing Toprak, U Torres-Quintero, MC Trimble, J Tripodi, AD Troemel, ER Tsuruta, K	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68, 161, 181, 182 182 56, Oral Fun#3 135 78 77, 162, 186 47 88 63 134, 221 100	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haichuan Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli Wang, Ping Wang, Qianqian Wang, Qianran Wang, Xiujuan Wang, Yueqin Wang, Zengxia Wang, Zeyu Wang, Zhenying Wang, Xi Warnecke, SA Wassermann, M Weavil-Abueg,ME	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#5 208 187 Oral Vir#5 119 79 80 122 79, 80 115 Oral Vir#1, Oral Vir#5 98 271	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Yalpani, N Yamamoto, T Yanagawa, A Yang, Kai Yang, Ming-Jun Yang, Shili Yang, Yang Yang, Yi-Ting Yao, Qin Yasin, M Yates, JR Ye, Sudan Yin, Mengyi Yokoyama, T Yoshimura, T Yousefelahi, MN	111, 291 126 Oral Vir#5 145 163 23 217 126 48, 49 109 96, 97, 99 , 195 165 163 165 107 145, 146, 147 294, 295 245, 246 236 Oral Vir#5 144 109
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D Theilmann, DA Thézé, J Thiéry, D Tokarev, Y Tong, Zongxing Toprak, U Torres-Quintero, MC Trimble, J Tripodi, AD Troemel, ER Tsuruta, K	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68, 161, 181, 182 182 56, Oral Fun#3 135 78 77, 162, 186 47 88 63 134, 221 100	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli Wang, Ping Wang, Qianqian Wang, Qianran Wang, Xiujuan Wang, Yueqin Wang, Zengxia Wang, Zeyu Wang, Zhenying Ward, GM Wang, Xi Warnecke, SA Wassermann, M Weavil-Abueg,ME Webb, TK	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#5 208 187 Oral Vir#5 119 79 80 122 79,80 115 Oral Vir#1, Oral Vir#5 98 271 114 289	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Yalpani, N Yamamoto, T Yanagawa, A Yang, Kai Yang, Ming-Jun Yang, Shili Yang, Yang Yang, Yi-Ting Yao, Qin Yasin, M Yates, JR Ye, Sudan Yin, Mengyi Yokoyama, T Yoshimura, T Yousefelahi, MN Yu, Dong-Shuai	111, 291 126 Oral Vir#5 145 163 23 217 126 48, 49 109 96, 97, 99 , 195 165 163 165 107 145, 146, 147 294, 295 245, 246 236 Oral Vir#5 144 109 103 165
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D Theilmann, DA Thézé, J Thiéry, D Tokarev, Y Tong, Zongxing Toprak, U Torres-Quintero, MC Trimble, J Tripodi, AD Troemel, ER Tsuruta, K	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68, 161, 181, 182 182 56, Oral Fun#3 135 78 77, 162, 186 47 88 63 134, 221 100	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haichuan Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli Wang, Ping Wang, Qianqian Wang, Qianran Wang, Xiujuan Wang, Yueqin Wang, Zengxia Wang, Zeyu Wang, Zhenying Wang, Xi Warnecke, SA Wassermann, M Weavil-Abueg,ME	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#5 208 187 Oral Vir#5 119 79 80 122 79,80 115 Oral Vir#1, Oral Vir#5 98 271 114	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei Yalpani, N Yamamoto, T Yanagawa, A Yang, Kai Yang, Ming-Jun Yang, Shili Yang, Yang Yang, Yi-Ting Yao, Qin Yasin, M Yates, JR Ye, Sudan Yin, Mengyi Yokoyama, T Yoshimura, T Yousefelahi, MN	111, 291 126 Oral Vir#5 145 163 23 217 126 48, 49 109 96, 97, 99 , 195 165 163 165 107 145, 146, 147 294, 295 245, 246 236 Oral Vir#5 144 109 103
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D Theilmann, DA Thézé, J Thiéry, D Tokarev, Y Tong, Zongxing Toprak, U Torres-Quintero, MC Trimble, J Tripodi, AD Troemel, ER Tsuruta, K Tummala, Y	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68, 161, 181, 182 182 56, Oral Fun#3 135 78 77, 162, 186 47 88 63 134, 221 100	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli Wang, Qianqian Wang, Qianran Wang, Xiujuan Wang, Yueqin Wang, Zengxia Wang, Zeyu Wang, Zeyu Wang, Zhenying Ward, GM Wang, Xi Warnecke, SA Wassermann, M Weavil-Abueg, ME Webb, TK Webster, TC Wedgwood, E	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#1, Oral Vir#5 208 187 Oral Vir#5 119 79 80 122 79,80 115 Oral Vir#1, Oral Vir#5 98 271 114 289	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei YY Yalpani, N Yamamoto, T Yanagawa, A Yang, Kai Yang, Ming-Jun Yang, Shili Yang, Yang Yang, Yi-Ting Yao, Qin Yasin, M Yates, JR Ye, Sudan Yin, Mengyi Yokoyama, T Yoshimura, T Yousefelahi, MN Yu, Dong-Shuai Yuan, Chuanfei	111, 291 126 Oral Vir#5 145 163 23 217 126 48, 49 109 96, 97, 99, 195 165 163 165 107 145, 146, 147 294, 295 245, 246 236 Oral Vir#5 144 109 103 165 Oral Vir#5
Takahashi, N Takamatsu, T Talaei-Hassanloui, R Tan, Sek Yee Tan, Yeping Tanaka, S Tang, Zhimin Tarver, MR Tatahash, D Tatianan, D Taylor, CG Thakuria, D Theilmann, DA Thézé, J Thiéry, D Tokarev, Y Tong, Zongxing Toprak, U Torres-Quintero, MC Trimble, J Tripodi, AD Troemel, ER Tsuruta, K	265 199 263 119 142 46 99, 195 288, 289 237 125 35 Poster Bac26 17, 68, 161, 181, 182 182 56, Oral Fun#3 135 78 77, 162, 186 47 88 63 134, 221 100	Walsh, T Wang, Chung-Hsiung Wang, Dun Wang, Haichuan Wang, Haidong Wang, Hanzhong Wang, Hualin Wang, JB Wang, Jinwen Wang, Kui Wang, Manli Wang, Qianqian Wang, Qianran Wang, Xiujuan Wang, Yueqin Wang, Zengxia Wang, Zeyu Wang, Zhenying Wang, Xi Warnecke, SA Wassermann, M Weavil-Abueg,ME Webb, TK Webster, TC	208 19 86 211 313 163 Oral Vir#1 110 191 Poster Bac25 Oral Vir#1, Oral Vir#5 208 187 Oral Vir#5 119 79 80 122 79,80 115 Oral Vir#5 98 271 114 289 222 90	Xia, Yuxian Xie, Weiping Xing, Longsheng Xing, Yali Xu, Congrui Xu, Jinshan Xu, Lei YY Yalpani, N Yamamoto, T Yanagawa, A Yang, Kai Yang, Ming-Jun Yang, Shili Yang, Yang Yang, Yi-Ting Yao, Qin Yasin, M Yates, JR Ye, Sudan Yin, Mengyi Yokoyama, T Yoshimura, T Yousefelahi, MN Yu, Dong-Shuai Yuan, Chuanfei Yu, Qian	111, 291 126 Oral Vir#5 145 163 23 217 126 48, 49 109 96, 97, 99, 195 165 163 165 107 145, 146, 147 294, 295 245, 246 236 Oral Vir#5 144 109 103 165 Oral Vir#5 145

Yuan, Meijin 96, 97, 99,

195

Yun, HwiGeon 194, 224, 273,

> **274**, Poster Msp06, Poster Vir21

Z	Z	Z	

Zack, M 119 Zaghloul, H 142**, 143** Zahrna, EB Poster Fun24 Zamanian, M Zamudio-Ramirez, A 139 Zeigler, DR 286 Zekeya, N 52 Zeng, Deyu 291 Zhan, Yiling 78 Oral Vir#1 Zhang, Fenghua Fang, Fengjuan 219 Zhang, Gaobo 163 Zhang, Hao 96 Zhang, Jie 122,

Poster Bac24, Poster Bac25

Zhang, Lingling 217 Zhang, Panpan 146 Zhang, Qinfen 187 Zhang, Tiantao 80 Zhang, Wei 111 Zhang, Yahui 147 Zhao, Jin 191 Zhao, Mingli 170 Zheng, Jinshui 287 Zheng, Yi 48 Zhou, Zishan 122 Zhu, Silei 96, **97** Zhu, Yu Cheng 278 Zitlalpopoca-Hernandez, G 234 Zou, Zhen Oral Vir#5

ADDITIONAL, LATE RECEIVED ABSTRACTS FOR POSTER PRESENTATION and OTHER PERTINENT CHANGES IN THE PROGRAM

MONDAY am

SIP Founders' Jubilee Celebration

(Theater)

Micrsporidia research through the ages

Ann Cali

¹Rutgers, The State University of New Jersey, Department of Biological Sciences , 195 University Ave, Newark, NJ 07102, USA Corresponding author: anncali@newark.rutgers.edu

Since the establishment of the SIP, a primary interest has been the discovery of potential biological control agents and pathogens of beneficial insects. The Microsporidia are part of this mission and the related studies of safety, epidemiology, organism survival, and impact data are necessary. An example was the development of the microsporidium Paranosema (Nosema) locustae. It is now a commercially available biological for the control of rangeland grasshoppers. While others have studied microsporidia for control of mosquitoes, lepidopterans, fire ants, beetles, honey bees etc. Microsporidia are obligate intracellular pathogens and an amazing amount of detail has been observed with the tools of the first 100 years. Approximately sixty years ago, Huger published the first electron micrographs (EM) of spores revealing their features. Subsequently, microsporidial EM studies have made huge advancements. Details of lifecycles and the host/parasite interface could be discerned, differences between organisms became apparent and the descriptions of genera were being modified. For example, at our first Microsporidia division meeting in 1970, EM of microsporidial development of three species of Nosema revealed their similarities and differences, which were sufficiently significant to separate them into two genera and Encephalitozoon was made a microsporidial genus. By 1976, Vavra and Sprague published "Biology of the Microsporidia" with EMs illustrating the presence and absence of cell organelles and the presence of new structures. In 1977, their second volume reviewed the known 46 genera. Publications of both morphology and taxonomy followed, but it was molecular techniques that became an integral part of biosystematic studies in the Microsporidia. The complete genome of Encephalitozoon cuniculi was published containing 2.9Mbases, one of the smallest known for a eukaryote. The small subunit rDNA became the standard for organismal comparison. This information lead to massive reclassification of the Microsporidia. They were considered very primitive, and since they lacked mitochondria, Cavalier-Smith (1987) moved them from Protozoa to the kingdom Archezoa. They were reconsidered because of features such as Golgi enzymes and microtubule sequence data to be Kingdom Protista but close to Fungi some consider them to belong in the Fungal kingdom. This is still a debate. Since 1985, several human infections reported especially Enterocytozoon bieneusi. It or similar identified in fish, pigs, and Arthropods.

TUESDAY am

Microbial Control Contributed Papers #1

(Marshall Room)

10:00

Mohammed I. Elbashir, P. Bishwajeet, K. Shankarganesh, P. Sharma. Performance of three Indian isolates of *Beauveria bassiana* (Balsamo) Vuillemin and three commercial mycoinsecticides against three developmental stages of *Bactrocera dorsalis* (Hendel) (Diptera:Tephritidae)

*Paper #280; MOVED from poster MCn22

TUESDAY am

DBI Contributed Papers #1

(Marshall Room)

10:45 STU Nina S. Lottsfeldt, Mariah E. Weavil-Abueg, Lisa M. Crosson,
Carolyn S. Friedman. Candidatus Xenohaliotis californiensis
viability and infectious dose in Haliotis rufescens

THIS IS A JUDGED STUDENT PRESENTATION.

WEDNESDAY pm

POSTERS

(West Ballroom)

FUNGI DIVISION POSTERS

Roles of two photolyases in sustaining UV-B STU Fun26 resistance and photoreactivation of Beauveria bassiana

Ding-Yi Wang, Sheng-Hua Ying, Ming-Guang Feng

Institute of Microbiology, College of Life Sciences, Zhejiang University, Hangzhou, Zhejiang, China Corresponding author: mgfeng@zju.edu.cn

Solar UV irradiation exerts a harmful effect on applied fungal biocontrol agents. Photolyases are enzymes that can repair DNA damage caused by UV irradiation. This study seeks to characterize functions of the CPD photolyase Phr1 and the 6-4 photolyase Phr2 in Beauveria bassiana, a filamentous fungal insect pathogen. Deletion of phr1 resulted in a loss of ~40% UV-B resistance and of >90% photoreactivation in conidia. Deletion of phr2 led to a significantly less loss of either UV-B resistance or photoreactivation. Short blue (450 nm) and violet (480 nm) lights were evidently optimal for the photoreactivation of both Phr1 and Phr2. Interestingly, the Δ phr2 mutant displayed a DNA repair activity increased with the time (0-5 h) of exposure to white light, greater tolerance to the cell wall stressor calcofluor white and increased yields of submerged blastospores and aerial conidia. Overexpression of phr2 in B. bassiana resulted in reversed changes in the aforementioned phenotypes. Conclusively, Phr1 and Phr2 played important, but differential, roles in the fungal response to harmful UV irradiation and hence in sustaining the fungal biocontrol potential against arthropod pests.

Daylight length-dependent translocation of VIVID photoreceptor in cells and its regulatory role in conidiation and virulence of *Beauveria bassiana*



Sen-Miao Tong, Sheng-Hua Ying, Ming-Guang Feng

Institute of Microbiology, College of Life Sciences
Zhejiang University, Hangzhou, China
Corresponding author: mgfeng@zju.edu.cn

The fungal insect pathogen *Beauveria bassiana* has the blue-light photoreceptor VIVID (VVD) but lacks a pigmentation pattern to trace its light responses. Here we show that the fungal VVD is transcriptionally expressed, and linked to other blue/red photoreceptors, in a light-dependent manner. GFP-tagged VVD fusion protein was localized to periphery, cytoplasm and vacuoles of hyphal cells in light/dark (L:D) cycles of 24:0 and 16:8 and aggregated in cytoplasm with shortening daylight until transfer into nuclei in full darkness. Deletion of *VVD* reduced conidiation capacity in L:D 12:12 cycle of 450/480 nm blue light (91%) more than yellow-to-red (540–760 nm) and

white lights (~70%). The conidiation defect worsened with shortened daylight in different L:D cycles of white light, coinciding well with drastic repression of key activator genes in central development pathway. Intriguingly, the deletion mutant displayed blocked secretion of cuticle-degrading Pr1 proteases, retarded dimorphic transition *in vivo* or *in vitro*, and hence a lethal action twice longer than those for control strains against *Galleria mellonella* regardless of the infection passing or bypassing insect cuticle. Conclusively, VVD sustains conidiation level in a daylight length-dependent manner and acts as a vital virulence factor in *B. bassiana*.

Transcriptomic analysis unveils the potential pathways regulated by autophagy-related protein 1 (BbATG1) during conidiation of *Beauveria bassiana*

Fun28

Wei-Xia Dong, Ming-Guang Feng, Sheng-Hua Ying

College of Life Sciences, Zhejiang University, Hangzhou, Zhejiang, 310058, China Corresponding author: yingsh@zju.edu.cn

Conidiation promotes fungal dispersal and survival in the environment, and is a determinant for the biocontrol potential of $Beauveria\ bassiana$. Autophagy-related protein 1 (BbATG1), a serine/threonine protein kinase, plays an important role in autophagic process and development of B. bassiana. To explore the down-stream targets regulated by BbATG1 under menadione stress, a comparative trancriptome was applied on the wild-type and the $\Delta BbATG1$ mutant strains during conidiation. Transcriptomic analysis indicated that the conidiation-related genes regulated by BbATG1 were significantly enriched in the functional catalogs of metabolism, protein fate, cell rescue, and transportation. Conclusively, these results indicate that BbATG1 contributes to conidial development of B. bassiana by mediating the potential pathway of metabolism and protein fate. These findings highlight the potential genetic network involved in fungal conidiation, and provide the new clues for improving condiation potency of entomopathogenic fungi.

Ethanol dehydrogenase I contributes to growth and sporulation under low oxygen condition via detoxification of acetaldehyde in *Metarhizium acridum*

Erhao Zhang^{1,2}, **Yueqing Cao**^{1,2}, Yuxian Xia^{1,2}

Fun29

¹School of Life Sciences, Chongqing University, Chongqing, China; ²Chongqing Engineering Research Center for Fungal Insecticides, Chongqing, Chin *Corresponding author: yuxiaxia@cqu.edu.cn*

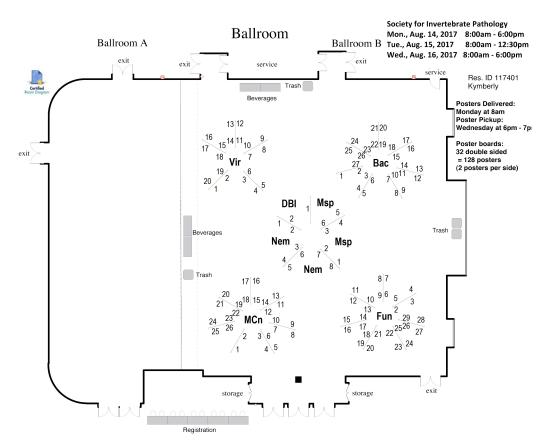
The entomopathogenic fungi encounter hypoxic conditions in nature and in culture. Alcohol dehydrogenases (ADHs) are a group of oxidoreductases that occur in many organisms. Here we demonstrate that an alcohol dehydrogenase I, MaADH1, in the locust-specific fungal pathogen, Metarhizium acridum, functions in acetaldehyde detoxification mechanism under hypoxic conditions in growth and sporulation. The MaADH1 was highly expressed in sporulation and had much higher expression under hypoxic conditions. Compared with a wild-type strain, the *\Delta MaADH1* deletion mutant showed inhibited growth and sporulation under hypoxic conditions, but no impairment under normal conditions. ΔMaADH1 mutant produced significant decreased alcohol, but significant increased acetaldehyde compared to wild type. Though alcohol and acetaldehyde could inhibited growth and sporulation of M. acridum, M. acridum was much more sensitive to exogenous acetaldehyde than alcohol. These results indicate that the MaADH1 plays an important roles in growth and sporulation under hypoxic stress by detoxification of acetaldehyde in M. acridum.

MICROBIAL CONTROL DIVISION POSTERS

MCn22

Mohammed I. Elbashir, P. Bishwajeet, K. Shankarganesh, P. Sharma. Performance of three Indian isolates of *Beauveria bassiana* (Balsamo) Vuillemin and three commercial mycoinsecticides against three developmental stages of *Bactrocera dorsalis* (Hendel) (Diptera:Tephritidae)

Paper #280 in program; rescheduled for oral presentation in Microbial Control #1 (Tuesday am, 10:00; Marshall Room)



V	VEDNESDAY – 16 August			THURSDAY – 17 August
8:00 10:00	Bacteria Division #3 Microsporidia Division	(Theater)	8:00 – 10:00	Bacteria Division Symposium: What is Bt? Current
	Symposium: The pas	t and		perspectives (Theater)
	future frontiers in			Microbial Control Division #4
	Microsporidiology			(the Forum)
	(Retrospective look			Diseases of Beneficial Invertebrates
	Microsporidia resear			Division Symposium:
	from the first meetir	•		The Pathobiobiome Concept:
	ago)	(The Forum)		An emerging view of
	Microbial Control Division			microbes and diseases
	•	oosevelt Room)		(Roosevelt Room)
10:00 – 10:30	Coffee break		10:00 – 10:30	Coffee break
10:30 – 12:30	Virus Division #3	(Theater)	10:30 – 12:30	SOCIETY FOR INVERTEBRATE
	Microbial Control Division			PATHOLOGY business meeting
	•	oosevelt Room)	12:30 – 1:30	Lunch
12:30 – 1:30	Lunch			Awards Committee (Green Room)
	Nematode Division busi		1:30 – 3:30	Fungi/Nematodes/Microsporidia
	•	oosevelt Room) •		X-Divisional Symposium:
	Student Worshop on Sc			Ecology of Invertebrate
	Communication	(The Forum)		Pathogens (Theater)
1:30 – 3:30	Virus Division #3	(Theater)		Microbial Control Division
	Bacteria Division #4	(The Forum)		Symposium: Biopesticides III:
	,	oosevelt Room)		Beyond entomopathogenicity –
	Microsporidia Division #			Reliable tools or just a novelty?
	· ·	Marshall Room)		(The Forum)
3:30 – 4:00	Refreshments break			Virus Division #5 (Roosevelt Room)
4:00 – 6:00	•	West Ballroom)	3:30 – 4:00	Refreshments break
8:00 – 10:00	Fungus Division busines	_	6:30 – 12:30	SIP BANQUET
	(Village West, Bldg. 2, Rooms 2A/2B)			(Estancia La Jolla Hotel)
	Microsporidia Division	ousiness		Cocktail Hour
	meeting/workshop			Banquet
	(Village West, Bl			Awards Ceremony
	Virus Division business r	•		Dancing
	and tribute to Micha			
	Strand's election to t			
	US National Academy	y of Sciences		

(Village Tower West, 15th fl., Rm. 15B)





















The support of these organizations for the 2017 Annual Meeting and Golden Jubilee Celebration of the Society for Invertebrate Pathology is gratefully acknowledged.

















thoughtseeders™

